



Proving that Traditional Knowledge Works: The antibacterial activity of Northern Peruvian medicinal plants

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Research

Abstract

Bacterial infections and inflammation are among the ailments treated by traditional healers. The World Health Organization has expressed high interest in Traditional Medicine (TM), and it is important to demonstrate scientifically that the remedies employed in folk medicine are indeed therapeutically active. In order to evaluate the antibacterial activity of species used in traditional medicine in Northern Peru, 525 plant samples of at least 405 species were tested in simple agar-bioassays for antibacterial activity under simple laboratory conditions in a private clinic in Trujillo, Peru. Antibacterial activity was investigated against *Staphylococcus aureus* Rosenbach 1884, *Escherichia coli* (Migula 1895) Castellani & Chalmers 1919, *Salmonella enterica* Typhi (ex Kauffmann & Edwards 1952) Le Minor & Popoff 1987, and *Pseudomonas aeruginosa* (Schröter 1872) Migula 1900. The aim of the study was to scientifically test whether plants used in TM for the treatment of infections showed antibacterial activity, and to delineate a number of candidates for further in-depth study of their Minimum Inhibitory Concentration (MIC) and toxicity. One-hundred-ninety-three ethanolic extracts and 31 water extracts were active against *S. aureus*. In twenty-one cases only the water extract showed activity. None of the aqueous extracts were active against the other three bacteria, with the activity of the ethanolic extracts also much reduced, as only 36 showed any activity against *E. coli*, and 3 each against *S. enterica* Typhi and *P. aeruginosa*. Two-hundred-twenty-five extracts came from species that are traditionally employed against bacterial infections. One-hundred-sixty-six (73.8%) of these were active against at least one bacterium. Of the three-hundred extracts from plants without traditional antibacterial use, only 96 (32%) showed any activity. Plants used for respiratory disorders, inflammation/infection, wounds, diarrhea, and to prevent post partum infections were efficacious in 70-88% of the tests. Plants used for "kidney inflammation"

had a much lower efficacy against bacteria and fell within the range of species that are traditionally used to treat other bodily disorders.

Resumen

Infecciones bacterianas e inflamación se encuentran entre las enfermedades tratadas por curanderos tradicionales. La Organización Mundial de Salud se ha expresada como altamente interesada en la Medicina Tradicional, y es importante demostrar científicamente que los remedios usados en la medicina popular de veras son terapéuticamente activos. En este trabajo evaluamos la propiedad antibacteriana de 525 muestras de plantas medicinales del Perú septentrional de mínimo 405 especies contra *Staphylococcus aureus* Rosebach 1884 *Escherichia coli* (Migula

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1895) Castellani & Chalmers 1919, *Salmonella enterica* Typhi (ex Kauffmann & Edwards 1952) Le Minor & Popoff 1987 e *Pseudomonas aeruginosa* (Schröter 1872) Migula 1900, **usando un método de difusión en agar bajo de condiciones simples de laboratorio en Trujillo, Perú. La meta de este estudio fue de científicamente probar si plantas usadas en la Medicina Tradicional para tratar infecciones mostraron actividad antibacteriana, y para delinear candidatos para estudios futuros de Concentración Inhibitoria Mínima y toxicidad. Ciento noventa y tres extractos etanolicos y 31 extractos en agua mostraron actividad en contra de *S. aureus*. En 21 casos solo los extractos acuosos fueron eficaces. Ninguno de los extractos acuosos tuvo actividad contra las otras bacterias, y solo 36 de los etanolicos mostraron eficaz contra *E. coli*, y 3 en cada caso contra *S. enterica* Typhi e *P. aeruginosa*. Doscientos veinticinco extractos pertenecieron a especies tradicionalmente usadas como antibacterianas. De estos 73.8% fueron activos. De los 300 extractos de plantas no tradicionalmente usadas contra bacterias, solo 32% tuvieron un efecto positivo. Plantas usadas para el tratamiento de infecciones respiratorias, inflamación/infección, heridas, diarrea e infecciones después del parto fueron eficaces en 70-88% de los casos. Plantas usadas para inflamaciones de los riñones y otros desordenes tuvieron una eficaz mucho mas baja en contra de bacterias.**

Introduction

In developing countries, Traditional Medicine (TM) is often the only accessible and affordable treatment available. In Latin America, the World Health Organization (WHO) Regional Office for the Americas (AMRO/PAHO) reports that 71% of the population in Chile and 40% of the population in Colombia has used TM. In many Asian countries TM is widely used, even though Western Medicine is often readily available. In Japan, 60–70% of allopathic doctors prescribe traditional medicines for their patients. In the US the number of visits to providers of Complementary Alternative Medicine (CAM) now exceeds by far the number of visits to all primary care physicians (WHO 1999a, b, 2002).

CAM is becoming increasingly popular in many developed countries. Forty-two percent of the population of the U.S.A. have used CAM at least once (WHO 1998), and a national survey reported the use of at least one of 16 alternative therapies increased from 34% in 1990 to 42% in 1997 (UNCTD 2000).

The expense for the use of TM and CAM is exponentially growing in many parts of the world. The 1997 out-of-pocket CAM expenditure was estimated at \$ 2.7 billion in the U.S.A. The world market for herbal medicines based on traditional knowledge was estimated at US\$60 billion

already over a decade ago (Breevort 1998). It is however important to demonstrate scientifically that the remedies employed in folk medicine are indeed therapeutically active (Baker *et al.* 1995, Cox & Balick 1994, Elisabetsky & Castilhos 1990, Farnsworth *et al.* 1985, Muñoz & Sauvain 2002, Schultes 1994).

Peru is a country rich in biodiversity. For millennia, traditional healers have used the flora to treat ailments. The same plants are still being used today. Traditional Medicine continues to be very popular since a large part of the population has either no access to, or cannot afford, Western Medicine. Bacterial infections and inflammation are among the ailments treated by traditional healers. Northern Peru is believed to be the center of the Central Andean Health Axis (Camino 1992, 1999), and traditional medicinal practices in this region remain an important component of everyday life (Bussmann 2006, Bussmann & Sharon 2006, De Feo 1992, Joralemon & Sharon 1993, Polia 1988, Sharon 1978, 1980, 1994, 2000, Sharon & Bussmann 2006). TM is also gaining respect by national governments and health providers. Peru's National Program in Complementary Medicine and the Pan American Health Organization recently compared Complementary Medicine to allopathic medicine in clinics and hospitals operating within the Peruvian Social Security System (Es-Salud 2000). According to the WHO (2002), the sustainable cultivation and harvesting of medicinal plant species is one of the most important challenges in the next few years. The WHO has expressed high interest in TM.

Plants with potential medicinal activity have recently come to the attention of Western scientists, and studies have reported that some are bioactive (e.g., Perumal Samy & Ignacimuthu 2000). Potentially active compounds have been isolated from a few of the plants tested (D'Agostino *et al.* 1995 a, b, Okuyama *et al.* 1994, Rodriguez *et al.* 1994, Umana & Castro 1990).

Plant species from the Cordillera Blanca, one of the high-altitude areas of Peru, have been studied in recent years for their antimicrobial, anti-cancer, and wound-healing activities (Bussmann *et al.* 2008, Hammond *et al.* 1998, Lee *et al.* 1999, Neto *et al.* 2002, Villegas *et al.* 1997). However, despite the fact that the center of healing traditions in Northern Peru is located in the Trujillo / Chiclayo coastal region, no studies had been undertaken in this area until the work of the Minority Health Disparity International Research Training (MHIRT) Peru.

In this communication we report on antibacterial assays for 391 plant species with a wide range of traditional uses, conducted under simple laboratory conditions in a private clinic in Trujillo. The goal of this investigation was to validate the medicinal use of these plants. We hypothesize that plants that are traditionally used to treat illnesses caused by bacteria have a higher likelihood to being efficacious than plants that are not used traditionally as anti-

bacterials, in particular plants that are normally employed for purely spiritual healing purposes. However, we hypothesize that plants that are employed as baths for spiritual cleansing, and thus come in direct contact with the skin of the patient, might have measurable antibacterial effects, while plants that are used in protective amulets (**seguros**) are not active.

Materials and Methods

Plant Material

Plants in Peru were collected in the field, in markets, and at the homes of traditional healers (**curanderos**) in Northern Peru (Figure 1) in August-September 2001, July-August 2002, July-August 2003, June-August 2004, July-August 2005, July-August 2006, June-August 2007, June-August 2008, March-April 2009 and June-August 2009. A total of 116 informants (6 healers and 110 market vendors, of which 20 also acted as healers) in the Trujillo and Chiclayo area were interviewed using structured questionnaires. The informants were always provided with fresh plant material, either collected with them, by them, or available at their market stands. The questionnaires did not include any leading reference as to disease concepts. The participants were only asked simple questions along the lines "What is this plant used for, which part, which quantity, how is it prepared, are any other plants added to the mixture." All questions were asked in the same order. All informants were of Mestizo origin, and spoke only Spanish as their native language. The study covered the four existing medicinal plant markets of the region, and included all vendors present. All interviews were conducted with the same set of participants. The allopathic disease concept of "infection" was found to cover not only topical bacterial, viral and fungal diseases, but also external and internal inflammatory processes, when looked at from the local healing perspective. For

this reason, all plants used to treat such symptoms were included in the study. The specimens are registered under the collection series "RBU/PL," "ISA," "GER," "JULS," "EHCHL," "VFCHL," "TRUBH," and "TRUVANERICA," depending on the year of fieldwork and collection location. Surveys were conducted in Spanish by fluent speakers. Surveyors would approach healers, collectors and market vendors and explain the premise for the study, including the goal of conservation of medicinal plants in the area.

Vouchers of all specimens were deposited at the Herbario Truxillensis (HUT, Universidad Nacional de Trujillo), and Herbario Antenor Orrego (HAO, Universidad Privada Antenor Orrego Trujillo). In order to recognize Peru's rights under the Convention on Biological Diversity, most notably with regard to the conservation of genetic resources in the framework of a study treating medicinal plants, the identification of the plant material was conducted entirely in Peru. Plant material was identified by the authors, using available floras as indicated in the nomenclature section, as well as herbarium vouchers in the herbaria where material was deposited. No plant material was exported in any form whatsoever.

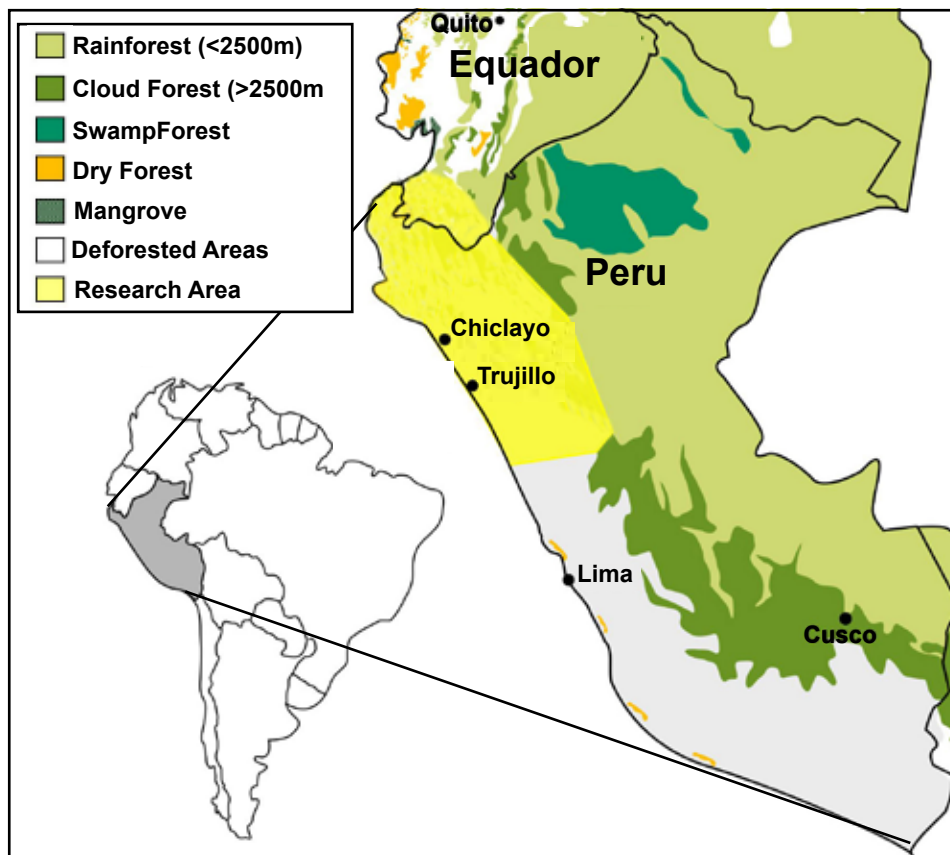


Figure 1. Trujillo and Chiclayo research area of northern Peru.

Nomenclature

The nomenclature of genera and species follows the Catalogue of the Flowering Plants and Gymnosperms of Peru (Brako Zarucchi 1993) and the Catalogue of the Vascular Plants of Ecuador (Jørgensen & León-Yanez 1999). Species were identified using the available volumes of the Flora of Peru (McBride 1936-1981), as well as Jørgensen and Ulloa Ulloa (1994), Pestalozzi (1998), and Ulloa Ulloa and Jørgensen (1993), and the available volumes of the Flora of Ecuador (Sparre & Harling 1978-2009), and reference material in the herbaria HUT, HAO, QCA, LOJA and QCNE. The plant family nomenclature was updated using the TROPICOS database (Tropicos 2010) which follows the Angiosperm Phylogeny Working Group III system of classification.

Disease concepts

In the context of our study, healers and market vendors used the terms “inflammation” and “infection” interchangeably for any indication that, to their understanding, was caused by a bacterial infection. We maintain this concept by simply using “inflammation” as a term that signifies potential antibacterial activity. In addition, we hypothesize that plants employed by healers for their concepts of “bronchitis, pneumonia, cough, wounds, diarrhea, kidney and urinary tract inflammation, blood purification and postpartum infections” also to be antibacterial, because in all these cases a bacterial infection could be suspected.

On the contrary, plants used to treat illnesses described as “diabetes, high blood pressure, arthritis, asthma, intestinal discomfort, or used as antihelminthics, anti-parasitics, menstrual regulation, abortion, contraceptive” etc. were clearly seen by the healers involved as non-antibacterial. We maintained this concept for our study.

A large part of the Peruvian pharmacopoeia (over 40%, Busmann & Sharon 2006) is employed for “spiritual” healing. In such cases patients are either subjected to a healing bath, thus an herbal concoction coming in direct contact with the skin of the patient, or receive a **seguro**, a flask or bottle containing herbs in alcohol. The latter however is not ingested or allowed to come in direct contact with the patient in any way, but rather serves basically as an amulet. Because of their high abundance, “spiritually” used plants provide an ideal comparison group to antibacterial plants. In addition, cleansing baths are often employed when the healer suspects that sorcery has caused an illness, often manifesting as visible inflammation. For this reason, we hypothesized that plants of this kind are in fact antibacterial, although the healers would not mention them when addressing the western effect concept.

Preparation of Extracts

For each species tested, above ground material (in case of trees: leaves or bark as indicated by the collaborating healers) was collected during the same time period to avoid any seasonal variations, and the entire material was used for extract preparation. This corroborates with the traditional preparation (Busmann & Sharon 2006). Plant material was dried at 35°C for three days. After drying, the material was ground with an industrial grinder, and 2 samples of 5g of plant material each were weighed out. Traditionally most plants are prepared in water extracts. In order to investigate if this method yielded the best antibacterial results, or if an ethanol extraction would prove more effective against bacteria, one sample was submerged in 100ml of 96% ethanol and left to macerate for 7 days, while another sample was submerged in 100ml of boiling distilled water and left to macerate for 24h. After maceration the plant material was filtered using standard 10cm filters and 100ml 96% ethanol was added to the water extracts to allow faster solvent removal. The solvent was then evaporated to complete dryness using a standard Buchi rotary-evaporator. The resulting dry extracts were re-suspended in 5ml distilled water. In order to determine the real concentration of each extract, 1ml of previous homogenization of the respective extracts was removed and again completely oven-dried and then weighed to determine the exact amount of extract per ml of final solution.

Antimicrobial assays

Bacteria

Staphylococcus aureus Rosenbach 1884 ATCC 25923, *Escherichia coli* (Migula 1895) Castellani & Chalmers 1919 ATCC 25922, *Salmonella enterica* (ex Kauffmann & Edwards 1952) Le Minor & Popoff 1987 serogroup Typhi (from a patient sample), and *Pseudomonas aeruginosa* (Schröter 1872) Migula 1900 (from a patient sample) were used for the current study.

Bioassays

The antibacterial activity of the crude plant extracts was determined using an agar-diffusion method (Koneman *et al.* 1997). Bacterial strains were obtained as listed above. Bacterial cultures were grown on 5% sheep red blood agar (SBA) and then inoculated onto Mueller-Hinton Agar (PML) for testing.

Following the initial incubation, bacteria were suspended in 10ml of distilled water and their concentration equilibrated to a 0.5 McFarland standard. Using a sterile cotton swab, each sample was transferred onto Mueller-Hinton Agar. Six millimeter blank paper disks were then saturated with 10µl of each plant extract, dried, and applied to the agar surface. Disks with Doxycycline (for *S. aureus* assays, 3µg/ml, 17mm inhibition), Ampicillin (for *E. coli*

assays 1µg/ml, 15mm inhibition), Ceftriaxone (for *S. enterica* Typhi assays, 3µg/ml, 8mm inhibition) and Ciprofloxacin (for *P. aeruginosa* assay, 0.5µg/ml, 10mm inhibition) were applied as controls. These respective antibiotics were chosen because they are often employed as first line antibiotics in the respective bacterial infections. Disks with distilled water were used as negative controls. Plates were incubated at 37°C. After 24h, zones of inhibition appearing around disks were measured and recorded in millimeters. At least three replicates were run for each assay. The antibiotic controls showed an inhibition zone of at least 6mm after 24h exposure in all assays. Extracts from plant species were considered anti-bacterially active if the inhibition zone was least 6mm.

Results

In order to evaluate the antibacterial activity of species used in TM in Northern Peru, 525 plant samples of at least 405 species were tested in simple agar-bioassays for antibacterial activity against *S. aureus*, *E. coli*, *S. enterica* Typhi and *P. aeruginosa*. Table 1 shows the results of the antibacterial assays. A much larger number of ethanolic plant extracts showed any antibacterial activity compared to water extracts for all antibacterial activity. One-hundred-ninety-three ethanolic extracts and 31 water extracts were active against *S. aureus*. In twenty-one cases only the water extract showed activity (for all bacterial species) compared to ethanol only. None of the aqueous extracts were active against the other three bacteria, with the activity of the ethanolic extracts also much reduced, as only 36 showed any activity against *E. coli*, and 3 each against

S. enterica Typhi and *P. aeruginosa*. Eighteen ethanol extracts were effective against both *E. coli* and *S. aureus*, while in two cases the ethanol extract showed activity against *E. coli* and the water extract against *S. aureus*. The ethanol extract of *Dioscorea trifida* L.f. was effective against *E. coli*, *S. aureus* and *P. aeruginosa*. *Caesalpinia spinosa* (Molina) Kuntze was the only species that showed high activity against all bacteria, including *S. enterica* Typhi and *P. aeruginosa*, when extracted in ethanol. This is of particular interest for potential further research.

Two-hundred-twenty-five extracts came from plant species that are traditionally employed against bacterial infections. One-hundred sixty-six (73.8%) of these were active against at least one bacterium. Of the three-hundred extracts from plants without traditional antibacterial use, only 96 (32%) showed any activity (Figure 2). This shows clearly that plants traditionally used as antibacterial had a much higher likelihood to be antibacterially active than plants without traditional anti-bacterial use. However, the efficacy of plants used traditionally for antibacterial related applications did vary, which underlines the need for studies aiming to clearly understand traditional disease concepts. Plants used for respiratory disorders, inflammation/infection, wounds, diarrhea, and to prevent postpartum infections were efficacious, based on antibacterial activity, in 70-88% of the tests. Plants used for "kidney inflammation" had a much lower efficacy against bacteria, and fell within the range of species that are traditionally used to treat other bodily disorders. Only species used for spiritual/ritual treatments scored worse. Of these only 22% showed some antibacterial activity (Figures 3, 4). However, amongst the "spiritual" plants 38% of the species used

for cleansing baths did in fact show activity, while only 15% of the plants often used in protective amulets (mostly species with the families of Lycopodiaceae and Caprifoliaceae) showed limited antibacterial activity.

A variety of species showed higher efficacy than the control antibiotics employed: *Ambrosia peruviana* Willd., *Iresine herbstii* Hook., *Niphogeton dissecta* (Benth.) J.F. Macbr., *Opuntia ficus-indica* (L.) Mill., *Smilax kunthii* Killip & C.V. Morton were particular effective against *E. coli*. *Berberis buceronis* J.F. Macbr., *Caesalpinia paipai* Ruiz & Pav., *C. spinosa*, *Cestrum strigilatum* Ruiz & Pav., *Cydista aequinoctialis* (L.) Miers, *D. trifida*, *Escallonia pendula* (Ruiz & Pav.) Pers., *Escobedia grandiflora* (L.f.) Kuntze, *Eucalyptus citriodora* Hook., *Eucalyptus globulus* Labill., *Eugenia obtusifolia* Cambess., *Eustephia coccinea* Cav., *Gallesia integrifolia* (Spreng.) Harms,

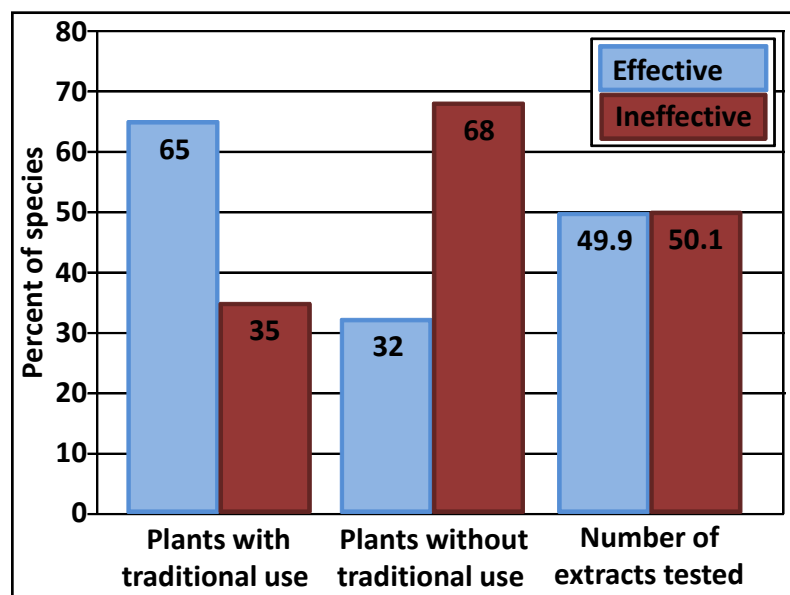


Figure 2. Evaluation of 225 plant species extracts that are traditionally employed against bacterial infections in the area of Trujillo and Chiclayo, northern Peru.

Table 1. Antibacterial activity of 525 plant samples from 405 species used in traditional medicine in Northern Peru. Each were tested in simple agar-bioassays for antibacterial activity against *Staphylococcus aureus* Rosenbach 1884, *Escherichia coli* (Migula 1895) Castellani & Chalmers 1919, *Salmonella enterica* (ex Kauffmann & Edwards 1952) Le Minor & Popoff 1987 serogroup Typhi and *Pseudomonas aeruginosa* (Schröter 1872) Migula 1900. Ethanol extracts noted in black, Water extracts noted in red. The positive control antibiotics led to bacterial inhibition in all assays. For details see Materials and Methods. The negative control (a disk with distilled water) did not cause any antibacterial effect in any assay. Antibacterial species are noted as:

Plants with efficacy AND traditional antibacterial use.

Plants with traditional antibacterial use BUT WITHOUT efficacy.

Plants with efficacy, but NOT traditionally used as antibacterial.

Traditional applications: Abortifacient (Ab); Asthma (As); Arthritis (Ar); Blood pressure (Bp); Blood purification (BI); Bronchitis (Br); Contraceptive (C); Diabetes (Db); Diarrhea (Dr); Infection (If); Inflammation (In); Intestinal (It); Kidney Inflammation (K); Nerves (N); Parasites (Pa); Postpartum infection (Pi); Respiration (R); Spiritual cleansing (S); Wounds (W).

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
Pteridaceae								
<i>Adiantum concinnum</i> Humb. & Bonpl. ex Willd.	Culantrillo	ACR91		8				BI
Amaranthaceae								
<i>Alternanthera halimifolia</i> (Lam.) Standl. ex Pittier	Sanguinara	KMM468	10					In
<i>Alternanthera porrigens</i> (Jacq.) Kuntze	Moradilla	AKT149/ KMM395						Blood circulation
<i>Alternanthera villosa</i> Kunth	Hierba de Oso	ACR 103						S
<i>Amaranthus hybridus</i> L.	Yuyo	GER190		11				In
<i>Chenopodium ambrosioides</i> L.	Paico	ACR31						Anthelmintic
<i>Gomphrena globosa</i> L.	Siempre viva	ACR101						S
<i>Iresine herbstii</i> Hook.	Color	ACR162	17					In
<i>Iresine herbstii</i> Hook.	Sanguinaria	JULS75	10	16				In
Amaryllidaceae								
<i>Allium sativum</i> L.	Ajo	ACR107						As
<i>Eustephia coccinea</i> Cav.	Pumapara	RBU/PL313		7				In
<i>Eustephia coccinea</i> Cav.	Para Para	ACR138		13	20			In
	Fosforito	KMM542						S
	Pumapara	ACR119						In
Anacardiaceae								
<i>Mauria heterophylla</i> Kunth	Chacur	ACR77/ JULS17	14	22				In
<i>Schinus molle</i> L.	Molle	KMM404						In
<i>Schinus molle</i> L.	Molle	JULS196		14				In

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
Annonaceae								
<i>Annona muricata</i> L.	Guanabana	ACR81		11				In
Apiaceae								
<i>Ammi visnaga</i> (L.) Lam. cf.	Bisnaga	KMM566						S
<i>Apium graveolens</i> L.	Apio del campo	KMM439		11				In
<i>Arracacia xanthorrhiza</i> Bancr.	Racacha de zorro	KMM466						S
<i>Coriandrum sativum</i> L.	Cilantro	KMM548		12				Dr
<i>Foeniculum vulgare</i> Mill.	Hinojo	ACR82		10				In
<i>Foeniculum vulgare</i> Mill.	Inojo	KMM409						In
<i>Niphogeton dissecta</i> (Benth.) J.F. Macbr.	Hornamo Toro	AKT1196	16	10				W
<i>Petroselinum crispum</i> (Mill.) Fuss	Perejil	JULS225		15				If
Apocynaceae								
<i>Mandevilla antennacea</i> (A.DC.) K. Schum.	Bejuco Colambo	GER236		8				S
<i>Mandevilla cf. trianae</i> Woodson	Bejuco Negro	ISA14	13	24				S
<i>Mandevilla cf. trianae</i> Woodson	Bejuco negro	AKT1221						S
<i>Nerium oleander</i> L.	Laurel	ACR34		13	11			W
<i>Thevetia peruviana</i> (Pers.) K. Schum.	Machil	KMM416						Ar
Aquifoliaceae								
<i>Ilex guayusa</i> Loes.	Gauyusa	KMM513		14				In
Araceae								
<i>Anthurium</i> sp.	Patecina negra	KMM486						S
Araliaceae								
<i>Hydrocotyle globiflora</i> Ruiz. & Pav.	Sombrero	KMM467						K
<i>Oreopanax eriocephalus</i> Harms	Maqui Maqui	KMM445						If
<i>Oreopanax eriocephalus</i> Harms	Maqui Maqui	JULS39		8				If
Aristolochiaceae								
<i>Aristolochia cf. ruiziana</i> (Klotzsch) Duch.	Bejuco Colambo / Bejuco del Aire	AKT1217/ KMM563						S
Asteraceae								
<i>Acanthoxanthium spinosum</i> (L.) Fourr.	Juan Alonzo	AKT1127						In

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Achillea millefolium</i> L.	Milenrama	AKT1188/ ACR139						Gastritis
<i>Achyrocline alata</i> (Kunth) DC.	Hierba de Ishpingo	AKT1199		10				Ar
<i>Achyrocline</i> sp.	Espina de Hoja	KMM547						
<i>Ambrosia peruviana</i> Willd.	Marco o Altamisa	ACR11	17	12				Pi
<i>Arctium lappa</i> L.	Lampazo	GER227		15				Urinary If
<i>Artemisia absinthium</i> L.	Ajenjo	KMM432/ AKT1099						Menstrual regulation
<i>Baccharis genistelloides</i> (Lam.) Pers.	Karqueja	AKT1144						If
<i>Baccharis glutinosa</i> Pers.	Pega Pega	KMM556						Db
<i>Baccharis latifolia</i> (Ruiz & Pav.) Pers.	Chilca Grande	ISA115		12				Ar
<i>Baccharis salicifolia</i> (Ruiz. & Pav.) Pers.	Cedron	AKT1126						S
<i>Baccharis salicifolia</i> (Ruiz. & Pav.) Pers.	Chilco Hembra	GER125		10				S
<i>Baccharis</i> sp	Chilca chica	KMM562						Ar
<i>Baccharis</i> sp.	Chilca	KMM498			9			Ar
<i>Baccharis</i> cf. <i>vaccinioides</i> Kunth	Sigueme Sigueme	KMM565						S
<i>Bidens pilosa</i> L.	Amor seco	KMM427						K
<i>Chersodoma deltoidea</i> M.O. Dillon & Sagást.	Arquitecta	ACR46/ KMM446						S
<i>Chromolaena</i> sp.	Asma chilca	ACR113						As
<i>Chromolaena</i> sp	Asma chilca	KMM555						As
<i>Chuquiraga spinosa</i> Less. ssp. <i>humanpita</i> C. Ezcurra	Huaman pinta	KMM434						In
<i>Chuquiraga weberbaueri</i> Tovar	Amaro	KMM551/ ACR85			10			Br
<i>Cynara cardunculus</i> L.	Alcachofa	KMM414		14				Bl
<i>Diplostephium sagasteguii</i> Cuatrec.	Gato Simuro	ACR124,153/ AKT1145, 1192/KMM478		13				S
<i>Eupatorium</i> cf. <i>gayanum</i> Wedd.	Asma chilca	KMM555						As
<i>Flaveria bidentis</i> (L.) Kuntze	Mata gusanos	KMM412		9				Br

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Gnaphalium americanum</i> Mill.	Lechugilla	ACR41/ KMM393		9				Db
<i>Loricaria cf. ferruginea</i> (Ruiz & Pav.) Wedd.	Pata de Gallina	ACR63/ AKT1174		15				S
<i>Loricaria pauciflora</i> Cuatrec.	Palma	KMM473	9					S
<i>Loricaria</i> sp.	Palma Bendita	ACR182		8				S
<i>Matricaria frigidum</i> (Kunth) Kunth.	Lavanda (la banda)	AKT1175						In
<i>Matricaria recutita</i> L.	Manzanilla	ACR6						W
<i>Munnozia lyrata</i> (A. Gray) H. Rob. & Brettell	Caniahuanga	KMM519						S
<i>Munnozia</i> sp.	Salvia blanco	ACR148						S
<i>Onoseris odorata</i> (D. Don) Hook. & Arn.	Hierba Aguila	ACR150		7				N
<i>Oritrophium peruvianum</i> (Lam.) Cuatrec.	Huamanripa	JULS58		14				Pneumonia
<i>Perezia multiflora</i> (Bonpl.) Less.	Escorsonera	KMM535/ AKT1153						As
<i>Picrosia cf. longifolia</i> D. Don.	Achicoria	KMM436,540/ ACR29, 39						Bl
<i>Porophyllum ruderale</i> (Jacq.) Cass.	Hierba de Gallinazo	KMM515		23				S
<i>Pseudogynoxys cordifolia</i> (Cass.) Cabrera	Hierba San Juan	AKT1168		8				S
<i>Schkuhria pinnata</i> (Lam.) Kuntze ex Thell.	Encanchallacha	ACR17	12					Urinary If
<i>Senecio canescens</i> (Humb. & Bonpl.) Cuatrec.	Vira Vira	ACR44		8				Br
<i>Senecio cf. tephrosioides</i> Turcz.	Huamanripa	ACR65						As, Br
<i>Senecio chionogeton</i> Wedd.	Hornamo Leon Amarillo	GER60	13	18				In
<i>Senecio cf. hypsiandinus</i> Cuatrec.	Ornama blanco	KMM523						S
<i>Senecio</i> sp.	Huamanripa	KMM449		10				As, Br
<i>Senecio</i> sp.	Ornamo	KMM480						S
<i>Senecio</i> sp.	Ornamo tigre	AKT1158						S
<i>Smallanthus sonchifolius</i> (Poepp. & Endl.) H. Rob.	Yacon	ACR197						Db, Prostate

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Sonchus oleraceus</i> L.	Serraja	KMM438						High blood pressure
<i>Stevia</i> sp.	Pastomiel	KMM588						Cholesterol
<i>Tagetes elliptica</i> Sm.	Culantrillo Serrano	GER184		13				Br
<i>Tagetes erecta</i> L.	Flor de Muerto	JULS156		20				In
<i>Tagetes filifolia</i> Lag.	Anis	KMM524						Dr
<i>Tanacetum parthenium</i> (L.) Sch. Bip.	Manzanilla de campo	ACR14						S
<i>Taraxacum officinale</i> F.H. Wigg.	Amargon	JULS150		20				In
<i>Tessaria integrifolia</i> Ruiz. & Pav.	Pajaro bobo	KMM465						In
<i>Trixis</i> cf. <i>cacalioides</i> (Kunth) D. Don	Añasquero	KMM497						S
<i>Verbesina</i> sp.	Sabadilla	ACR154		13				W
<i>Werneria nubigena</i> Kunth	Hierba de halago	ACR205						S
<i>Werneria</i> sp.		AKT1178						S
	Churguis	KMM405						S
	Hierba de Amor	KMM522						S
	Miscichilca	KMM490						S
	Olvido	KMM538						S
Balanophoraceae								
<i>Corynaea crassa</i> Hook. f.	Huanarpo	ACR130						Fertility
<i>Corynaea crassa</i> Hook. f.	Huanarpo	AKT1169		11				Fertility
Berberidaceae								
<i>Berberis buceronis</i> J.F. Macbr.	Palo Amarillo	KMM573	8	21				Hepatitis
Betulaceae								
<i>Alnus acuminata</i> Kunth	Aliso	KMM418			10			W
<i>Alnus acuminata</i> Kunth	Aliso	ACR75		14				W
Bignoniaceae								
<i>Crescentia cujete</i> L.	Turuma	AKT1157	9					Healing of belly button
<i>Cydista aequinoctialis</i> (L.) Miers	Bejuco Amarillo	ISA6	7	21				In from sorcery
<i>Jacaranda acutifolia</i> Humb. & Bonpl.	Arabisco	ACR89	7	16				Br

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
Bixaceae								
<i>Bixa orellana</i> L.	Achiote	KMM429		11				K
Boraginaceae								
<i>Borago officinalis</i> L.	Borraja	ACR9						Bl
<i>Cordia alliodora</i> (Ruiz. & Pav.) Oken.	Ajo Sacha	KMM489		13				Br
<i>Cordia lutea</i> Lam.	Obero	AKT1114						K
<i>Heliotropium curassavicum</i> L.	Alacran	ACR57						S
<i>Tiquilia paronychioides</i> (Phil.) A.T. Richardson	Flor de arena	KMM406		12				In
Brassicaceae								
<i>Brassica oleracea</i> L.	Col	JULS147		11				Gallstones
<i>Capsella bursa-pastoris</i> (L.) Medik.	Bolsa pastor	KMM451						K
<i>Rorippa nasturtium-aquaticum</i> (L.) Hayek	Berros	ACR94/ AKT1163		7				Br
Bromeliaceae								
<i>Puya hamata</i> L.B. Sm.	Hierba del carnero	KMM520						S
<i>Tillandsia cf. cacticola</i> L.B. Sm.	Siempre viva	ACR183						S
<i>Tillandsia</i> sp.	Brocamedia/ Brocamelia	KMM571						S
Burseraceae								
<i>Bursera graveolens</i> (Kunth) Triana & Planch.	Palo Santo	ACR132						Br
<i>Commiphora</i> sp.	Mirra	KMM579						S
Cactaceae								
<i>Echinopsis pachanoi</i> (Britton & Rose) H. Friedrich & G.D. Rowley	San Pedro	ACR95						W caused by witchcraft
<i>Opuntia ficus-indica</i> (L.) Mill.	Tuna	AKT1220	21					Db
	Orillo de Brujo	ACR93						S
Calceolariaceae								
<i>Calceolaria percaespitosa</i> Wooden	Canillahuanga	AKT1195						S
Calophyllaceae								
<i>Mammea americana</i> L.	Mamey	KMM568						Dr
Campanulaceae								
<i>Centropogon articulatus</i> Drake	Conchalalay	EHCHL119		6				S

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			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Centropogon</i> sp.	Canchalagua	KMM536/ ACR17						S
<i>Centropogon</i> sp.	Canchalaly	ACR147						S
<i>Centropogon</i> sp.	Trinoso	KMM545	16	16				Skin problems
<i>Lobelia decurrens</i> Cav.	Contoya	KMM443/ ACR157						S
Capparidaceae								
<i>Capparis crotonoides</i> Kunth	Simura	KMM586						Ar
<i>Capparis scabrida</i> Kunth	Zapote	KMM554						In
Caprifoliaceae								
cf. <i>Belonanthus</i>	Baton de Oro	ACR127/ KMM494						S
<i>Dipsacus fullonum</i> L.	Cardo Santo	EHCHL90		12				Db
<i>Lonicera</i> cf. <i>japonica</i> Thunb. ex Murray	Madre de Selva	KMM390						Depression
<i>Lonicera japonica</i> Thunb. ex Murray	Madre Selva	JULS28		9				Depression
<i>Phyllactis rigida</i> (Ruiz & Pav.) Pers.	Estrella	KMM481		15				S
<i>Phyllactis rigida</i> (Ruiz & Pav.) Pers.	Valeriana estrella	AKT1117						S
<i>Sambucus peruviana</i> Kunth	Sauco	ACR66/ AKT1103						Br, Yellow Fever
<i>Sambucus peruviana</i> Kunth	Sauco	KMM539		6				Br, Yellow Fever
<i>Scabiosa atropurpurea</i> L.	Ambarilla	ACR158		10				Br
<i>Valeriana</i> cf. <i>bonplandiana</i> Wedd.	Fortuna	ACR181						S
cf. <i>Valeriana plantaginea</i> Kunth	Ornamo caballero	ACR120		9				S
<i>Valeriana</i> sp.	Hornamo del Caballo	AKT1140						S
<i>Valeriana</i> sp.	Ornamo	AKT1141			11			S
<i>Valeriana</i> sp.	Valeriana	AKT1213		10				S
<i>Valeriana</i> sp.	Valeriana Delgada	KMM394		12				S
<i>Valeriana</i> sp.	Valeriana gruesa	KMM396		13	13			S

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
Caryophyllaceae								
<i>Dianthus caryophyllus</i> L.	Claveles	AKT1125						Depression
Chloranthaceae								
<i>Hedyosmum racemosum</i> (Ruiz. & Pav.) G. Don.	Asancsito	KMM505		18				Br
Clethraceae								
<i>Clethra castaneifolia</i> Meisn.	Hierba del olvido	KMM549		11				S
<i>Clethra castaneifolia</i> Meisn.	Olvido	ACR109	8					S
Clusiaceae								
<i>Clusia minor</i> L.	Chusgon	JULS280		16				N
<i>Hypericum laricifolium</i> Juss.	Chinchango	AKT1912/ KMM533						S
<i>Hypericum laricifolium</i> Juss.	Pachuli	AKT1172		12				S
<i>Hypericum silenoides</i> Juss.	Cintaura	AKT1154/ KMM387/ ACR152		13				Dr
Convolvulaceae								
<i>Cuscuta foetida</i> Kunth	Pelo de duende	ACR98						Goiter
<i>Ipomoea pauciflora</i> M. Martens & Galeotti	Huanarpo	GER222		16				Cold
Crassulaceae								
<i>Echeveria peruviana</i> Meyen	Pin Pin	ACR169/ AKT1165						K
Cucurbitaceae								
<i>Sicana odorifera</i> (Vell.) Naudin	Cecana	ACR96		11				S
Cupressaceae								
<i>Cupressus sempervirens</i> L.	Pino silvestre	AKT1181/ KMM506						Hemorrhage
Cyperaceae								
<i>Oreobolus goeppingeri</i> Suess.	Hierba del Carpintero	KMM493/ ACR126		12				S
<i>Scirpus californicus</i> Steud. ssp. <i>tatora</i> (Kunth) T. Koyama	Tatora	ACR204						Fever
Dioscoreaceae								
<i>Dioscorea</i> sp.	Papa de Buenas Tardes	KMM462						K

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			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Dioscorea tambillensis</i> R. Kunth	Papa semintona	KMM583						K
<i>Dioscorea tambillensis</i> R. Kunth	Papa semitona	KMM583		12				K
<i>Dioscorea trifida</i> L.f.	Papa madre	KMM503	11	30			11	W, Cancer
Ephedraceae								
<i>Ephedra americana</i> Humb. & Bonpl. ex Willd.	Diego Lopez	AKT1159						Fractures
<i>Ephedra americana</i> Humb. & Bonpl. ex Willd.	Diego Lopez	KMM511	8	22				Fractures
Equisetaceae								
<i>Equisetum bogotense</i> Kunth	Cola de caballo	ACR1		9				W
Ericaceae								
<i>Bejaria aestuans</i> Mutis ex L.	Hierba de la postema	KMM527		12				In
<i>Bejaria aestuans</i> Mutis ex L.	Purunrosa/ FlorPostema	AKT1109						In
<i>Gaultheria erecta</i> Vent.	Mullaca	KMM472/ JULS288		13	11			Br
<i>Gaultheria reticulata</i> Kunth	Maike	KMM531		9				Ar
Escalloniaceae								
<i>Escallonia pendula</i> (Ruiz & Pav.) Pers.	Chuque	ISA23		18				Ar
Euphorbiaceae								
<i>Acalypha mandonii</i> Müll. Arg.	Chilca Dulce	RBU/PL294		11				Liver In
<i>Hura crepitans</i> L.	Habilla	AKT1225						S
<i>Jatropha macrantha</i> Müll. Arg.	Piñones	AKT1230/ KMM487						S
	Chilca Dulce	ACR151						BI
Fabaceae								
<i>Acacia macracantha</i> Humb. & Bonpl. ex Willd.	Faique	JULS172		15				W
<i>Caesalpinia paipai</i> Ruiz & Pav.	Pai pai	KMM581/ GER40		14	20			W
<i>Caesalpinia spinosa</i> (Molina) Kuntze	Taya or Tara	ACR111	10	23		10	11	Tonsillitis, Skin In, W
<i>Cajanus cajan</i> (L.) Huth	Chibato	ACR10						S
<i>Cassia fistula</i> L.	Caña Fistula	RBU/PL386		12				N

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			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Cassia fistula</i> L.	Ceiña Fistala	ACR88						N
<i>Desmodium molliculum</i> (Kunth) DC.	Manda yupa	AKT1162/ KMM392		8				W
<i>Leucaena leucocephala</i> (Lam.) De Wit	Arabisca	JULS104	7					W
<i>Lupinus mutabilis</i> Sweet	Chochos	ACR102						Nutrition
<i>Medicago sativa</i> L.	Trebol de agua	KMM463/ ACR51	10	7				Br
<i>Melilotus albus</i> Medik.	Fenogreco	KMM459						lfs
<i>Mimosa albida</i> Humb. & Bonpl. ex Willd.	Tapa Tapa	ACR79						S
<i>Mimosa albida</i> Humb. & Bonpl. ex Willd.	Tutapure	KMM560/ ACR190/ AKT1118		12				W
<i>Myroxylon balsamum</i> (L.) Harms	Quina Quina	JULS287		12				Br
<i>Myroxylon balsamum</i> (L.) Harms	Quina Quina	AKT1223						Br
<i>Prosopis pallida</i> (Humb. & Bonpl. ex Willd.) Kunth	Algarrobina	ACR203		15				Br
<i>Senna cf. bicapsularis</i> (L.) Roxb.	Alcaparilla	ACR194		15				Bl
<i>Senna monilifera</i> H.S. Irwin & Barnaby	Sen	KMM470/ AKT1112						Purgative
<i>Senna monilifera</i> H.S. Irwin & Barnaby	Sen	KMM470		17				Purgative
<i>Spartium junceum</i> L.	Ratanica	AKT1222/ KMM407		14				Bl
<i>Spartium junceum</i> L.	Retama	ACR108						Bl
<i>Trifolium repens</i> L.	Trebol	AKT1194/ KMM577						K
Gentianaceae								
<i>Coutoubea ramosa</i> Aubl.	Genciana	GER173		12				W
<i>Gentiana sedifolia</i> Kunth	Horma de amarilla	ACR92						S
<i>Gentianella bicolor</i> (Wedd.) Fabris ex J.S. Pringle	Amargon	ACR156						Ar
<i>Gentianella bicolor</i> (Wedd.) Fabris ex J.S. Pringle	Corpusway	KMM526		18				Ar
<i>Gentianella brunneotincta</i> (Gilg.) J.S. Pringle	Yanga macha	KMM403						Uterus If after birth

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			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Gentianella crassicaulis</i> J.S. Pringle	Agencia	ACR20/ KMM422		13	12			Db
<i>Gentianella dianthoides</i> (Kunth) Fabris ex J.S. Pringle	Chagape	ACR155		10				BI
<i>Gentianella dianthoides</i> (Kunth) Fabris ex J.S. Pringle	Genciana	KMM576						BI
<i>Gentianella graminea</i> (Kunth) Fabris	Sumaran	AKT1164						BI
<i>Gentianella</i> sp.	Hornamo asmarillo	ACR92						BI
Geraniaceae								
<i>Erodium cicutarium</i> (L.) L'Hér. ex Aiton	Agujilla	AKT1171						Br
<i>Erodium cicutarium</i> (L.) L'Hér. ex Aiton	Cachujillo	KMM578		15				Br
<i>Geranium ayavacense</i> Willd. ex Kunth	Pasuchaca	JULS48		12				If
<i>Geranium</i> cf. <i>ayavacense</i> Willd. ex Kunth	Pasuchaca	KMM401						If
<i>Geranium sessiliflorum</i> Cav.	Pasuchaca	KMM400		20	15			If
<i>Geranium sessiliflorum</i> Cav.	Pasuchaca	ACR38		12				If
<i>Pelargonium</i> cf. <i>odoratissimum</i> (L.) L'Hér.	Malva de Olor	ACR26		8				BI
Gigartinaceae								
<i>Gigartina chamissoi</i> (C. Agardh) J. Agardh	Cochayuyo	AKT393, 1226						Cholesterol
Illiciaceae								
<i>Illicium verum</i> Hook. f.	Anis estrella	AKT1208						Expel residues from the stomach of newborns
Iridaceae								
<i>Hesperoxiphion niveum</i> (Ravenna) Ravenna	Hierba de la justicia	AKT1100/ ACR45			9			S
Juglandaceae								
<i>Juglans neotropica</i> Diels	Nogales	AKT1111/ KMM435/ ACR71		11				W

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			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
Krameriaceae								
<i>Krameria lappacea</i> (Dombey) H.M. Burdet & B.B. Simpson	Retamia	ACR48	12	15				In
Lamiaceae								
<i>Clinopodium pulchellum</i> (Kunth) Govaerts	Panizara	ACR97						S
<i>Hyptis sidifolia</i> (L'Hér.) Briq.	Albaca serrana	ACR18			12			Colic
<i>Hyptis sidifolia</i> (L'Hér.) Briq.	Perroreya	ACR69, 18	10	13				Colic
<i>Marrubium vulgare</i> L.	Cordon de Muerto	KMM452						S
<i>Melissa officinalis</i> L.	Toronjil	ACR4						N
<i>Mentha X piperita</i> L.	Poleo	JULS29		6				Colic
<i>Mentha X piperita</i> L.	Poleo	ACR68						Colic
<i>Mentha spicata</i> L.	Menta	KMM453		12				Anthelmintic
<i>Minthostachys mollis</i> (Kunth) Griseb.	Chancas de muerto	AKT1142						Colic
<i>Ocimum basilicum</i> L.	Albaca	KMM437, 428/ ACR82		13				lfs after birth
<i>Origanum majorana</i> L.	Mejorana/ Tomillo	ACR24		13				Colic
<i>Origanum vulgare</i> L.	Oregano	KMM509		13	17			Colic
<i>Otholobium glandulosum</i> (L.) J.W. Grimes	Culen	KMM419/ AKT1134		10				Dr
<i>Otholobium mexicanum</i> (L.f.) J.W. Grimes	Culen	ACR67		14				Dr
<i>Rosmarinus officinalis</i> L.	Romero	ACR16/ AKT1129		17	11			Br
<i>Salvia cuspidata</i> Ruiz & Pav.	Salvia Blanca	RBU/PL315		18				S
<i>Salvia discolor</i> Kunth	Llatama	ISA151		13				Cough
<i>Salvia sagittata</i> Ruiz & Pav.	Salvia Negra	RBU/PL318		12				Cough
<i>Salvia</i> sp.	Alamo Silvestre	ACR184		14				Cough
<i>Salvia</i> sp.	Hierba del Aire	AKT1160						S
<i>Salvia</i> sp.	Hierba de los sietea vientos	KMM572						S
<i>Salvia</i> sp.	Paja Amargoza	KMM567						S
<i>Salvia</i> sp.	Paja del Aire	ACR115						S

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			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Salvia</i> sp.	Unquia Real	KMM550						S
<i>Satureja pulchella</i> (Kunth) Briq.	Panizara	KMM543		11				Br
<i>Satureja sericea</i> (C. Presl & Benth.) Briq.	Romerio	KMM397/ ACR100						S
<i>Scutellaria</i> cf. <i>scutellarioides</i> (Kunth) Harley	Poleo Gentil	ACR178						S
<i>Stachys lanata</i> Crantz	Veronica Macho	JULS13		12				Br
<i>Thymus vulgaris</i> L.	Tomillo	EHCHL169		16				Cough
	Salvia Real	ACR25		18				S
Lauraceae								
<i>Cinnamomum verum</i> J. Presl	Canela	KMM575		14				Br
<i>Persea americana</i> Mill.	Palta	AKT1120/ KMM444		10				Cough
Lichenes								
<i>Siphula</i> sp.	Pelo de Pierda	ACR187		10				Liver In
Linaceae								
<i>Linum</i> cf. <i>usitatissimum</i> L.	Linaza	KMM504						K
Loganiaceae								
<i>Buddleja utilis</i> Kraenzl.	Flora Blanca	AKT1131						Menstruation
Lycopodiaceae								
<i>Huperzia</i> sp.	Condor	AKT1170						S
<i>Huperzia</i> sp.	Condor	KMM541						S
<i>Huperzia</i> sp.	Condor crespó	KMM477						S
<i>Huperzia</i> sp.	Condor Misha	KMM479						S
<i>Huperzia</i> sp.	Condor Purga	KMM482						S
<i>Huperzia</i> sp.	Condor Purga	AKT1212						S
<i>Huperzia</i> sp.	Corontilla	KMM391						S
<i>Huperzia</i> sp.	Enredadera	AKT1183						S
<i>Huperzia</i> sp.	Trencilla blanca	KMM483						S
<i>Lycopodium thyooides</i> Humb. & Bonpl. ex Willd.	Trencilla	AKT1198, 1191						S
<i>Lycopodium</i> sp.	Guamingo	AKT1206	10					S
<i>Lycopodium</i> sp.	Simba	ACR116						S
Lythraceae								
<i>Cuphea</i> sp.	Hierba del Toro	KMM448		13				BI

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Cuphea strigulosa</i> Kunth	Sanguinaria	JULS33		12				BI
Malpighiaceae								
<i>Banisteriopsis caapi</i> (Spruce ex Griseb.) C.V. Morton	Ayohuasca	ACR135						S
Malvaceae								
<i>Gossypium cf. barbadense</i> L.	Algodon Pardo	AKT1227						W (dressing)
<i>Malva cf. parviflora</i> L.	Malva rosa	AKT1200		8				Br
<i>Malva cf. sylvestris</i> L.	Malva Blanca Alta	ACR8						W
<i>Malva</i> sp.	Malva Real	AKT1201						Br
<i>Ochroma cf. pyramidale</i> (Cav. ex Lam.) Urb.	Balsa	ACR206						Construction
<i>Urena cf. lobata</i> L.	Buenas Horas	ACR185						Dementia
Maranthaceae								
<i>Monotagma plurispicatum</i> (Körn.) K. Schum.	Patiquina	ACR114	7					S
Melastomataceae								
<i>Brachyotum naudinii</i> Triana	Carcilleja	ACR140		14				Blood circulation
<i>Brachyotum naudinii</i> Triana	Zarcilleja	AKT1189						Blood circulation
<i>Brachyotum tyrianthinum</i> J.F. Macbr.	Sarzilleja	EHCHL55		10				Blood circulation
<i>Miconia salicifolia</i> ((Bonpl. ex Naudin) Naudin	Mococho/ Porotillo	ACT204/ KMM544		11				Burns
Menispermaceae								
<i>Abuta grandifolia</i> (Mart.) Sandwith	Abuta	ACR136	10	12		8		C
Monimiaceae								
<i>Peumus boldus</i> Molina	Boldo (Chilean)	AKT1132		15				K
Moraceae								
<i>Brosimum rubescens</i> Taub.	Palo Sangre	KMM570						Ar
<i>Ficus carica</i> L.	Higo	ACR73		8				Db
<i>Ficus carica</i> L.	Higo	AKT1128						Db
Myrtaceae								
<i>Eucalyptus citriodora</i> Hook.	citradora	KMM 454		20	10			Db

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Eucalyptus globulus</i> Labill.	Eucalipto	AKT1110/ KMM408		20	18			Br
<i>Eugenia cf. obtusifolia</i> Cambess.	Rumilanche	AKT1143		14	18			In
<i>Eugenia obtusifolia</i> Cambess.	Limoncillo	ACR180, 76		15				In
<i>Eugenia</i> sp.	Arrayn	ACR19		13				In
<i>Myrcianthes discolor</i> (Kunth) McVaugh	Lanche	RBU/PL271		16				In
<i>Psidium guajava</i> L.	Guanabana	KMM339		7				Cancer
<i>Scutia spicata</i> (Humb. & Bonpl. ex Willd.) Weberb.	Pial	ACR207		12				S
<i>Syzygium aromaticum</i> (L.) Merr. & L.M. Perry	Clavo de olor	ACR188						Pain
<i>Syzygium jambos</i> (L.) Alston	Poma Rosa	GER173/ ACR174		10				Dr
Olacaceae								
<i>Heisteria acuminata</i> (Humb. & Bonpl.) Engl.	Chuchu Wassi	KMM507	12	14				Cough
<i>Ximenia americana</i> L.	Limoncillo	JULS184	12	10				Menstrual regulation
Oleaceae								
<i>Jasminum</i> sp.	Jasmin	AKT1211		15				S
Onagraceae								
<i>Fuchsia</i> sp.	Añasquero	AKT1187						Ar
Orchidaceae								
<i>Aa paleacea</i> (Kunth) Rchb.f.	Hierba Sola	ACR144/ AKT1158/ KMM530						C
<i>Epidendrum</i> sp.	Caballero	ACR168						S
<i>Epidendrum</i> sp.	Espadilla	ACR165						S
<i>Epidendrum</i> sp.	Hierba de la Espada	AKT1177		8				S
<i>Epidendrum</i> sp.	Hierba del Gallo	AKT1176						S
<i>Epidendrum</i> sp.	Lancilla	KMM484						S
<i>Maxillaria</i> sp.	Huome Huome	AKT1193						S
<i>Stelis</i> sp.	Hierba del Olor	KMM492						S
<i>Stelis</i> sp.	Oro	AKT1152						S

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
Orobanchaceae								
<i>Escobedia grandiflora</i> (L.f.) Kuntze	Azafran	JULS110		22				Br, Pneumonia
Papaveraceae								
<i>Argemone mexicana</i> L.	Cardo Santo	KMM433						S
<i>Argemone mexicana</i> L.	Cardo Santo	ACR61/ AKT1135			16			S
Passifloraceae								
<i>Malesherbia ardens</i> J.F. Macbr.	Veronica	AKT1232		9				Br
<i>Passiflora ligularis</i> A. Juss.	Hoja de granada	KMM420		8				In
<i>Passiflora ligularis</i> A. Juss.	Hoja de granadilla	AKT1097/ AKT1105/ KMM420						In
<i>Passiflora punctata</i> L.	Norgo	KMM510						Menstrual pain
<i>Passiflora quadrangularis</i> L.	Ajo Sacha	ACR121						Menstrual pain
<i>Passiflora</i> sp.	Hierba del Partero	AKT1122						Menstrual pain
Phyllanthaceae								
<i>Phyllanthus niruri</i> L.	Chanca Piedra	KMM517/ AKT1151		18				In
<i>Phyllanthus urinaria</i> L.	Chanca Piedra	JULS133		14				In
Phytolaccaceae								
<i>Petiveria alliacea</i> L.	Mocura	ACR52/ KMM447						S
<i>Phytolacca bogotensis</i> Kunth	Ailambo	KMM457		7				Malaria, Dengue, Yellow Fever
<i>Gallesia integrifolia</i> (Spreng.) Harms	Palo de Ajo	ACT116		19				Br
Piperaceae								
<i>Peperomia fraseri</i> C.DC.	Hierba de Dolor	AKT1133						S
<i>Peperomia inaequalifolia</i> Ruiz. & Pav.	Congona	KMM534/ AKT1148						S
<i>Peperomia quadrifolia</i> (L.) Kunth	Piri Piri	KMM532						S
<i>Peperomia</i> sp.	Congonilla	AKT1190						S
<i>Peperomia</i> sp.	Lancetillo	ACR21						S
<i>Piper acutifolium</i> Ruiz.& Pav.	Matico (sierra)	ACR15						Liver In

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Piper aduncum</i> L.	Matico	ACR12/ AKT1150	12					W, Br
Plantaginaceae								
<i>Galvezia fruticosa</i> J.F. Gmel.	Macacha	ACR64						Ar
<i>Plantago linearis</i> Kunth	Llanten Serrano	JULS35		10				W
<i>Plantago major</i> L.	llanten	KMM411		11				Br
<i>Plantago sericea</i> Ruiz. & Pav.	Pajilla blanca	ACR186						S
<i>Plantago sericea</i> Ruiz. & Pav. var. <i>lanuginosa</i> Griseb.	Paja Blanca	KMM499/ AKT1182/ ACR186		13	15			Vaginal discharge
Poaceae								
<i>Arundo donax</i> L.	Carrizo	KMM389		12				Haemorrhoids
<i>Cymbopogon citratus</i> (DC.) Stapf	Hierba Luisa	ACR5						Panacea
<i>Cynodon dactylon</i> (L.) Pers.	Gramadulce	KMM450/ ACR64						Ovarian cysts
<i>Digitaria ciliaris</i> (Retz.) Koeler	Pata de Gallina	JULS220		8				S
<i>Distichlis spicata</i> (L.) Greene	Gramadulce	AKT1096						S
<i>Gynerium sagittatum</i> (Aubl.) P. Beauv.	Carrizo	ACR47						S
<i>Saccharum officinarum</i> L.	Cana cana	AKT121/ KMM502						K
	Barra de oro	KMM415						S
Polemoniaceae								
<i>Cantua buxifolia</i> Juss. ex Lam.	Cantuta o Cando	ACR86						S
<i>Cantua quercifolia</i> Juss.	Adormidora	AKT1161						S
Polygonaceae								
<i>Polygonum hydropiperoides</i> Michx.	Pica Pica	ACR80/ JULS223		9				W
<i>Rumex crispus</i> L.	Mala Hierba	KMM496						K
Polypodiaceae								
<i>Polypodium cf. crassifolium</i> L.	Calaguala	AKT1137						K
	Contra Hierba	KMM464			9			S
Proteaceae								
<i>Oreocallis grandiflora</i> (Lam.) R. Br.	Proteaceae	ACR176/ KMM514/ AKT1173		10				K

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
Pteridaceae								
<i>Argyrochosma nivea</i> (Poir.) Windham	Doradillo	KMM460						S
<i>Cheilanthes pruinata</i> Kaulf.	Cuty Cuty	KMM461		13				S
<i>Cheilanthes</i> sp.	Cuti Cuti	AKT1108		14				S
<i>Jamesonia alstonii</i> A.F. Tryon	Baston del Inca	ACR112						S
<i>Notholaena sulphurea</i> (Cav.) L. Sm.	Doradilla	KMM458						S
Ranunculaceae								
<i>Laccopetalum giganteum</i> (Wedd.) Ulbr.	Pacra	AKT1119		9				Br
<i>Ranunculus krapfia</i> DC. ex Deless.	Marrajudio	ACR179						S
<i>Thalictrum decipiens</i> B. Bovin	Chontilla	ISA15	12	14				Mumps
Rosaceae								
<i>Cydonia oblonga</i> Mill.	Membrillo	JULS194		11				Depression
<i>Cydonia oblonga</i> Mill.	Membrillo	ACR56						Depression
<i>Margyricarpus pinnatus</i> (Lam.) Kuntze	China Lincla Verde	ACR146		8				S
<i>Polylepis racemosa</i> Ruiz. & Pav.	Quinal	ACR3		14				If after birth
<i>Prunus serotina</i> Ehrh.	Capuli	ACR172						Ar
<i>Prunus serotina</i> Ehrh. ssp. <i>capuli</i> (Cav.) McVaugh	Helialiso	EHCHL94		12				W
<i>Rubus robustus</i> C. Presl.	Sarsa mora	ACR70			16			Br
<i>Sanguisorba minor</i> Scop.	Pinpinela	ACR23		13	13			Menstrual regulation
	Ornamo Leon	AKT1214						S
Rubiaceae								
<i>Cinchona officinalis</i> L.	Cascarilla	ACR123/ KMM525		8				Cough
<i>Morinda citrifolia</i> L.	Noni	ACR160						Panacea
<i>Uncaria tomentosa</i> (Willd. ex Roem & Schult.) DC.	Uña de Gato	ACR198	8	10				Br
Rutaceae								
<i>Citrus limetta</i> Risso	Lima	KMM425		10				N
<i>Citrus limon</i> (L.) Osbeck	Limon	KMM424						K
<i>Citrus sinensis</i> (L.) Osbeck	Naranja	JULS202		16				N

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Ruta graveolens</i> L.	Ruda Seca	AKT1105/ KMM430		14				Ab
Salicaceae								
<i>Salix chilensis</i> Molina	Sause	ACR42/ AKT1006		12				Malaria, Fever
Santalaceae								
<i>Phoradendron</i> sp.	Suelda con suelda	ACR189	7		10			S
Scrophulariaceae								
<i>Capraria peruviana</i> Benth.	Flor Arenilla	RBU/PL374		11				K
<i>Capraria peruviana</i> Benth.	Te de Judío	KMM574			12			K
	Chiciricoma	KMM440			17			W
Smilacaceae								
<i>Smilax kunthii</i> Killip & C.V. Morton	Palo de la China	KMM20	16	18				Cancer
<i>Smilax</i> sp.	Palo China	KMM516		10				Cancer
Solanaceae								
<i>Brugmansia candida</i> Pers.	Misha Blanca	KMM564				8		S
<i>Brugmansia sanguinea</i> (Ruiz & Pav.) D. Don	Misha	KMM528						S
<i>Brugmansia</i> sp	Misha	ACR90						S
<i>Brugmansia</i> sp	Misha	AKT1149						S
<i>Brugmansia suaveolens</i> (Humb. & Bonpl. ex Willd.) Bercht. & C. Presl	Floripondio	AKT1116						S
<i>Capsicum chinense</i> Jacq.	Aji Panca	GER203	9	15				S
<i>Cestrum auriculatum</i> L'Hér.	Hierba Santa (Sierra)	ACR36		10				W
<i>Cestrum humboldtii</i> Francey	Hierba Santa	ACR83		9				W
<i>Cestrum</i> sp.	Agrasejo	AKT1121						W
<i>Cestrum</i> sp.	Hierba Santa	KMM426						W
<i>Cestrum strigilatum</i> Ruiz & Pav.	Santa Maria	JULS245	13	19				Menstrual regulation
<i>Datura ferox</i> L.	Floripondio	KMM441, 582						S
<i>Lycopersicon hirsutum</i> Dunal	Ambulluco de Muerto	ISA31		26				S
<i>Nicotiana tabacum</i> L.	Tabaco	KMM388, 557		8				S
<i>Nolana</i> cf. <i>humifusa</i> (Gouan) I.M. Johnst.	Hierba de Señorita	AKT1124						S

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
<i>Solanum americanum</i> Mill.	Hierba Mora/ Cushay	ACR37, 99		16				Sinusitis
<i>Solanum mammosum</i> L.	Torito	AKT1138						S
Tropaeolaceae								
<i>Tropaeolum minus</i> L.	Mastuerzo	JULS81		17				In of the stomach
Ulmaceae								
<i>Celtis loxensis</i> C.C. Berg	Palo Huaco	KMM561		6				Fertility, Br
Urticaceae								
<i>Pilea microphylla</i> (L.) Liebm.	Contra Hierba	RBU/PL282		8				K
<i>Urtica magellanica</i> Juss. ex Poir.	Ortiga Negra	ACR72						BI
<i>Urtica urens</i> L.	Chin Chin	ACR199						S
Verbenaceae								
<i>Aloysia scorodonioides</i> (Kunth) Cham.	Santa Maria	ACR43		12				Depression, Pain
<i>Aloysia triphylla</i> Royle	Cedron	KMM398		13				Depression, Pain
<i>Aloysia triphylla</i> Royle	Cedron	AKT1126/ ACR159						Depression, Pain
<i>Lantana scabrosiflora</i> Kunth	Pacha Rosa	AKT1123/ ACR50						Cold
<i>Lantana</i> sp.	Hierba del Hombre	AKT1202			9			Cold
<i>Verbena litoralis</i> Kunth	Verbena	JULS77		12				W
	Hierba de Susto	AKT1210						S
Violaceae								
<i>Viola</i> sp.	Pensamiento	ACR27						S
<i>Viola</i> sp.	Violeta	KMM426						S
<i>Viola tricolor</i> L.	Cinta de Novia	ACR195						S
Zingiberaceae								
<i>Zingiber officinale</i> Roscoe	Gengibre	ACR196						Br
	Chimipampana Blanca	KMM485		12				W
Unknown								
	Ajo Caspi	KMM518		9				S
	Ajo Caspi	ACR133						S

Plant Family Species	Common name	Collector & Col#	Inhibition diameter (mm). Blank cells indicate lack of antibacterial activity					Traditional application
			<i>E. coli</i>	<i>S. aureus</i>	<i>S. aureus</i>	<i>S. enterica</i>	<i>P. aeruginosa</i>	
	Anti ajo	AKT1219						S
	Arnica	ACR193		11			2	W
	Bejuco de montaña	ACR173						S
	Bledaco	KMM501		13				
	Canchalagua Morada	ACR143						S
	Cardo Bendito	KMM508						S
	Chimipanpana	AKT1139						S
	Chivato	KMM569						S
	Choloque	ACR84						S
	Cina Cina	ACR164						S
	Contra Hechizo	ACR106						S
	Cucho	ACR161						S
	Flor Dracaena	KMM559						S
	Hercampuri	ACR200						S
	Huarate	AKT1209		11				Db
	Lengua perro	ACR117						S
	Llanten Blanco	AKT1231			14			S
	Llanten Blanco	AKT1123						S
	Milagroso	AKT1130						S
	Misqui Chilca	ACR137						S
	Motelilla	KMM488						S
	Paja del Susto	ACR104						S
	Pegajosa	ACR60						S
	Polen de Zapote	AKT1230						Tuberculosis
	Poma Rosa	ACR174			8			Nutrition
	Violeta	KMM423						S
	Yelama	ACR110						S
	Zarzaparrilla	AKT1136						S

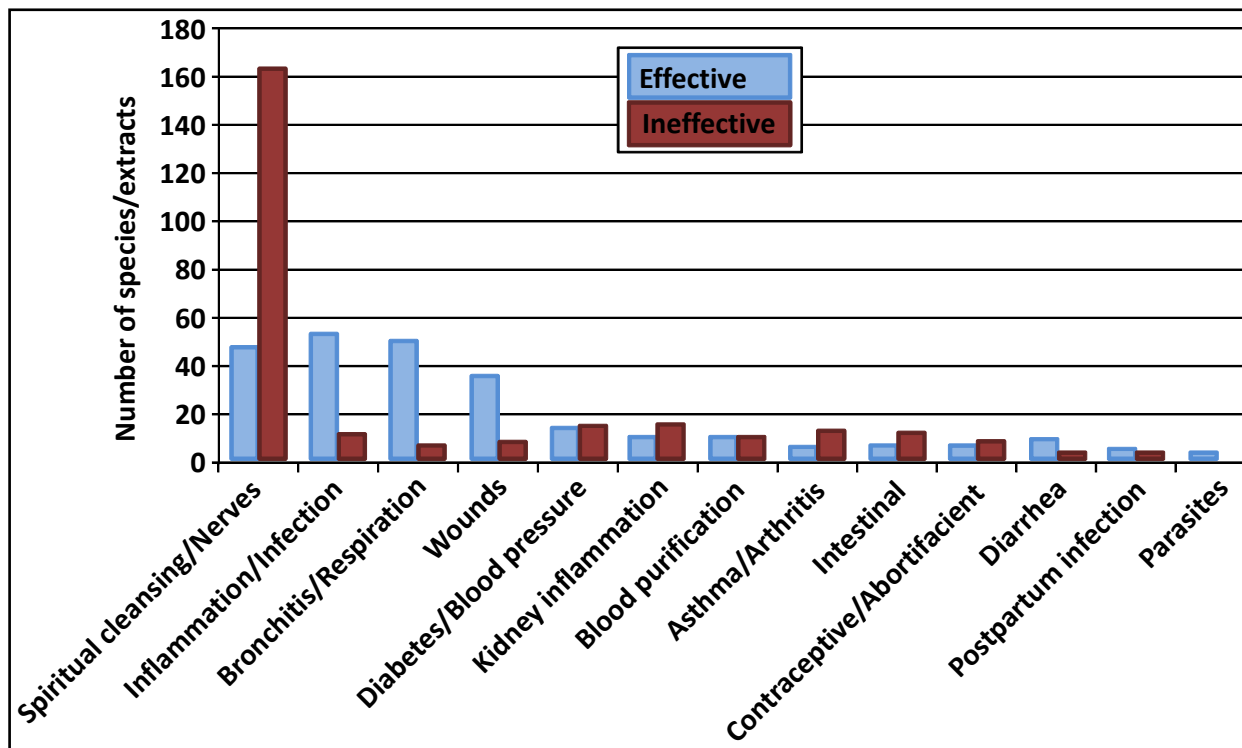


Figure 3. Antibacterial efficacy of plants traditionally used as anibacterials in the area of Trujillo and Chiclayo, northern Peru.

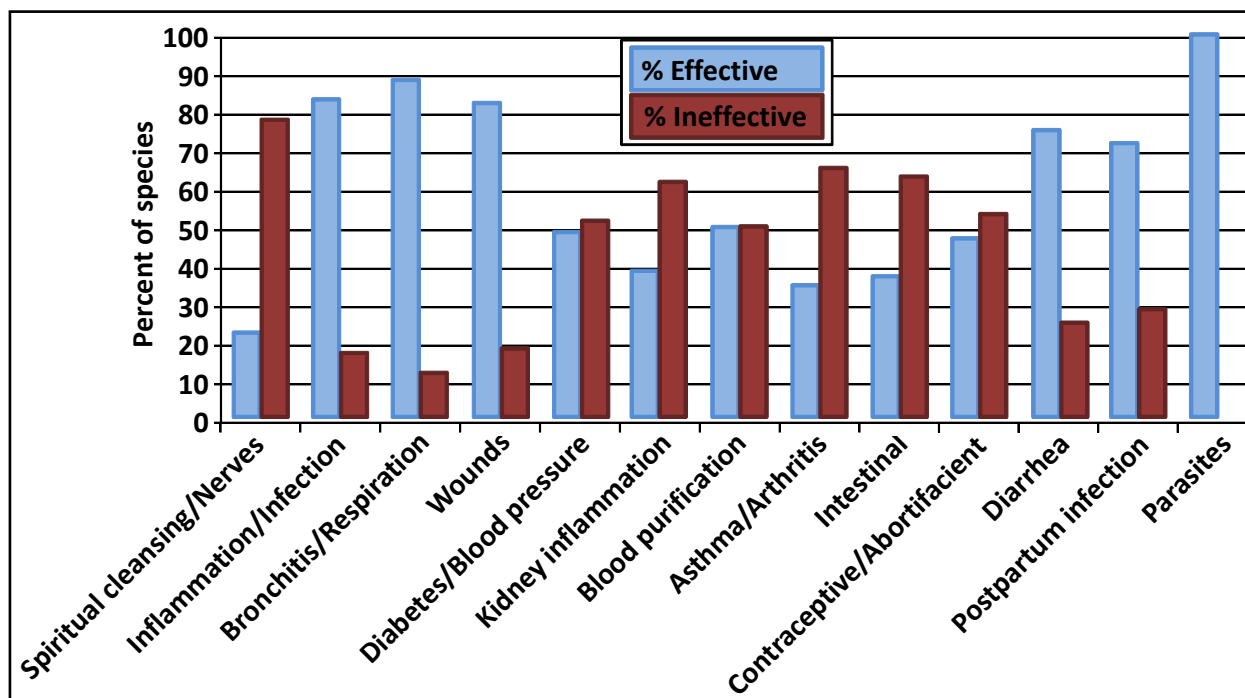


Figure 4. Efficacy percent per application category Trujillo and Chiclayo research area of northern Peru.

Geranium sessiliflorum Cav., *Hedyosmum racemosum* (Ruiz. & Pav.) G. Don., *I. herbstii*, *Lycopersicon hirsutum* Dunal, *Mauria heterophylla* Kunth, *Phyllanthus niruri* L., *Porophyllum ruderale* (Jacq.) Cass., *Salvia cuspidata* Ruiz & Pav., *Senecio chionogeton* Wedd., *Smilax kunthii* Killip & C.V. Morton, *Tagetes erecta* L. and *Taraxacum officinale* F.H. Wigg. showed high activity against *S. aureus*. The same holds true for *Ephedra americana* Humb. & Bonpl. ex Willd., *Gentianella bicolor* (Wedd.) Fabris ex J.S. Pringle, *Mandevilla trianae* Woodson. However, extracts of these three species were highly inconsistent in their efficacy.

Discussion

The results of the presented study give potentially interesting leads for future antibiotic research. They do however also give close insight into traditional plant selection, and last, caution against the simple transfer of traditional applications into Western science. The comparison of closely related species traditionally employed for different purposes (e.g., different *Alternanthera* spp., *Passiflora* spp., *Senecio* spp. and *Salvia* spp. for spiritual purposes and against bacterial infections) showed that the “spiritual” species normally were not effective against bacteria, while the species used as antibacterials had increased effectiveness. The example of *Plantago sericea* Ruiz. & Pav. var. *sericea* (used in **seguros**, no efficacy) and *Plantago sericea* Ruiz. & Pav. var. *lanuginosa* Griseb. (used for vaginal infections, high efficacy against *S. aureus*) is a particularly compelling case that indicates the sophistication of traditional knowledge. However, we did find examples like *Chuquiragua* spp., where closely related species were used as antibacterials, but only one of them did in fact show efficacy, clearly indicating that in this case traditional knowledge did not produce reliable results.

On the other hand, extracts of the same species traditionally used to treat infections often produced vastly diverging results when collected from different localities. Good examples are *Iresine herbstii* Hook., *Schinus molle* L., *Eustephia coccinea* Cav., *Oreopanax eriocephalus* Harms, *Myroxylon balsamum* (L.) Harms, *Spartium junceum* L., or *Gentianella dianthoides* (Kunth) Fabris ex J.S. Pringle. These species did not produce particularly high inhibition rates in any case, and were not the first choice of healers when trying to find remedies for bacterial infections. Many traditional remedies for concepts like “kidney inflammation” did not produce any antibacterial results, which underlines that research into efficacy does need to closely take traditional disease concepts into account.

Many remedies used for spiritual healing and other non-infection purposes did show antibacterial efficacy *in-vitro*, but were not listed as such by the local healers. This might be explained by the fact that they either are very inconsistent in their activity (e.g., *Brachyotum naudinii* Triana, *Cy-*

donia oblonga Mill., *Hypericum laricifolium* Juss., *Hyptis sidifolia* (L'Hér.) Briq., *Lonicera japonica* Thunb. ex Murray, *Loricaria* spp., *Mentha X piperita* L., *M. trianae*), or are so closely related that identification, especially when dried, can be a problem, e.g. in the case of *Baccaris* spp., *Gentianella* spp., and *Valeriana* spp., or are prone to toxic side effects like *E. americana* and *Brugmansia* spp.

Almost all remedies are traditionally prepared as water extracts, although ethanol (in the form of sugarcane spirit) is readily available. This might at a first glance seem astonishing, given the low efficacy of water extraction found in this study. However, initial results from Brine-Shrimp toxicity assays indicate that the ethanolic extracts are by far more toxic than water extracts of many species, and thus ethanolic extraction might in many cases not be suitable for application in patients. This again indicates the considerable sophistication and care with which traditional healers in northern Peru chose their remedies for a specific purpose.

Conclusions

Simple agar assays alone are not reliable for determining the efficacy of plants used in traditional medicine. The present study however confirms that simple laboratory methods are indeed well suited for the initial assessment of the efficacy of traditionally used medicinal plants in inhibiting bacterial growth. A comparison to the traditional uses also indicated that local knowledge can provide important leads for the development of new drugs: Our hypotheses that plants traditionally used as antibacterials do have a much higher incidence of efficacy than other medicinally used species, and that plants used for cleansing baths are more likely to have antibacterial properties than other species used for ritual purposes could be verified. However, if traditional knowledge is to serve as a lead, the results clearly indicate, that a very careful assessment of traditional disease concepts is needed in order to draw the right conclusions. It also needs to be taken into account that many plants are usually traditionally prepared in complex mixtures, and single species are rarely applied (Bussmann & Sharon 2006, 2007). An efficacy analysis of these mixtures, as well as assays to evaluate the toxicity of single species as well as mixtures would be an interesting comparative study.

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