



# The singularity of the medicinal knowledge of the Huni Kuĩ people from the western Brazilian Amazon

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## Correspondence

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## Research

### Abstract

*Background:* Studies that aim to analyze the transcultural traditional knowledge from different countries still in their infancy but are essential to favor an equal and fair division of the benefits resulting from the exploration of genetic resources and to develop sustainable strategies for conservation. The goal of this research is to compare the traditional knowledge about medicinal plants by the Huni Kuĩ people with the knowledge from other cultures, in order to assess the singularity of the Huni Kuĩ knowledge and to identify the convergence in medicinal plants use with other cultures.

*Methods:* We did a literature review for 83 plant species and compiled all traditional knowledge available on the Scielo PubMed and Google Scholar. We searched for the scientific name of each species and its synonyms (382) plus the words "ethnobotany", "ethnobotanical" or "medicinal" as inclusion criteria.

*Results:* We found 625 papers for 54 plants. About 73% of the medicinal services found for these species were considered, and 90% of all medicinal services cited were from the Huni Kuĩ people. The other 10% (20 species) were shared with other communities. Most of the research was conducted in the Brazilian Amazon, but also in the other countries.

*Conclusion:* Our results show the singularity of the Huni Kuĩ knowledge and the complexity in the distribution of traditional knowledge, which highlights the importance of projects that document the traditional knowledge, in order to create new conservation strategies and public policies.

*Keywords:* traditional knowledge, indigenous people, indigenous language, medicinal services, ethnobotany

### Background

The United Nations recognized that the preservation of native languages around the world is of such importance and proclaimed the years between 2022 and 2032 as the International Decade of Indigenous Languages, in order to raise global awareness of their importance for sustainable development and risk of extinction. The extinction of

indigenous languages is one of the main reasons for loss of traditional indigenous knowledge (Cámara-Leret & Bascompte 2021) and the loss of knowledge about plants is directly related to the reduction in use of these languages (Saynes-Vasquez *et al.* 2013). Cámara-Leret & Bascompte (2021) pointed out that the whole spectrum of indigenous languages need to be considered in conservation plans, as each society and its language can have unique knowledge that can be complementary to other societies in search for good health practices. Initially, many researchers assumed that traditional medicinal knowledge was shared among different indigenous language, but when they investigated this matter further, they found high rates of singularity in traditional medicinal knowledge from North America, northwest Amazon and New Guinea (Cámara-Leret & Bascompte 2021), suggesting that there are singularities in traditional knowledge even in languages that exist close to each other in areas of high floristic similarity (Cámara-Leret & Dennehy 2019).

The Huni Kuĩ people live in western Amazon, between Acre state in Brazil and the foothills of the Andes (Iglesias 2014). The Hatxa Kuĩ is their original language, and it is part of the pano linguistic family, one of the smallest linguistic families from South America. Muru & Quinet (2014) published a book with a list of medicinal plants used by the Huni Kuĩ people from Jordao River, to register in writing the knowledge about medicinal plants for future generations, a long-time wish of the shaman elders, that fear their accumulated knowledge may be lost (Muru & Quinet 2014). Although the Jordao river region has the highest number of Hatxa Kuĩ speakers, the preservation of the language was impacted in several localities (Kaxinawa 2011).

Muru & Quinet (2014) mention several widespread plants occurring in Brazil and across Brazilian borders, and many of those are found in degraded areas (Penedo 2023). We can assume that species with wide distributions that occur in degraded areas have higher probabilities of being used, as they are readily available in human settlements, and territorial occupation patterns and intercultural exchange can favor knowledge exchange among different communities. In this context, we ask the following questions: does the traditional knowledge about medicinal plants of the Huni Kuĩ people is similar to the knowledge of other people and communities, or is it unique?

Studies that aim to analyze the transcultural traditional knowledge from different countries still in their infancy but are essential to favor an equal and fair division of the benefits resulting from the exploration of genetic resources, a goal of the Nagoya Protocol (Cámara-Leret *et al.* 2014). The use of a plant through time can represent its "tradition" related to the way it is used (Ricardo *et al.* 2018) and this criterium can help the inclusion of certain plants in primary healthcare (Kniess & Wiesner 2019, Pires *et al.* 2020) and on prioritization of conservation and sustainable development efforts (Janni & Bastien 2000), which are goals of the Biological Diversity Convention.

In this study we compare the recovered traditional knowledge about medicinal services of 83 plants used by the Huni Kuĩ people from the Jordao River, mentioned in Muru & Quinet, with other ethnobotanical studies, to quantify how much of this knowledge is unique to the Huni Kuĩ culture, and identify potential strategies to preserve this traditional knowledge. We also aim to classify diseases and medicinal uses into an international classification system and identify which relationship between other study regions in order to of study to populations make use of these plants, for how long they use them, and where these communities live, contribute with of the understanding the convergences and singularities of Huni Kuĩ knowledge.

## Material and Methods

In order to assess and compare the Huni Kui traditional knowledge, we used the list of 83 plant species reported by Muru & Quinet (2014) and performed a literature review of all the traditional uses of these species found in peer-reviewed journals, books, thesis and dissertations, using Scielo, Pubmed and Google Scholar online databases. In our online search we used as inclusion criteria (Félix Silva *et al.* 2017) the scientific names and synonyms of the 83 plant species used by the Huni Kui (Muru & Quinet, 2014) and the following terms: "ethnobotany", "ethnobotanical" or "medicinal". We considered studies available until between May 2020 and May 2021. Studies where the medicinal use of the plants were not directly stated were excluded.

We used Microsoft Excel to organize and compile the data, and we recorded: year published, medicinal use, country where the research was conducted and region. In order to include as much information as possible and because of the heterogeneity of information available, "region" was considered as any specific biogeographic space, such as "Amazon", "Andes", "Caribbean", "Atlantic Forest", for example.

The first step on this review was to select papers with at least one medicinal use for each plant, excluding references with repeated information, such as published in a thesis and in a paper that was published later. For this, we checked the medicinal use mentioned, the locality, authors and year of the publication.

The medicinal use of plants by the Huni Kuĩ (Muru & Quinet 2014) and the use of plants from the literature review were classified according to the International Classifications of Primary Care - ICPC, currently the most adequate classification for traditional knowledge data (Weckerle *et al.* 2018). We created 29 additional categories based on descriptions of use, because some could not fit any of the ICPC classes. X01 - Inflammation, X02 - Hallucinogenic, X03- Analgesic, X04 Tumor/Cancer, X05 - Astringent, X06 - Antibiotic, X07 - Ulcer, X08 - Spasm, X09 - Infection, X10 - To attract wealth, X11 - Bad luck, X12 - Evil eye, X13 - Prevention, X14 - For babies, X15 - Repellent, X16 For kids, X17- Veterinary use, X18 - Body cleanse, X19 - Refresh, X20 - Hunting, X21 - Protection, X22 - Witchcraft, X23 - Rituals, X24 - Winti ritual, X25 - Amaci Ritual, X26 - For *ramo de ar*, X27 - Shamanic use, X28 - *Nisu*, X29- Climate.

After this classification, we used a combination of the plants and medicinal use classes to assess the medicinal services for each taxon, following Cámara-Leret & Bascompte (2021). We classified as "unique" the medicinal service that was reported in only one study (Cámara Leret & Bascompte 2021), and we identified the "old" medicinal services, those with more than 30 years of documentation (Knowss & Wiesner 2019). We then analyzed the organized data qualitatively and quantitatively.

## Results

We recorded 625 references (supplementary material) for 54 plant species (Table 1). All references had at least one specific medicinal use for one of these taxa. We found 57 references related to 28 synonyms. The following taxa had records of medicinal use without enough information to include them into any of the categories of use considered in this study: *Adenocalymma bracteatum*, *Begonia maynensis*, *Catasetum barbatum*, *Faramea torquata*, *Geogenanthus poeppigii*, *Gloeospermum sphaerocarpum*, *Heliconia schumanniana*, *Hirtella triandra*, *Nautilocalyx bullatus*, *Psychotria pedunculosa*, *Renealmia breviscapa*, *Senna loretensis*, *Solanum anceps*, *Solanum sessile*, *Vatairea fusca* and *Xylopia polyantha*. We did not find any record of research about the medicinal use of *Aphelandra acrensis*, *Floscopa elegans*, *Goepertia microcephala*, *Herpetacanthus rotundatus*, *Nautilocalyx pallidus*, *Peperomia sulcata*, *Philodendron burle-marxii*, *Philodendron toshibai*, *Pilocarpus manuensis*, *Piper crassistilum*, *Piper nigropunctatum*, *Sloanea garckeana* and *Stigmaphyllon florosum*.

Table 1. Plant species, their synonyms and number of references for each species. Nref = number of references.

Botanical Family Scientific Name	Synonyms	Nref
<b>Acanthaceae</b>		
<i>Fittonia albivenis</i> (Lindl. ex Veitch) Brummitt	<i>Fittonia verschaffeltii</i> Coem.	7
<b>Anacardiaceae</b>		
<i>Astronium graveolens</i> Jacq.	<i>Astronium gracile</i> Engl., <i>A. graveolens</i> var. <i>planchonianum</i> (Engl.) Engl., <i>A. planchonianum</i> Engl.	8
<b>Annonaceae</b>		
<i>Annona ambotay</i> Aubl.		10
<i>Unonopsis guatterioides</i> (A.DC.) R.E.Fr.	<i>Uvaria guatterioides</i> A.DC., <i>Trigynaea angustifolia</i> Benth., <i>Unonopsis angustifolia</i> (Benth.) R.E.Fr., <i>U. lindmanii</i> R.E.Fr.	3
<b>Apocynaceae</b>		
<i>Couma macrocarpa</i> Barb. Rodr.	<i>Couma capiron</i> Pittier, <i>C. caurensis</i> Pittier, <i>C. guatemalensis</i> Standl., <i>C. sapida</i> Pittier	5
<i>Tabernaemontana sananho</i> Ruiz & Pav.	<i>Bonafousia sananho</i> (Ruiz & Pav.) Markgr., <i>Tabernaemontana poeppigii</i> (Müll.Arg.) Miers, <i>Tabernaemontana poeppigii</i> Müll.Arg.	19
<b>Araceae</b>		
<i>Anthurium croatii</i> Madison		2
<i>Monstera spruceana</i> (Schott) Engl.	<i>Tornelia spruceana</i> Schott	4
<b>Araliaceae</b>		

<i>Oreopanax capitatus</i> (Jacq.) Decne. & Planch	<i>Aralia capitata</i> Jacq., <i>A. multiflora</i> Pohl ex DC., <i>A. ovata</i> Sessé & Moc., <i>Botryodendrum capitatum</i> (Jacq.) Endl., <i>Hedera capitata</i> (Jacq.) Sm., <i>H. frondosa</i> Salisb., <i>H. multiflora</i> DC., <i>Mesopanax capitatus</i> (Jacq.) R.Vig., <i>Oreopanax capitatus</i> var. <i>multiflorum</i> (DC.) Marchal, <i>O. capitatus</i> var. <i>multiflorus</i> (DC.) Marchal, <i>O. destructor</i> Seem., <i>Sciadophyllum capitatum</i> (Jacq.) Griseb.	1
<b>Bignoniaceae</b>		
<i>Bignonia aequinoctialis</i> L.	<i>Anemopaegma tonduzianum</i> Kraenzl., <i>Arrabidaea pseudochica</i> Kraenzl., <i>Bignonia aequinoctialis</i> var. <i>spectabilis</i> (Vahl) DC., <i>B. hostmannii</i> E.Mey., <i>B. incarnata</i> var. <i>caribaea</i> DC., <i>B. nitidissima</i> DC., <i>B. pilosa</i> A.Dietr. ex Steud., <i>B. sarmentosa</i> Bertol., <i>B. sarmentosa</i> var. <i>hirtella</i> Benth., <i>B. spectabilis</i> Vahl, <i>B. villosa</i> Vahl, <i>Cydista aequinoctialis</i> (L.) Miers, <i>C. aequinoctialis</i> (L.) Miers var. <i>aequinoctialis</i> , <i>C. aequinoctialis</i> var. <i>hirtella</i> (Benth.) A.H.Gentry, <i>C. aequinoctialis</i> var. <i>sarmentosa</i> (Bertol.) Govaerts, <i>C. amoena</i> Miers, <i>C. incarnata</i> (Aubl.) Miers, <i>C. picta</i> (Kunth) Miers, <i>C. pubescens</i> S.F.Blake, <i>C. sarmentosa</i> (Bertol.) Miers, <i>C. seemannii</i> Miers, <i>C. spectabilis</i> (Vahl) Miers, <i>Levyia nicaraguensis</i> Bureau, <i>Macfadyena bifolia</i> Miers, <i>Temnocydia aequinoctialis</i> (L.) Mart. ex DC., <i>T. spectabilis</i> Mart. ex DC.	4
<i>Callichlamys latifolia</i> (Rich.) K.Schum.	<i>Bignonia latifolia</i> Rich., <i>B. crucifera</i> Bertol. ex DC., <i>B. hondensis</i> Kunth, <i>B. rufinervis</i> Mart., <i>Callichlamys garnieri</i> Standl. & L.O.Williams, <i>C. peckoltii</i> Bureau ex K.Schum., <i>C. riparia</i> Miq., <i>C. rubiginosa</i> Miers, <i>C. rufinervis</i> (DC.) Miers, <i>C. splendida</i> Miers, <i>Lundia schomburgkii</i> Klotzsch, <i>Spathodea mansoana</i> DC., <i>Tabebuia latifolia</i> (Rich.) DC., <i>T. mansoana</i> DC., <i>T. rufinervis</i> DC., <i>T. speciosa</i> Standl.	8
<i>Handroanthus serratifolius</i> (A.H.Gentry) S. Grose	<i>Bignonia serratifolia</i> Vahl, <i>B. araliacea</i> Cham., <i>B. conspicua</i> Rich. ex DC., <i>B. flavescens</i> Vell., <i>B. patrisiana</i> DC., <i>Gelsemium araliaceum</i> (Cham.) Kuntze, <i>G. speciosum</i> (DC. ex Mart.) Kuntze, <i>Handroanthus araliaceus</i> (Cham.) Mattos, <i>H. atractocarpus</i> (Bureau & K.Schum.) Mattos, <i>H. flavescens</i> (Vell.) Mattos, <i>Tabebuia araliacea</i> (Cham.) Morong & Britton, <i>T. monticola</i> Pittier, <i>T. serratifolia</i> (Vahl) G.Nichols., <i>Tecoma araliacea</i> (Cham.) DC., <i>T. atractocarpa</i> Bureau & K.Schum., <i>T. conspicua</i> DC., <i>T. nigricans</i> Klotzsch, <i>T. patrisiana</i> DC., <i>T. serratifolia</i> (Vahl) G.Don, <i>T. speciosa</i> DC. ex Mart.	18
<i>Tanaecium bilabiatum</i> (Sprague) L.G.Lohmann	<i>Adenocalymma bilabiatum</i> (Sprague) Sandwith, <i>Arrabidaea bilabiata</i> (Sprague) Sandwith, <i>A. cuminaensis</i> A.Samp., <i>A. kuhlmannii</i> A.Samp., <i>Memora bilabiata</i> Sprague, <i>M. nobilis</i> Miers, <i>Pseudocalymma kuhlmannii</i> (A.Samp.) J.C.Gomes	2
<b>Boraginaceae</b>		
<i>Cordia nodosa</i> Lam.	<i>Cordia hispidissima</i> DC., <i>C. miranda</i> DC., <i>C. nodosa</i> var. <i>hispidissima</i> Fresen.	9
<b>Cannabaceae</b>		
<i>Trema micrantha</i> (L.) Blume		20
<b>Costaceae</b>		
<i>Dimerocostus strobilaceus</i> Kuntze		2
<b>Cyclanthaceae</b>		
<i>Cyclanthus bipartitus</i> Poit. ex A.Rich.	<i>Cyclanthus bifolius</i> Perr., <i>C. bipartitus</i> var. <i>gracilis</i> Drude, <i>C. cristatus</i> Klotzsch, <i>C. plumieri</i> Poit. ex A.Rich., <i>Discanthus odoratus</i> Spruce	6
<b>Euphorbiaceae</b>		
<i>Croton cuneatus</i> Klotzsch	<i>Croton kaeteuri</i> Jabl., <i>C. martii</i> Müll.Arg., <i>C. martii</i> var. <i>latifolius</i> Müll.Arg., <i>C. martii</i> var. <i>longifolius</i> Müll.Arg., <i>C.</i>	4

	<i>mimeticus</i> S.Moore, <i>C. monachinoensis</i> Jabl., <i>C. surinamensis</i> Müll.Arg., <i>C. tonantinensis</i> Jabl., <i>Macrocroton cuneatus</i> (Klotzsch) Klotzsch, <i>M. surinamensis</i> Klotzsch, <i>Oxydectes cuneata</i> (Klotzsch) Kuntze, <i>O. martii</i> (Müll.Arg.) Kuntze, <i>O. surinamensis</i> (Müll.Arg.) Kuntze	
<i>Euphorbia hyssopifolia</i> L.	<i>Anisophyllum hyssopifolium</i> (L.) Haw., <i>Chamaesyce brasiliensis</i> (Lam.) Small, <i>C. hyssopifolia</i> (L.) Small, <i>C. jenningsii</i> Millsp. ex Britton, <i>C. jonesii</i> (Millsp.) Millsp., <i>C. nirurioides</i> Millsp., <i>Euphorbia blanchetii</i> Miq. ex Boiss., <i>E. brasiliensis</i> Lam., <i>E. brasiliensis</i> var. <i>blancheti</i> Lam., <i>E. brasiliensis</i> var. <i>blanchetii</i> (Miq. ex Boiss.) Boiss., <i>E. brasiliensis</i> var. <i>genuina</i> Chodat & Hassl., <i>E. brasiliensis</i> var. <i>hyssopifolia</i> (L.) Boiss., <i>E. brasiliensis</i> var. <i>paraguayensis</i> Chodat, <i>E. brasiliensis</i> var. <i>pruinosa</i> (Chodat) Chodat & Hassl., <i>E. brasiliensis</i> var. <i>pulchella</i> Boiss., <i>E. brasiliensis</i> var. <i>uniflora</i> Chodat & Hassl., <i>E. domingensis</i> Spreng. ex Boiss., <i>E. hypericifolia</i> var. <i>falciformis</i> Klotzsch, <i>E. hyssopifolia</i> var. <i>blanchetii</i> (Miq. ex Boiss.) Oudejans, <i>E. hyssopifolia</i> var. <i>paraguayensis</i> (Chodat) Oudejans, <i>E. hyssopifolia</i> var. <i>pruinosa</i> (Chodat) Oudejans, <i>E. hyssopifolia</i> var. <i>pulchella</i> (Boiss.) Oudejans, <i>E. hyssopifolia</i> var. <i>uniflora</i> (Chodat & Hassl.) Oudejans, <i>E. jonesii</i> Millsp., <i>E. klotzschiana</i> Miq., <i>E. nirurioides</i> (Millsp.) Fawc. & Rendle, <i>E. pulchella</i> Kunth, <i>E. serrulata</i> Vell., <i>E. stenomeris</i> S.F.Blake	15
<b>Fabaceae</b>		
<i>Myroxylon balsamum</i> (L.) Harms		31
<b>Gesneriaceae</b>		
<i>Drymonia semicordata</i> (Poepp.) Wiehler	<i>Alloplectus semicordatus</i> Poepp., <i>Columnnea semicordata</i> (Poepp.) Kuntze, <i>Crantzia semicordata</i> (Poepp.) Fritsch	1
<b>Hernandiaceae</b>		
<i>Sparattanthelium burchellii</i> Rusby		1
<b>Hypericaceae</b>		
<i>Vismia cayennensis</i> (Jacq.) Pers.	<i>Hypericum cayennensis</i> Jacq., <i>Vismia cayennensis</i> (Jacq.) Pers. var. <i>cayennensis</i>	5
<i>Vismia guianensis</i> (Aubl.) Choisy	<i>Hypericum guianense</i> Aubl., <i>Vismia acuminata</i> (Lam.) Pers., <i>V. caparosa</i> Kunth, <i>V. ferruginea</i> Kunth, <i>V. guianensis</i> var. <i>acuminata</i> (Lam.) M.E.Berg, <i>V. reichardtiana</i> (Kuntze) Ewan	34
<b>Lamiaceae</b>		
<i>Vitex triflora</i> Vahl	<i>Macrostegia ruiziana</i> Nees, <i>Pyrostoma ternatum</i> G.Mey., <i>Vitex triflora</i> var. <i>angustiloba</i> Huber, <i>V. triflora</i> var. <i>coriacea</i> Huber, <i>V. triflora</i> var. <i>floribunda</i> Huber, <i>V. triflora</i> var. <i>hirsuta</i> Moldenke, <i>V. triflora</i> var. <i>kraatzii</i> Huber, <i>V. triflora</i> var. <i>quinquefoliolata</i> Moldenke, <i>V. triflora</i> var. <i>tenuifolia</i> Huber	5
<b>Loganiaceae</b>		
<i>Strychnos panurensis</i> Sprague & Sandwith		1
<b>Lygodiaceae</b>		
<i>Lygodium venustum</i> Sw		21
<b>Malvaceae</b>		
<i>Herrania mariae</i> (Mart.) Decne. ex Goudot	<i>Theobroma mariae</i> (Mart.) K.Schum.	3
<i>Quararibea guianensis</i> Aubl.	<i>Quararibea martini</i> Baill.	1
<b>Menispermaceae</b>		
<i>Chondrodendron tomentosum</i> Ruiz & Pav.		7

<b>Moraceae</b>		
<i>Sorocea guilleminiana</i> Gaudich.	<i>Sorocea grandifolia</i> S. Moore, <i>S. houlettianae</i> Gaudich., <i>Trophis hilariana</i> Casaretto	9
<b>Phytolaccaceae</b>		
<i>Petiveria alliacea</i> L.	<i>Petiveria alliacea</i> var. <i>octandra</i> (L.) Moq., <i>P. corrientina</i> Rojas Acosta, <i>P. hexandria</i> Sessé & Moc., <i>P. octandra</i> L., <i>P. paraguayensis</i> D. Parodi	282
<b>Piperaceae</b>		
<i>Piper callosum</i> Ruiz & Pav.	<i>Piper benianum</i> Trel., <i>P. callosum</i> var. <i>franciscoanum</i> C.DC., <i>P. poiretianum</i> C.DC.	23
<i>Piper hispidum</i> Sw.	<i>Artanthe asperifolia</i> (Ruiz & Pav.) Miq., <i>A. olfersiana</i> (Kunth) Miq., <i>Piper asperifolium</i> Ruiz et Pav., <i>P. bullatum</i> Vahl, <i>P. hirsutum</i> Sw., <i>P. hispidum</i> var. <i>magnifolium</i> C.DC., <i>P. hispidum</i> var. <i>trachydermum</i> (Trel.) Yunck., <i>P.</i> <i>trachydermum</i> Trel.	19
<i>Piper marginatum</i> Jacq.	<i>Artanthe caudata</i> (Vahl) Miq., <i>Piper catalpifolium</i> Kunth, <i>P. marginatum</i> var. <i>anisatum</i> (Kunth) C.DC., <i>P.</i> <i>marginatum</i> var. <i>catalpifolium</i> (Kunth) C.DC.	35
<i>Piper nudilimbium</i> C. DC.		3
<i>Piper reticulatum</i> L.		2
<b>Polygalaceae</b>		
<i>Caamembeca spectabilis</i> (DC.) J. F. B. Pastore	<i>Caamembeca spectabilis</i> (DC.) J.F.B.Pastore var. <i>spectabilis</i> , <i>Polygala spectabilis</i> DC., <i>P. spectabilis</i> DC. var. <i>spectabilis</i>	9
<b>Polypodiaceae</b>		
<i>Microgramma percussa</i> (Cav.) de la Sota	<i>Pleopeltis percussa</i> (Cav.) Hook & Grev., <i>Polypodium</i> <i>percussum</i> Cav.	3
<i>Phlebodium decumanum</i> (Willd.) J.Sm.	<i>Polypodium decumanum</i> Willd.	33
<b>Primulaceae</b>		
<i>Clavija weberbaueri</i> Mez		3
<b>Rosaceae</b>		
<i>Prunus myrtifolia</i> (L.) Urb.	<i>Celastrus myrtifolius</i> L., <i>Prunus sellowii</i> Koehne, <i>P.</i> <i>sphaerocarpa</i> Sw.	8
<b>Rubiaceae</b>		
<i>Hamelia patens</i> Jacq.	<i>Duhamelia odorata</i> Willd. ex Schult., <i>D. patens</i> (Jacq.) Pers., <i>D. sphaerocarpa</i> (Ruiz & Pav.) Pers., <i>Hamelia</i> <i>brachystemon</i> Wernham, <i>H. brittoniana</i> Wernham, <i>H.</i> <i>coccinea</i> Sw., <i>H. corymbosa</i> Sessé & Moc., <i>H. erecta</i> Jacq., <i>H. intermedia</i> Urb. & Ekman, <i>H. lanuginosa</i> M.Martens & Galeotti, <i>H. latifolia</i> Rchb. ex DC., <i>H. nodosa</i> M.Martens & Galeotti, <i>H. patens</i> var. <i>axillaroides</i> Wernham, <i>H. patens</i> var. <i>glabra</i> Oerst., <i>H. patens</i> var. <i>quinifolia</i> DC., <i>Hamelia patens</i> Jacq. var. <i>patens</i> , <i>H.</i> <i>pedicellata</i> Wernham, <i>H. sphaerocarpa</i> Ruiz & Pav., <i>H.</i> <i>suaveolens</i> Kunth, <i>H. tubiflora</i> Wernham, <i>H. verticillata</i> Moc. & Sessé ex DC., <i>H. viridifolia</i> Wernham	54
<i>Palicourea justiciifolia</i> (Rudge) Delprete & J.H.Kirkbr	<i>Cephaelis justiciifolia</i> Rudge, <i>Palicourea lupulina</i> (Benth.) Borhidi, <i>Psychotria flavovirens</i> Suess., <i>P. langsdorffiana</i> Müll.Arg., <i>P. leucophaea</i> Poepp., <i>P. lupulina</i> Benth., <i>P.</i> <i>lupulina</i> subsp. <i>rhodoleuca</i> (Müll.Arg.) Steyer., <i>P.</i> <i>lupulina</i> var. <i>genuina</i> Müll.Arg., <i>P. lupulina</i> var. <i>maypurensis</i> (Humb. & Bonpl. ex Schult.) Steyer., <i>P.</i> <i>lupulina</i> var. <i>stipulacea</i> Müll.Arg., <i>P. maypurensis</i> Humb. & Bonpl. ex Schult., <i>P. persimilis</i> Müll.Arg., <i>P. rhodoleuca</i> Müll.Arg., <i>P. rhodophylla</i> Standl., <i>P. rudgei</i> Bremek.	2
<i>Palicourea racemosa</i> (Aubl.) G.Nicholson	<i>Nonatelia panamensis</i> DC., <i>N. racemosa</i> Aubl., <i>Oribasia</i> <i>racemosa</i> (Aubl.) J.F.Gmel., <i>Palicourea racemosa</i> (Aubl.) Borhidi, <i>Psychotria longistipula</i> Benth., <i>P. quinquecupis</i>	1

	Müll.Arg., <i>P. racemosa</i> (Aubl.) Raeusch., <i>Uragoga racemosa</i> (Aubl.) Kuntze	
<b>Rutaceae</b>		
<i>Ertela trifolia</i> (L.)Kuntze	<i>Monniera trifolia</i> L., <i>Moniera trifolia</i> L., <i>Monniera trifolia</i> Loefl.	12
<i>Zanthoxylum rhoifolium</i> Lam.	<i>Fagara acutifolia</i> (Engl.) Engl., <i>F. astrigera</i> R.S.Cowan, <i>F. obscura</i> (Engl.) Engl., <i>F. pubescens</i> (A.St.-Hil. & Tul.) Herzog, <i>F. regnelliana</i> (Engl.) Chodat & Hassl., <i>F. rhoifolia</i> (Lam.) Engl., <i>F. rhoifolia</i> var. <i>intermedia</i> R.S.Cowan & L.B.Sm., <i>F. rhoifolia</i> var. <i>peltophorum</i> (Turcz.) Chodat & Hassl., <i>F. rhoifolia</i> var. <i>petiolulata</i> (Engl.) Chodat & Hassl., <i>F. rhoifolia</i> var. <i>surparanaensis</i> Najera, <i>F. ruiziana</i> (Klotzsch ex Engl.) Engl., <i>Langsdorfia instrumentaria</i> Leandro, <i>Pohlana instrumentaria</i> (Leandro) Nees & Mart. ex Engl., <i>P. langsdorfii</i> Nees & Mart., <i>Zanthoxylum acutifolium</i> Engl., <i>Z. acutifolium</i> var. <i>petiolulatum</i> Engl., <i>Z. astrigerum</i> (R.S.Cowan) P.G.Waterman, <i>Z. langsdorfii</i> (Nees & Mart.) A.St.-Hil., <i>Z. obscurum</i> Engl., <i>Z. obscurum</i> var. <i>ruizianum</i> Klotzsch ex Engl., <i>Z. peltophorum</i> Turcz., <i>Z. perrottetii</i> DC., <i>Z. pubescens</i> A.St.-Hil. & Tul., <i>Z. regnellianum</i> Engl., <i>Z. rhoifolium</i> var. <i>formosanum</i> (Lillo) P.G. Waterman, <i>Z. rhoifolium</i> var. <i>peltophorum</i> (Turcz.) Engl., <i>Z. rhoifolium</i> var. <i>petiolulatum</i> Engl., <i>Z. rhoifolium</i> var. <i>pubescens</i> (A.St.-Hil. & Tul.) Engl., <i>Z. rhoifolium</i> var. <i>sessilifolium</i> Engl., <i>Z. rhoifolium</i> var. <i>surparanaense</i> (Najera) P.G.Waterman, <i>Z. ruizianum</i> (Klotzsch ex Engl.) J.F.Macbr., <i>Z. sorbifolium</i> A.St.-Hil.	30
<b>Salicaceae</b>		
<i>Casearia javitensis</i> Kunth		2
<i>Casearia sylvestris</i> Sw.	<i>Anavinga samyda</i> C.F. Gaertn., <i>Casearia affinis</i> Gardner, <i>C. attenuata</i> Rusby, <i>C. benthamiana</i> Miq., <i>C. carpinifolia</i> Benth., <i>C. caudata</i> Uittien, <i>C. chlorophoroidea</i> Rusby, <i>C. ekmanii</i> Sleumer, <i>C. formosa</i> Urb., <i>C. herbert-smithii</i> Rusby, <i>C. lindeniana</i> Urb., <i>C. lingua</i> Cambess., <i>C. onacaensis</i> Rusby, <i>C. ovoidea</i> Sleumer, <i>C. parviflora</i> var. <i>microphylla</i> Schltld., <i>C. punctata</i> Spreng., <i>C. samyda</i> (C.F. Gaertn.) A. DC., <i>C. schulziana</i> O.C. Schmidt, <i>C. serrulata</i> Sw., <i>C. subsessiliflora</i> Lundell, <i>C. sylvestris</i> Sw. var. <i>sylvestris</i> , <i>C. sylvestris</i> var. <i>angustifolia</i> Uittien, <i>C. sylvestris</i> var. <i>benthamiana</i> (Miq.) Uittien, <i>C. sylvestris</i> var. <i>carpinifolia</i> (Benth.) Briq., <i>C. sylvestris</i> var. <i>chlorophoroidea</i> (Rusby) Sleumer, <i>C. sylvestris</i> var. <i>eichleri</i> Briq., <i>C. sylvestris</i> var. <i>lingua</i> (Cambess.) Eichler, <i>C. sylvestris</i> var. <i>myricoides</i> Griseb., <i>C. sylvestris</i> var. <i>paraensis</i> Uittien, <i>C. sylvestris</i> var. <i>platyphylla</i> A. DC., <i>C. sylvestris</i> var. <i>tomentella</i> Rusby, <i>C. sylvestris</i> var. <i>wydleri</i> Briq., <i>Guidonia sylvestris</i> (Sw.) M. Gómez, <i>Samyda sylvestris</i> (Sw.) Poir.	48
<i>Prockia crucis</i> P. Browne ex L.	<i>Prockia septemnervia</i> Spreng.	1
<b>Smilacaceae</b>		
<i>Smilax longifolia</i> Rich.	<i>Sarsaparilla acuminata</i> (Willd.) Kuntze, <i>Smilax grandifolia</i> Regel, <i>S. papyracea</i> Duhamel	10
<b>Solanaceae</b>		
<i>Brunfelsia grandiflora</i> D.Don		39
<b>Violaceae</b>		
<i>Leonia glycyarpa</i> Ruiz & Pav.	<i>Clavija sparsifolia</i> Miq., <i>Leonia glycyarpa</i> Ruiz & Pav. var. <i>glycyarpa</i> , <i>L. glycyarpa</i> var. <i>racemosa</i> (Mart.) L.B.Sm. & A.Fernández, <i>L. racemosa</i> Mart.	3
<b>Zamiaceae</b>		
<i>Zamia ulei</i> Dammer	<i>Zamia cupatiensis</i> Ducke	2

We found 1535 records of medicinal services for the 83 species. The major of medicinal services (77%) were mentioned only once, while 378 (24%) medicinal uses were mentioned in more than one study. This means that 38 species had medicinal uses reported more than once in the literature, while 45 plant species had their medicinal services reported only once. The Huni Kuí people had 226 medicinal services (90%) that were mentioned only once in the literature for this people.

Only 20 plants had records in past studies of medicinal uses similar to those of the Huni Kuí people (Table 2), meaning that only 10% of the Huni Kuí traditional knowledge is shared with other cultures, while 90% of this knowledge is unique to their culture. Were 61 references that reported medicinal uses similar to those of the Huni Kuí people, with most of the studies carried out in Brazil, but also in other countries, such as Argentina, Colombia, Costa Rica, Ecuador, Honduras, Mauritius, India, Mexico, Nicaragua, Panama, Pakistan, Peru, Suriname and Venezuela.

Table 2. Plant species with medicinal uses similar to those of the Huni Kuí people.

Scientific Name	Medicinal uses	References
<i>Brunfelsia grandiflora</i>	Muscle pain	Desmarchelier & Schaus 2000, Duke & Vasquez 1994, Giovannini 2015, Jernigan 2011, Jernigan 2012, Kvist et al. 2001, Mejía & Rengifo 2000, Polesna et al. 2011, Pratt 2007, Rengifo-Salgado et al. 2017, Rutter 1990, Sanz-Biset & Cañigüeral 2011, Sanz-Biset et al. 2009, Schultes et al. 1998, Stasi & Hiruma-Lima 2002, Torre et al. 2008
<i>Annona ambotay</i>	Energy level	Milliken et al. 1999
<i>Caamembeca spectabilis</i>	Animal or human bite	Agra et al. 2008
<i>Caamembeca spectabilis</i>	Vaginal discharge	Coelho-Ferreira 2009
<i>Cordia nodosa</i>	Animal or human bite	Saltos et al. 2016
<i>Couma macrocarpa</i>	Animal or human bite	Zapata 2015
<i>Cyclanthus bipartitus</i>	Fever	Valadeau et al. 2010
<i>Fittonia albivenis</i>	Pain, face	Vickers & Plowman 1984
<i>Fittonia albivenis</i>	Muscle pain	Vickers & Plowman 1984
<i>Handroanthus serratifolius</i>	Health promotion related to growth, development and ageing	Ruysschaert et al. 2009
<i>Lygodium venustum</i>	Animal or human bite	Butt et al. 2015, Coe 2008, Hamayun et al. 2006, Hernandez et al. 2007, Upreti et al. 2009, Vásquez et al. 2015
<i>Lygodium venustum</i>	Multiple trauma and injuries	Gupta et al. 1986
<i>Myroxylon balsamum</i>	Animal or human bite	Piso & Marcgrave 1648, Silva 2017
<i>Myroxylon balsamum</i>	Pain, face	Bussmann et al. 2010, Iglesias 1987, Suroowan & Mahomoodally 2016, Vergara et al. 2018
<i>Palicourea justiciifolia</i>	Skin colour change	Elisabetsky & Posey 1994
<i>Petiveria alliacea</i>	Other specified breathing problems	Carvalho 2015
<i>Phlebodium decumanum</i>	Cough	Amorozo & Gély 1988, Brack 1999, Carbonó-Delahoz & Dib-Diazgranados 2013, Gupta 1995, Milliken & Albert 1996, Milliken et al. 1999, Silva 2019
<i>Phlebodium decumanum</i>	Pertussis	Cibrián & Sutherland 2007
<i>Piper callosum</i>	Pain, face	Carvalho 2019, Cassino 2010
<i>Piper marginatum</i>	Pain, face	Hazlett 1986
<i>Prockia crucis</i>	Other specified and unknown infectious diseases	Tournon et al. 2015
<i>Prunus myrtifolia</i>	Malaria	Milliken 1997
<i>Tabernaemontana sananho</i>	Animal or human bite	Sanz-Biset et al. 2009



<i>Tabernaemontana sananho</i>	Hunt	Bennett & Alarcón 2015, Luzuriaga-Quichimbo et al. 2018
<i>Trema micrantha</i>	Pain, face	Crovetto 2012
<i>Zanthoxylum rhoifolium</i>	Fever	Agra et al. 2008, Cavalheiro & Guarim-Neto 2018, Figueiró-Leandro & Citadini-Zanette 2008, Messias et al. 2015, Thomas 2001

*Brunfelsia grandiflora* was the plant with more mentions in the literature (16 in total) of the same use by the Huni Kuĩ, in the category of Muscle pain. *Phlebodium decumanum* had seven mentions in the Cough category, *Lygodium venustum* had six uses for Animal or human bite, *Zanthoxylum rhoifolium* had five uses for Fever, *Myroxylon balsamum* had four uses for Pain, face, and *Tabernaemontana sananho*, *Myroxylon balsamum* and *Piper callosum* had two uses for Hunt, Animal or human bite and Pain, face.

*Annona ambotay* had a record of use for Energy level, *Caamembeca spectabilis* for Vaginal discharge. *Caamembeca spectabilis*, *Cordia nodosa*, *Couma macrocarpa* and *Tabernaemontana sananho* had records for Animal or human bite, *Cyclanthus bipartitus* for Fever, *Fittonia albivenis* for Muscle pain. *Fittonia albivenis*, *Piper marginatum* and *Trema micrantha* had records of use for Pain, face, *Handroanthus serratifolius* for Health promotion related to growth, development and ageing. *Lygodium venustum* had records for Multiple trauma and injuries, *Palicourea justiciifolia* for Skin colour change, *Petiveria alliacea* for Other specified breathing problems, *Phlebodium decumanum* for Pertussis. *Prockia crucis* for Other specified and unknown infectious diseases and *Prunus myrtifolia* had one record of use to treat Malaria.

In total, 28 species presented 165 medicinal services previously recorded in the literature for more than 30 years. Among these, six species had seven old medicinal services similar to those used by the Huni Kuĩ people. *Myroxylon balsamum* had records for two medicinal services, Pain, face and Animal or human bite; while five other species had records of one old medicinal service similar to the one used by the Huni Kuĩ: *Brunfelsia grandiflora* is used for Muscle pain; *Fittonia albivenis* for Pain, face; *Lygodium venustum* for Multiple trauma and injuries; *Phlebodium decumanum* for Cou et al. gh; and *Piper marginatum* for Pain, face.

Other 16 plant species had historical records of their medicinal use (Table 3). *C. sylvestris*, *P. marginatum* and *T. micrantha* are used by the Guaraní people (Pereira et al. 2016); *Ertela trifolia* and *Microgramma percussa* had their medicinal use in the amazon reported in the XIX century by the naturalist Martius (Breitbach et al. 2013), *Myroxylon balsamum* have several records of its medicinal use from the XVII, XIX and XX centuries (Janni & Bastien 2000, Mackonochie & Heinrich 2019, Martins et al. 2019, Ricardo et al. 2017, Silva 2017). The medicinal and spiritual use of *Petiveria alliacea* in Brazil and Argentina is recorded since the XIX century (Magalhães et al. 2019, Pereira et al. 2016, Ricardo et al. 2017, Scarpa & Anconatani 2017, Scarpa et al. 2016, Scarpa & Rosso 2019). *Smilax longifolia* had its medicinal use in the Amazon and in the Caatinga reported in the XIX and XX centuries (Alencar et al. 2010, Breitbach et al. 2013, Magalhães et al. 2019). *Vismia guianensis* has records of medicinal use in Brazil from the XIX century (Ricardo et al. 2017, Santos-Fonseca et al. 2019), while *Zanthoxylum rhoifolium* has records of historical medicinal use in Argentina (Scarpa & Anconatani 2017) and Brazil (Magalhães et al. 2019, Pereira et al. 2016).

Table 3. Plant species with historical records of their medicinal use.

Scientific Name	Historical medicinal use	References
<i>Microgramma percussa</i>	medicinal use in the amazon reported in the XIX century by the naturalist Martius	Breitbach et al. 2013
<i>Trema micrantha</i>	used by the Guaraní people	Pereira et al. 2016
<i>Piper marginatum</i>	used by the Guaraní people	Pereira et al. 2016
<i>Casearia sylvestris</i>	used by the Guaraní people	Pereira et al. 2016
<i>Ertela trifolia</i>	medicinal use in the amazon reported in the XIX century by the naturalist Martius	Breitbach et al. 2013
<i>Myroxylon balsamum</i>	medicinal use in the XVII, XIX and XX centuries	Janni & Bastien 2000, Mackonochie & Heinrich 2019, Martins et al. 2019, Ricardo et al. 2017, Silva 2017
<i>Petiveria alliacea</i>	historical medicinal and spiritual use in Brazil and Argentina in the XIX century	Magalhães et al. 2019, Pereira et al. 2016, Ricardo et al. 2017, Scarpa & Anconatani 2017, Scarpa et al. 2016, Scarpa & Rosso 2019

<i>Smilax longifolia</i>	medicinal use in the Amazon and in the Caatinga reported in the XIX and XX centuries	Alencar et al. 2010, Breitbach et al. 2013, Magalhães et al. 2019
<i>Vismia guianensis</i>	medicinal use in Brazil in the XIX century	Ricardo et al. 2017, Santos-Fonseca et al. 2019
<i>Zanthoxylum rhoifolium</i>	historical medicinal use in Argentina and Brazil	Scarpa & Anconatani 2017, Magalhães et al. 2019, Pereira et al. 2016

In total, 32 plant species were reported in 295 references for Brazil, while 52 species were reported in 315 references for other countries. Peru, Colombia, Mexico, Ecuador and Argentina were the countries with more studies, while Peru, Colombia, Ecuador, Suriname and Bolivia had more species recorded as medicinal (Figure 1). Amazon, Atlantic Forest and Caatinga, shared many medicinal species with other biomes (45, 11 and 11 shared species, respectively). Most of the research was done in the Amazon (175), Atlantic Forest (75), Cerrado (46) or Caatinga (31) (Figure 2).

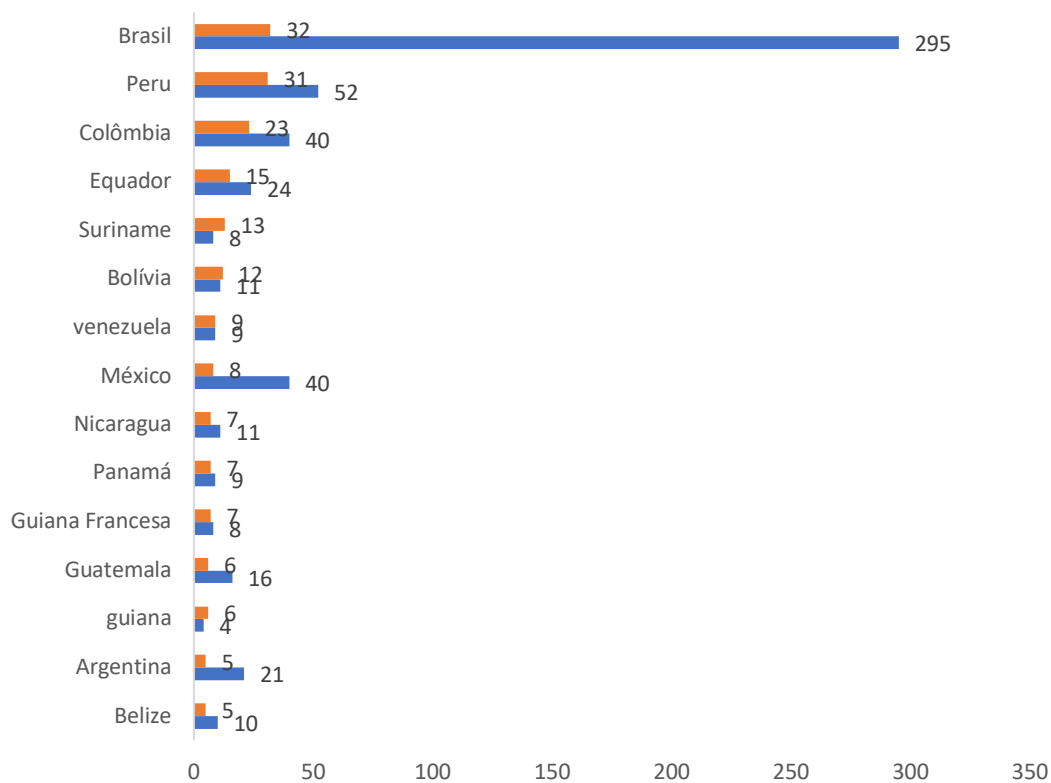


Figure 1. Number of references found on consulted databases for 83 medicinal plants used by the Huni Kui people, divided by country where research took place. Databases used: Scielo, PubMed and Google Scholar.

## Discussion

Our results have relevant numbers of references and highlight the importance of considering valid botanical names and its synonyms in literature reviews. We also point that part of the literature about medicinal uses of plants could not be considered as they did not have their medicinal use specified, emphasizing how much the traditional knowledge of indigenous people from the Amazon is still under sampled (Cámara-Leret *et al.* 2014b). Besides that, our results suggest that for 13 species used as medicinal by the Huni Kuĩ people and with no former record of medical use, this study is the first formal record of these medicinal uses in the literature (Muru & Quinet 2014), although this could be because of our scientific knowledge gaps or any search bias in this review. In this way, any further analysis about these species should be done with care.

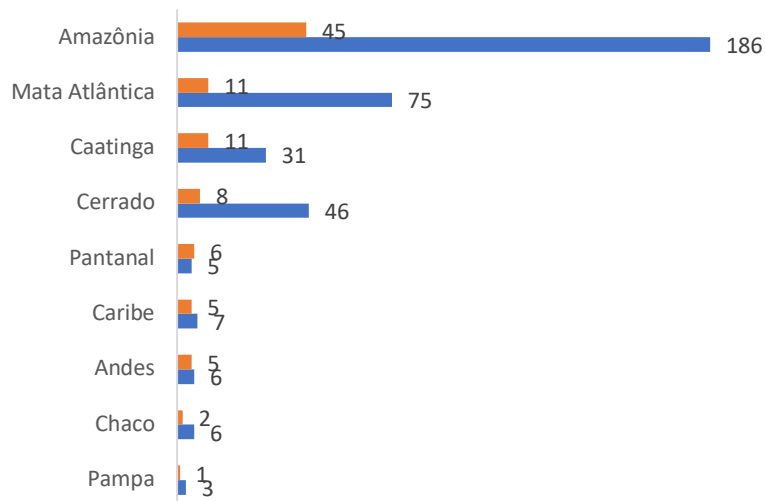


Figure 2. Number of references found on consulted databases for 83 medicinal plants used by the Huni Kui people, divided by region where research took place. Databases used: Scielo, PubMed and Google Scholar.

Many of the medicinal services compiled in this study are considered unique, even those of widespread species, and were recorded only by the Huni Kuĩ people. This result is similar to those reported in the literature (Cámara-Leret & Bascompte 2021), or when compared with nearby populations (Junsongduang *et al.* 2014) and the reason behind it is the cultural singularity of the Huni Kuĩ knowledge regarding the use of medicinal plants.

It is fundamental to improve our knowledge about the geographic distribution of traditional knowledge, especially in areas with high biocultural diversity, if we want to propose protocols for a fair division of the benefits and profits resulting from the exploration of genetic resources (Cámara-Leret *et al.* 2014) and this is also true for the traditional knowledge of the Huni Kuĩ people. We found a rate of 90% of singularity of the Huni Kuĩ's traditional knowledge, a similar rate to other northwestern Amazon people's traditional knowledge singularity (Cámara-Leret & Bascompte 2021), reinforcing the notion that the Americas are a hotspot of indigenous traditional knowledge and, therefore, a priority area for new documentation efforts (Cámara-Leret & Bascompte 2021).

Even with its high singularity, the medicinal services of plant species used by the Huni Kuĩ have similarities with other cultures, which may represent some level of "tradition" in the use of these species. The records of certain use through time (Ricardo *et al.* 2018), at least by two generations or more (Shils 1981), can be used to conceptualize the term "tradition" for medicinal use of plants and the use of historical documents could be a successful strategy to improve our knowledge about these traditions (Pires *et al.* 2020). Another strategy is the "social validation" (Crellin 2008), a concept in which more ethnobotanical records of the same medicinal service of a certain plant makes the claim of its "tradition" stronger. To make it easier, we can adopt quantitative criteria to set the time frame to consider the use of a plant "traditional", similar to the criteria adopted by the European Union (Knoess & Wiesner 2019). Nevertheless, the adoption of a single time frame without other criteria and associated data can be vague and frivolous to truly attest the tradition of a medicinal use of a plant (Jütte *et al.* 2017) and although it is paramount to indicate the tradition in the use of a plant, it is necessary to consider all available evidence, positive or negative, in the evaluation, especially if our goal is to subsidize governmental policies (Jütte *et al.* 2017).

Muru & Quinet (2014) stated that the traditional knowledge about medicinal plants was mainly recorded by the older shamans. In this context, we can conclude that the recorded knowledge spans at least two generations, but looking at the tradition, the medicinal services of some taxa shared among other indigenous people needs to be highlighted, especially *Brunfelsia grandiflora*, used for Muscle pain, *Phlebodium decumanum*, for Cough, *Lygodium venustum*, for Animal or human bite, *Zanthoxylum rhoifolium* for Fever and, *Myroxylon balsamum* for Pain, face. Besides the common use, that can confer "social validation" to these "old" services, as some of these records were published more than 30 years ago and because of it they should be considered strategic in governmental policies of primary and basic healthcare in the Amazon. This perspective can include policies of social inclusion (Filocreão *et al.* 2017) and equalize health policies within the targeted territory (Pires *et al.* 2020).

The majority of species in this study are widespread and this may explain why we can find many of these plants in other countries and in different vegetation types. Our results suggest that even with the low rates of shared medicinal services of these plants, the knowledge about these services is spread among several countries and in different cultural and social contexts, reinforcing the complexity in the distribution patterns of traditional knowledge (Cámara-Leret *et al.* 2014, Vandebroek 2010), the importance non-indigenous cultures also have in the preservation of the traditional knowledge and the need for cooperation among countries in order to reach the goals of the Nagoya protocol (Cámara-Leret *et al.* 2014).

The predominance of the Amazon (in Brazil and other countries) regarding the medicinal services, can be a consequence of a geographic distribution bias of the species group, as they are Amazonian native plants and in some cases they are also endemic. Although the traditional knowledge of South America is still under-documented (Cámara-Leret *et al.* 2014b), it is possible that these results are related to historical research efforts in other regions and for some indigenous people. Therefore, this literature review corroborates the idea of a singularity pattern of medicinal knowledge (Cámara-Leret & Bascompte 2021, Junsongduang *et al.* 2014) and indicate the necessity to study the under-documented traditional knowledge (Cámara-Leret *et al.* 2014), because each indigenous people can have unique knowledge that, in face of the global climate emergency, are fundamental to devise new mitigation, adaptation and sustainable development policies (Leonard *et al.* 2013, Salgotra *et al.* 2018).

## Conclusions

Our results show how important are the projects documenting the Amazonian indigenous traditional knowledge, as they can safeguard unique knowledge about natural resources. The Huni Kuĩ traditional knowledge about medicinal plants is rich and singular, as other indigenous people traditional knowledge, and the proclamation of the International Decade of Indigenous Languages by the UN shows that preservation of indigenous languages is paramount, if we want to keep this knowledge and to achieve the goals of conservation and sustainable development proposed in the Nagoya Protocol and Biological Diversity Convention.

The cross-border analysis of the knowledge shows how rich, and complex is the distribution of traditional knowledge, making it clear the importance of studies that focus on the development of mechanisms and policies for sharing the benefits and profits resulting from the exploration of genetic resources. Beyond that, the evidence that informs about the traditionality and shared use among different cultures can help the development of new strategies to incorporate plant species in primary healthcare, improving the life quality of local communities.

## Declarations

**List of abbreviations:** International Classifications of Primary Care (ICPC).

**Ethics approval and consent to participate:** Not applicable.

**Consent for publication:** Not applicable.

**Availability of data and materials:** The data was not deposited in public repositories.

**Competing interests:** The authors do not have any competing interests.

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**Authors' contributions:** The first author, Thiago Serrano de Almeida Penedo, contributed to research design, carried out data analysis and to manuscript writing. While Ariane Luna Peixoto, Alexandre Quinet and Moacir Haverroth contributed to research design. All authors have read and agreed to the published version of the manuscript.

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## Supplementary material. Botanical family, plant species and all of references for each species.

Botanical family	Scientific names	References
ACANTHACEAE	<i>Fittonia albivenis</i> (Lindl. ex Veitch) Brummitt	Sanz-Biset J, Campos-de-la-Cruz J, Epiqui�n-Riverac MC, Canigual S. 2009. A first survey on the medicinal plants of the Chazuta valley (Peruvian Amazon). <i>Journal of Ethnopharmacology</i> 122:333–362
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