



# Ethnobotanical and ethnopharmacological study of medicinal plants used in treating some liver diseases in the Al-Haouz Rehamna region (Morocco)

Ouafae Benkhnigue, Hamid Khamar, Rainer W. Bussmann, Noureddine Chaachouay and Lahcen Zidane

## Correspondence

**Ouafae Benkhnigue<sup>\*1</sup>, Hamid Khamar<sup>1,2</sup>, Rainer W. Bussmann<sup>3,4</sup>, Noureddine Chaachouay<sup>5</sup> and Lahcen Zidane<sup>2</sup>**

<sup>1</sup>Department of Botany and Plant Ecology, Scientific Institute, University Mohammed V, B. P. 703, Rabat 10106, Morocco.

<sup>2</sup>Plant, Animal Productions and Agro-industry Laboratory, Department of Biology, Faculty of Sciences, Ibn Tofail University, B.P. 133 14000, Kenitra, Morocco.

<sup>3</sup>Department of Botany, State Museum of Natural History, Erbprinzenstrasse 13, 76133 Karlsruhe, Germany.

<sup>4</sup>Department of Ethnobotany, Institute of Botany and Bakuriani Alpine Botanical Garden, Ilia State University, 1 Botanical Str., Tbilisi, Georgia.

<sup>5</sup>Agri-Food and Health Laboratory (AFHL), Higher School of Education and Training, University Hassan, 1st, 50 Rue Ibnou Lhaytham B.P. 577, 26002 Settat, Morocco.

\*Corresponding Author: benkhnigue@gmail.com

**Ethnobotany Research and Applications 25:34 (2023)** - <http://dx.doi.org/10.32859/era.25.34.1-32>

Manuscript received: 18/02/2023 – Revised manuscript received: 28/02/2023 – Published: 02/03/2023

## Research

### Abstract

**Background:** Medicinal plants have always been important in therapeutic and preventive folk medical remedies for humans and cattle. Plants are also quite important in today's global economy.

**Objectives:** This study is part of the development of plant resources in the Al-Haouz Rehamna region. Its general objective was to inventory the medicinal plants used in traditional pharmacopeia against certain liver diseases.

**Methods:** Ethnobotanical field surveys were conducted using 1700 questionnaire forms. Ethnobotanical indices such as the informant agreement ratio (IAR), the family use value (FUV), the use-value (UV), and the Plant Part Value (PPV) were employed in the data analysis rate.

**Results:** The findings enabled us to classify 86 medicinal plants into 79 genera and 37 families, among which four predominate: Asteraceae (17), Lamiaceae (9), and Fabaceae (8 species), and Apiaceae (7). Among the reported species, 21 are toxic, and the population of the said region widely uses ten: *Ridolfia segetum* (Guss.) Moris, *Curcuma longa* L., *Ononis natrix* L., *Rhamnus alaternus* L., *Cladanthus arabicus* (L.) Cass, *Rhaponticum acaule* (L.) DC, *Corrigiola telephifolia* Pourr, *Cynara cardunculus* L., *Cicer arietinum* L. and *Aframomum melegueta* K. Schum. The main parts used are leaves (PPV=0.183) and seeds (PPV= 0.165). A decoction is the most used method (34.88%).

The recipes are mainly administered orally and rarely by the cutaneous route as a poultice on the abdomen. The diversity of therapies identified in the study area is a cultural richness.

*Conclusion:* Thus, the data reported by this study could be a precious reference of data for this region and could be a basis for further study in the field of phytochemistry to produce and identify new natural drugs that could be endowed with interesting hepatoprotective properties in the treatment or prevention of certain liver diseases.

*Keywords:* Ethnobotany, ethnopharmacology, traditional medicine, liver disease, hepatoprotective, toxicity, Al-Haouz Rehamna, Morocco.

## **Background**

Liver disorders are a serious global health issue; they are the leading cause of human mortality globally, accounting for roughly two million fatalities each year, one million from cirrhosis complications, and one million from hepatitis viruses and hepatocellular carcinoma (Mokdad *et al.* 2014). Furthermore, these disorders are distinguished by development from steatosis to chronic hepatitis, fibrosis, cirrhosis, and hepatocellular cancer (Notas *et al.* 2009). Several specific causes can be the origin of these conditions: parasitic, obstructive, viral hepatitis, alcohol abuse, drug abuse, metabolic diseases due to iron or copper overload, autoimmune attack of hepatocytes, bile duct epithelium, or congenital anomalies (Michel *et al.* 2009). Patients with the chronic liver disease suffer from fatigue, anxiety, depression, decreased work productivity, and other emotional problems that significantly impair their quality of life (Afendy *et al.* 2009). In addition to a heavy clinical burden, the management of patients represents an economic burden for health care systems. Despite significant advancements in current medicine, no effective medications are available that protect the liver from harm or boost its activity (Akther *et al.* 2013). In recent decades, herbal medicine has played a crucial role in primary health care worldwide; it can serve as a curative or preventive drug regarding safety and efficacy. Thus, the WHO encourages using herbal medicine to treat several diseases, including liver diseases.

In Morocco, thanks to the richness of its vascular flora, about 5211 species and subspecies (Dobignard and Chatelain 2013; Fennane and Ibn Tattou 2012), the over-the-counter access, and the possibility of self-medication, the Moroccans have a long and illustrious history in herbal remedies (Bellakhdar 1997). They often use medicinal herbs to treat various disorders, especially liver problems. In the Al-Haouz Rehamna region, phytotherapy remains integral to the local Moroccan culture. Thus, using medicinal plants is a matter of common tradition for most of this population. Moreover, the Moroccan medicinal reference study reveals that investigations on medicinal plants are of relative relevance in Morocco's health system. Indeed, various works have been published in the last decades on Moroccan ethnobotanical knowledge. But works related to medicinal plants' inventory against liver diseases are rare in Morocco. Thus, the requirement to conduct an ethnobotanical investigation in phytotherapy contributes to the search for other traditional medicines of natural origin that could be endowed with impressive therapeutic effects (anti-hepatoprotective, anti-inflammatory, and antioxidant) in the therapy or precluding of these diseases. Given these factors, the current preliminary study was undertaken to catalog the traditionally used plants in treating some hepatic affections (jaundice, hepatitis, and liver cancer), which seem to be more frequent in the local population of the said region.

## **Material and Methods**

### **Description of the study area.**

Due to its geographical position and climate, Morocco can be subdivided into different phytogeographical areas (Fennane & Ibn Tattou 1998, 2005; Ibn Tattou & Fennane 2008). The Al-Haouz Rehamna region belongs to the Middle Atlantic Morocco group. It is bounded to the north by the Chaouia-Doukkala region, to the northeast by the Middle Oum-Errabiâ region, to the east by the Mgoun region, to the south by Ida-Ou-Tanane, Seksoua and the Central High Atlas and to the west by the Abda-Haha region (Fig. 1).

The study area is characterized by four major natural geographic areas (Piqué *et al.* 1993): Plateau areas with a moderate altitude below 1000m, including the plateaus of Rehamna and Bahira; Plain areas, including the plains of Al-Haouz Rehamna, and Tassaout Upstream and Downstream. This region contains large agricultural areas: the Basin area, i.e. the Essaouira-Chichaoua basin, characterized by depressions and elevations in the shape of cereal-producing land or rangeland and the Djebilets, a mountainous region of medium height with minimal vegetation.

Administratively, the study area is located within the Marrakech-Safi region. This region has a total area of 39 167 km<sup>2</sup> or 5.51 % of the national land (MGRMS 2015). According to the most recent general census of population and

housing, the estimated population of the Marrakech-Safi area is 4,520,569 (4,511,933 Moroccans and 8,636 foreigners; 13.36% of the national population and 115 persons per km<sup>2</sup>). With a proportion of 57.12%, the bulk of its population (2 582 553) is rural. The high rural population rate demonstrates the country's agricultural commitment (HCP 2014).

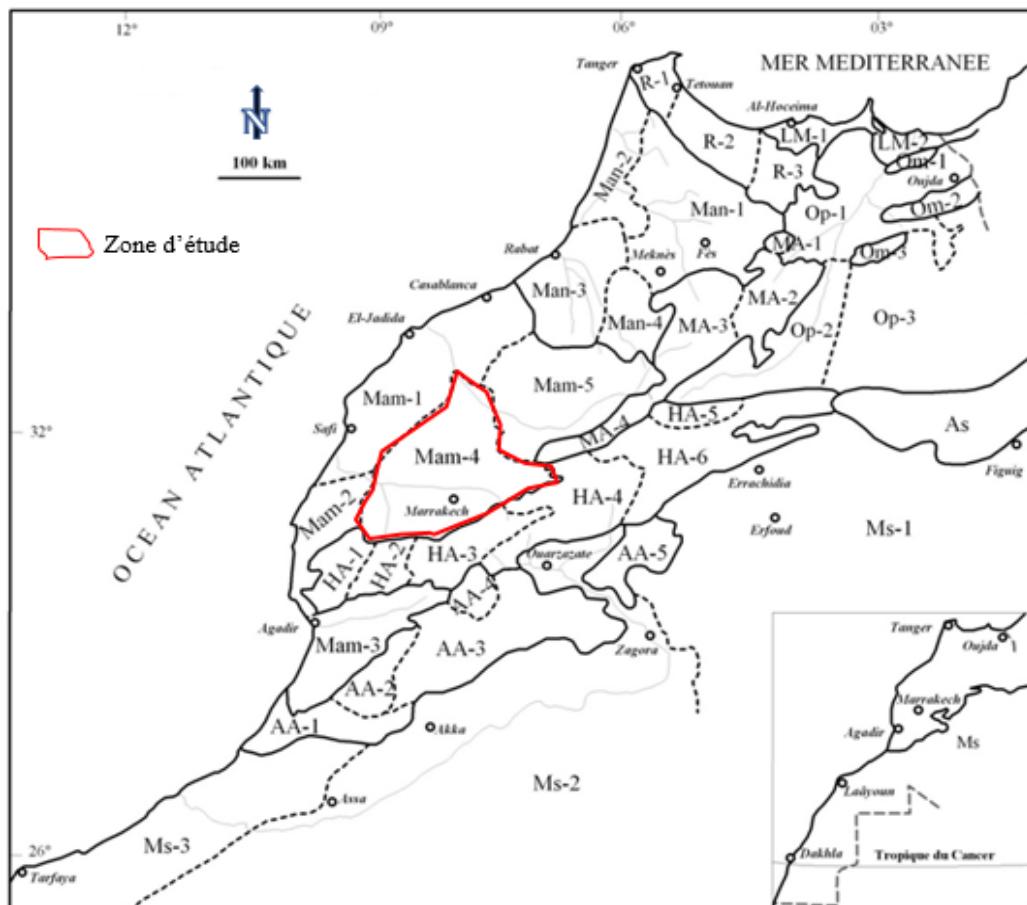


Figure 1. Carte Map of phytogeographic divisions of northern Morocco. (from Fenanne & Ibn Tattou 1998, 2005).

Soil studies conducted throughout the study area (Billaux and Bayssine 1967; Cavallar 1950) have shown the presence of eight types of soils: isohumic, red on shale, calco-magnesimorphic, vertisols (Tirs), halomorphic, hydromorphic, regosols, and soils not significantly evolved erosions, their texture is generally silty-clayey, silty and silty-sandy.

The area gets average annual precipitation of 800 millimeters in the hilly region and 190 millimeters on the plain (Ait El Mekki 2017; Knippertz *et al.* 2003). The yearly average temperature is around 18.5°C. The average maximum temperature is 37.7 degrees Celsius, while the average lowest temperature is 4.9 degrees Celsius (Samia *et al.* 2018). The area generally sits between arid and temperate winter bioclimates and semi-arid and mild winter bioclimates (Negre 1959).

The region's hydrographic system contains a vast watershed, the Tensift, and a portion of the Oum Errabia watershed, which is formed by multiple sub-watersheds and drains the northern slope of the High Atlas, with pluvial inputs (Abourida 2007).

From the point of view of the altitudinal zonation of ecosystems, two vegetation stages are defined in the study area: the thermo-Mediterranean stage (individualized in the north and south of the region) and the sub-Mediterranean stage, which is individualized in the center of the study area (Benabid 2000). Thus, the vegetation is classified in the arid Mediterranean vegetation stage; in Haouz-Tadla and Rehamna, it is represented by a bush with *Ziziphus lotus* (L.) Lam, *Withania frutescens* (L.) Pauquy and *Acacia gummifera* Willd; *Pistacia atlantica* Desf. here, it is scarce. However, in the Djebilets chain that forms the top of this whole arid zone of Western Morocco, the vegetation cover is generally precarious and appears highly reduced on the slopes. A few stunted bushes of

*Acacia gummifera* and *Ziziphus lotus*, very scattered here and there, a *Withania frutescens* or an *Ephedra*, are the only representatives of the shrubby vegetation that we have encountered (Emberger 1938; Negre 1959).

Thus, the region of Al-Haouz Rehamna is characterized by a diversity of hydro-geographical, climatic, and Phylogenetic resources. This geographical, structural, and climatic diversity is reflected by a specific richness of spontaneous plants, incredibly aromatic and medicinal.

### Ethnobotanical Methodology

#### Survey

To establish the list of plants used in traditional herbal medicine against liver diseases in the Al-Haouz Rehamna region, we conducted ethnobotanical field surveys during five campaigns from 2012 to 2017 to document plants employed in herbal medicine against liver diseases in the Al-Haouz Rehamna region, using 1700 questionnaires with herbalists and traditional practitioners and the local population. The interviews were conducted in an open discussion format so that people could respond without constraint and were conducted individually or in groups. All interviews were conducted after obtaining oral prior informed consent. The time devoted to each interview was approximately 15-120 minutes, depending on the availability of the participants. The questionnaires were previously prepared on a survey form (Appendix I). The data collected was recorded in this form, inspired by previous work (Bellakhdar 1997; Benkhnigue *et al.* 2010) and the questionnaires were adopted according to our study. Using stratified probabilistic sampling (Godron and Daget, 1982; Godron 1971; Benkhnigue *et al.* 2022), the locations of the different environments of ethnobotanical surveys and floristic surveys were determined in order to have the most comprehensive floristic inventory possible and to conduct ethnobotanical surveys in the studied region. The research region was subdivided into 34 identical strata (Fig. 2, Table 1). Using simple random selection, 50-person samples were drawn from each of the 34 strata.

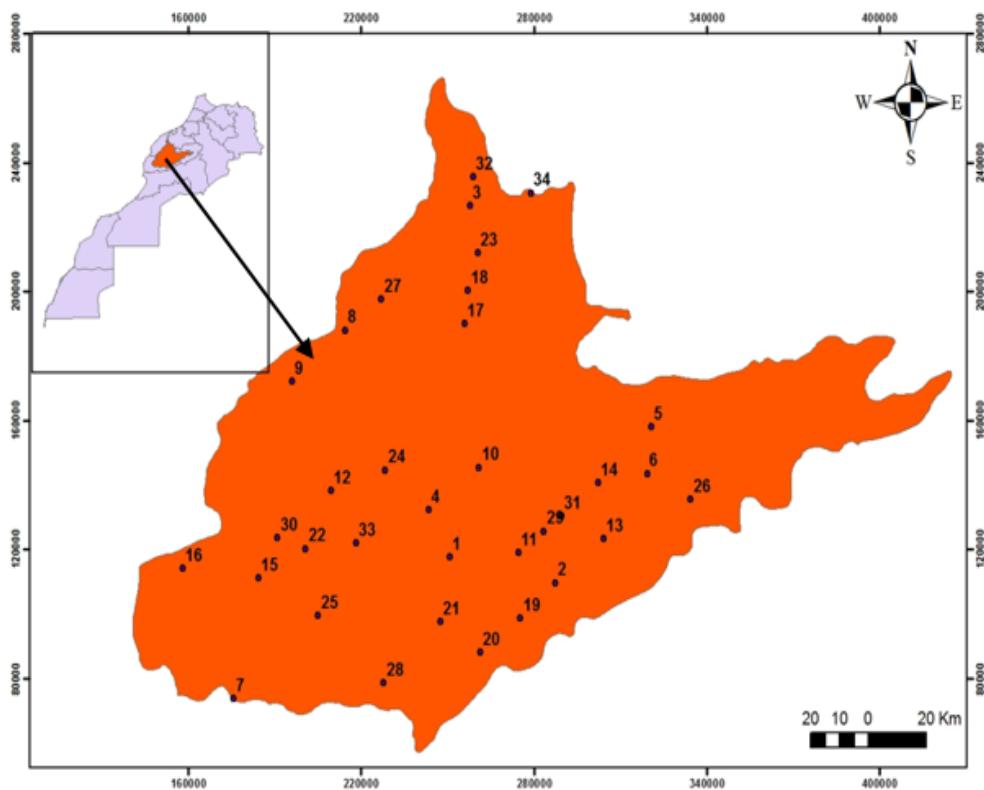


Figure 2. Map of the study area showing surveyed stations (Benkhnigue *et al.*, 2022).

#### Collection and identification

To identify each plant used, we organized plant sampling missions in natural environments from March 1, 2012, to September 30, 2017. Identification of the collected plants and those observed was based on the main floras (Bellakhdar 1997; Fennane *et al.* 1999; 2007; 2014; Fennane and Ibn Tattou 1998; Hmamouchi, 1999; Negre, 1961; 1962), and samples checked herbarium (RAB), as well as via the Jstore (<http://www.plants.jstore.org>), Global Biodiversity Information Facility (<http://www.gbif.org/occurrence>), and The Plant List databases (<http://www.theplantlist.org/>). The nomenclature follows (Dobignard and Chatelain 2013) as well as (Fennane and

Ibn Tattou 2005; Ibn Tattou and Fennane 2008). Plant families follow Angiosperm Phylogeny Group IV (APG 2016). The assessment of plant biological types (life forms) followed Raunkiaer (1934).

Table 1. List of floristic and ethnobotanical survey points.

<b>01-Marrakech</b>	<b>09-Echemmaia</b>	<b>17-Benguerir</b>	<b>25-Majjate</b>	<b>33-Sid Zwine</b>
<b>02-Aït-Ouirir</b>	10-Sidi Bou Othmane	18-Sebt Brikyne	26-Assahrij	34-Mechraa-Ben Aabou
<b>03-Ksiba</b>	11-Ouelad hassoune	19-Jemaat-Ghmate	27-Tnin Bouchane	
<b>04-Tamen sourte</b>	12-Si Thami	20-Tahennaoute	28-Mtal	
<b>05-El-Kelaâ des Sraghna</b>	13-Sidi Rahal	21-Tameslouhete	29-Jaidate	
<b>06-Laataouia</b>	14-Tamellalte	22-Lamzoudia	30-Mguedgua	
<b>07-Imintanoute</b>	15-Chichaoua	23-Skhour rehamna	31-Ras Ain Rehamna	
<b>08-Youssoufia</b>	16-Sid L'Mokhtar	24-Tlat Ouelad-Dlim	32-Sidi Ghanem	

## Statistical analysis and data processing

### Statistical analysis

To extract the different information concerning the four axes of the research: the informant, the plant material, the mode of use and administration, and the treated disease, we processed the survey data using SPSS version 21 (Statistical Package for the Social Sciences, IBM, New York, United States).

### Quantitative indices

To further examine and understand the data, we used quantitative and descriptive analyses using ethnobotanical indices such as:

#### Informant Agreement Ratio (IAR)

The IAR analysis has recently been used as an essential tool for ethnobotanical data analysis (Uniyal *et al.* 2011; Upadhyay *et al.* 2010). It depends on the availability of plants in the study area, and it informed us about the degree of shared knowledge about the medical use of plants among informants to treat specific disease categories. This rate is calculated as follows:

$$IAR = \frac{Nur - Nt}{Nur - 1}$$

where **Nur**: number of user records in each use category and **Nt**: number of use taxa in each use category. The IAR value ranges from 0 to 1; the higher value indicates taxon selection agreement among informants, while the lower value indicates disagreement. The value of 1 indicates that the taxa are used by many informants (Inta *et al.* 2013).

#### Plant Part Value (PPV).

The plant part value (PPV) was calculated using the following formula:

$$PPV = \frac{RU_{plant\ part}}{RU}$$

Where: **RU<sub>plant part</sub>** is the sum of reported uses per plant part, and **RU** is the number of reported uses of all plant parts. The part with the highest PPV is the most used by respondents (Benkhnigue *et al.* 2022; Chaachouay *et al.* 2020).

#### Use Value (UV).

It was calculated as follows:

$$UV = \frac{\sum U_i}{N}$$

where **Ui**: is the number of uses cited by each informant for a given species, and **N**: is the total number of informants (Hudaib *et al.* 2008; Vitalini *et al.* 2013). Their calculation allowed us to determine the relative importance of locally known species and the most frequently reported species in treating disease.

**Family Use Value (FUV).**

The FUV allowed us to identify the importance of medicinal plant families. It was calculated using the following formula:

$$\text{FUV} = \text{UVs/Ns}$$

where **UVs**: use-value of the species, and **Ns**: total number of species within each family (Cadena-González *et al.* 2013). The calculation of FUVs is an index of cultural importance that can be applied in ethnobotany to calculate a natural plant taxon value (Benkhnigue *et al.* 2022; Chaachouay *et al.* 2020; Gakuubi and Wanzala 2012).

## Results and Discussion

### Socio-demographic profile of respondents

Out of a total of 1700 persons interviewed, 85% (1530) opted to utilize traditional medicine (TM), either alone (75%) or in conjunction with modern medicine (MM), with a 10% share. A total of 648 (42.35%) utilized plants to cure specific liver ailments (hepatitis, jaundice, and liver cancer), and they provided:

- Information on the vernacular name of each species,
- The season of availability in the field,
- Its efficacy,
- The technique of preparation and mode of use.

We could define the consumers of traditional medicine for liver disorders in this area using the various characteristics (Table 2).

Table 2. Socio-demographic profile of respondents (n=648).

Variables	Categories	Number of informants	Frequency (%)
Age category (years)	Between 18 and 30	88	13.58
	Between 31 and 40	112	17.28
	Between 41 and 50	204	31.48
	>50 years	244	37.66
Gender	Female	364	56.17
	Male	284	43.83
Family situation	Married	434	66.98
	Single	214	33.02
Educational level	Illiterate	430	66.36
	Primary	102	15.74
	Secondary	88	13.58
	Superior	28	4.32
Monthly income	Low	251	38.74
	Medium	53	8.18
	High	2	0.30
	No income	342	52.78
Place of residence	Dour	398	61.42
	Village	108	16.67
	City	142	21.91

The data suggest that medicinal species usage was prevalent across all age categories, with a definite preference for persons over 50 (37.66%), indicating that many elderly persons still hold traditional knowledge of herbal treatment. Women outnumbered men (43.83% to 56.17%), with an unbalanced sex ratio (F/M = 1.28). Women's predominance may be explained by being responsible for their families' health and using readily accessible,

effective, and less costly means. These findings corroborate previous ethnobotanical research undertaken in several locales (Belhaj *et al.* 2021; Benkhnigue *et al.* 2010; 2022; Tahraoui *et al.* 2007). Regarding the family situation, 66.98% of the participants were married, while only 33.02% were single. Regarding the academic level, we found a clear predominance of illiterate people (66.36%), followed by the categories of people who have a primary and secondary level, with percentages of 15.74% and 13.58%. At the same time, people at a university level use the most miniature medicinal plants with a percentage of 4.32%. The results also show that 52.78% of these users have no monthly income, 38.74% had a low income, and 8.18% had a medium income. On the other hand, people with higher income represent only 0.30%. In our study, we found that (61.42%) of these people lived in rural areas (Douars). These findings corroborate previous ethnobotanical research in the Rif area (Chaachouay *et al.* 2020).

### **Floristic analysis**

The plants identified and collected during this study are presented in Table 3, categorized according to the alphabetical order of botanical families, the local vernacular name, the biological type, the nature of the plant (wild, cultivated, and imported), the portion utilized, the method of preparation employed by the local community, and the ethnobotanical indices (UV and FUV). To enhance our findings further, we did a literature analysis of ethnobotanical and pharmacological investigations to demonstrate the medicinal efficacy of the plant (hepatoprotective, antioxidant, anti-cancer, and anti-inflammatory).

### **Specific analysis of botanical families**

In this ethnobotanical and ethnopharmacological study we found 86 medicinal species divided into 79 genera and belonging to 37 botanical families, of which four families accounted for 47.67% of the total of species recorded: Asteraceae (17 species), Lamiaceae (9 species), Fabaceae (8 species) and Apiaceae (7 species). The importance of these families can be explained on the one hand by their richness within the Moroccan flora, by their ability to adapt to often diversified environments, and on the other hand by their biogeographic range (Mediterranean). In addition, the other families (33) contribute to 52.33% (45 species) of the total number, of which only one species represent twenty-eight. In addition, the high use of the family Asteraceae can be explained by the fact that they are the wealthiest family of angiosperms, with 128 genera and about 550 species spread throughout Morocco (Fennane and Ibn Tattou 2012). Compared to less wealthy families such as Poaceae, Fabaceae, or Solanaceae, the Asteraceae family has fewer species of economic interest. However, it provides food plants such as *Artemisia arborescens* (Vaill.) L., *Helianthus annuus* L., *Lactuca sativa* L., and *Cynara scolymus* L. are cultivated to extract its vegetable oil. Other species are used in pharmacy as *Artemisia cina* Berg, *Arnica montana* L., *Matricaria chamomilla* L., and *Tussilago farfara* L. Finally; some Asteraceae are used as ornamental plants, including species belonging to the genera *Tagetes*, *Chrysanthemum*, *Dahlia*, *Tanacetum*, *Rudbeckia*, *Zinnia*, *Cosmos*, *Callistephus*, and *Calendula* (El Bouzidi 2013). Phytochemical studies on this family mainly show terpenes, alkaloids, and flavonoids (Benn and Gul 2007; Emerenciano *et al.* 2001; Ferrari *et al.* 2005).

### **Quantitative analysis according to the FUV and UV index**

Based on the FUV index, we found six families being most used in traditional phytotherapy against liver diseases: Caryophyllaceae and Zingiberaceae occupy the first place (FUV= 0.331 for each), followed by Rhamnaceae (FUV= 0.259), Rubiaceae (FUV= 0.228), Apiaceae (FUV=0.187) and Fabaceae (FUV=0.174) (Fig. 3).

## Ethnobotany Research and Applications

Table 3. List of medicinal plants used against liver diseases in the Al-Haouz Rehamna region; their local use, ethnobotanical and ethnopharmacological literature that justifies their therapeutic properties.

<b>Family and species</b>	<b>Vernacular name</b>	<b>Plant type</b>	<b>Biological type</b>	<b>Part used</b>	<b>Preparation</b>	<b>Local use</b>	<b>UV</b>	<b>FUV</b>	<b>Ethnobotanical evidence</b>	<b>Literature to support the therapeutic claims</b>
<b>Amaryllidaceae</b> <b><i>Allium sativum</i> L.</b>	Touma	Cultivated	Geophyte	Bulb	Nature	H, I	0.157	0.157	Jaundice (Miara et al., 2019).	Anti-inflammatory and antioxidant (Shang et al. 2019).
<b>Anacardiaceae</b> <b><i>Pistacia lentiscus</i> L.</b>	Dro	Spontaneous	Phanerophyte	Leaf	Decoction	H	0.123	0.123	Jaundice (Ljubuncic et al., 2005).	Hepatoprotective and antioxidant (Janakat and Al-Merie; Maameri et al. 2015; Mehenni et al. 2016).
<b>Apiaceae</b> <b><i>Ammodaucus leucotrichus</i> Coss et Durieu.</b>	Kammûn essôfi	Cultivated	Therophyte	Seed	Infusion	Cf	0.185	0.187		Anti-inflammatory (Es-Safi et al., 2020).
<b>Apiaceae</b> <b><i>Carum carvi</i> L.</b>	El-Karwiya	Cultivated	Therophyte	Seed	Infusion	H, I	0.201			Hepatoprotective (Samojlik et al. 2010)
<b>Apiaceae</b> <b><i>Coriandrum sativum</i> L.</b>		Cultivated	Therophyte	Seed	Infusion	H	0.083		Blood depurative (Bouayyadi et al., 2015).	Hepatoprotective (Pandey et al. 2011).
<b>Apiaceae</b> <b><i>Petroselinum sativum</i> Hoffman.</b>	Maadenouss	Cultivated	Therophyte	Seed	Decoction	H	0.123		Hepatic (Ben Akka, et al., 2015; Tahri et al., 2012); liver detoxification (Bouayyadi et al., 2015).	Anticancer (Farshori et al. 2013).
<b>Apiaceae</b> <b><i>Pimpinella anisum</i> L.</b>	Habate hlawa	Cultivated	Therophyte	Seed	Infusion	H	0.227		Cholagogue (Tahri et al., 2012); liver disorders (Al-Asmari et al., 2014); anti-inflammatory and antioxidant (Ghlissi et al., 2020).	Anti-inflammatory and antioxidant (Martins et al. 2016).
<b>Apiaceae</b> <b><i>Ridolfia segetum</i> Moris.</b>	Tebche, Silou	Spontaneous	Therophyte	Leafy stem Flower	Infusion	H	0.525		Liver disease (Ben Akka, et al., 2015) and jaundice (Hseini, 2008).	Antitumour (Beeby et al. 2021).
<b>Apiaceae</b> <b><i>Thapsia transtagana</i> Brot.</b>	Deryâs, daryoussa	Spontaneous	Geophyte	Leaf	Poultice	H	0.046			Antioxidant (Alilou and Akssira 2021).

## Ethnobotany Research and Applications

9

<b>Asparagaceae</b> <i>Drimia marítima</i> (L.) Stearn.	El-aansla, Bassal dib, El bassila	Spontaneous	Geophyte	Leaf, Bulb	Poultice	H	0.031	0.031	Anti-inflammatory, antioxidant (Nejatbakhsh et al., 2017); Jaundice (Bellakhdar, 1997; Hseini, 2008); Hepatitis (Ouarghidi et al., 2013).	Antioxidant (Tahri <i>et al.</i> 2020).
<b>Asteraceae</b> <i>Artemisia atlantica</i> Coss et Dur. var. <i>maroccana</i> (Coss) Maire.	Chih Ourika	Spontaneous	Chamephyte	Leafy stem, Flower	Decoction	H	0.117	0.168		
<b>Asteraceae</b> <i>Artemisia herba-alba</i> Asso.	Chih Dwidi	Spontaneous	Chamephyte	Leafy stem, Flower	Decoction	H, I	0.170		Cholagogue (Tahri <i>et al.</i> , 2012) and liver diseases (Eddouks <i>et al.</i> , 2017).	Antioxidant (Boukhenoufa <i>et al.</i> , 2021).
<b>Asteraceae</b> <i>Artemisia huguetii</i> Caball.	Chih beldi	Spontaneous	Chamephyte	Leafy stem, Flower	Decoction	H	0.046			
<b>Asteraceae</b> <i>Calendula stellata</i> Cav.	Jemra	Spontaneous	Therophyte	Flower , Root	Infusion	H, I	0.167		Hepatic (Miara <i>et al.</i> , 2021).	
<b>Asteraceae</b> <i>Carlina gummifera</i> (L.) Less.	Addad	Spontaneous	Hemicryptophyte	Root	Fumigation	H	0.056			Antioxidant (Bouabid <i>et al.</i> , 2020).
<b>Asteraceae</b> <i>Carthamus tinctorius</i> L.	El-aossfore, Kaff essabea, Zaafour	Cultivated	Therophyte	Flower , Leafy stem	Powder	H	0.239			Hepatoprotective and antioxidant (Wang <i>et al.</i> , 2015; Wu <i>et al.</i> , 2013).
<b>Asteraceae</b> <i>Centaurea maroccana</i> Ball.	Bejjâe n-nhal, Negguûr	Spontaneous	Therophyte	Root	Decoction	H, Cf	0.250			Against cancer (Aissous <i>et al.</i> , 2021).
<b>Asteraceae</b> <i>Chrysanthemum coronarium</i> L.	El-gahwâne, Hmessou, Maloule o Aali	Spontaneous	Therophyte	Flower	Decoction	H	0.043			Antioxidant and hepatoprotective (Bardawel <i>et al.</i> , 2015; M. Donia, 2014).
<b>Asteraceae</b> <i>Cichorium intybus</i> L.	Bou-Aggad	Spontaneous	Hemicryptophyte	Leaf	Decoction	H, Cf, I	0.120		Choleretic (Ben Akka <i>et al.</i> , 2015); cholagogue and Choleretic (Bellakhdar, 2006).	Hepatoprotective (Neha <i>et al.</i> , 2014).
<b>Asteraceae</b>	Tâfsse	Spontaneous	Therophyte	Flower	Decoction	H, I, Cf	0.440		Icterus (Benzaabane and Abbad, 1994).	Antioxidant (Aghraz <i>et al.</i> , 2017).

***Cladanthus arabicus***  
(L.) Cass.

									Liver disease (Bouayadi et al., 2015; Hachi et al., 2015; Hseini, 2008); cholagogue and choleretic (Hmamouchi, 1999).
<b>Asteraceae</b> <i>Cynara cardunculus</i> L.	Kharchouf	Cultivated	Geophyte	Stem, Root,	Nature, Decoction	Cf, H	0.324		
<b>Asteraceae</b> <i>Cynara humilis</i> L.	Timta	Spontaneous	Geophyte	Root	Decoction	H, Cf	0.193	Liver diseases (Bellakhdar, 1997; Lahsissène, 2010).	
<b>Asteraceae</b> <i>Cynara scolymus</i> L.	El-qôq	Cultivated	Geophyte	Root, Flower , Leaf	Nature, Decoction	H, Cf, I	0.148	Hepatic (Ben-Akka et al., 2015; Lahsissène, 2010).	Liver improvement (Mejri et al., 2020); Antioxidant, anti- inflammatory, and Anti-hepatitis C (Elsebai et al., 2016).
<b>Asteraceae</b> <i>Matricaria chamomilla</i> L.	Babounj romi	Naturalized	Therophyte	Leafy stem, Flower	Infusion	H	0.025	Jaundice (Bouayyadi et al., 2015).	Hepatoprotective (Shebbo et al., 2020).
<b>Asteraceae</b> <i>Rhaponticum acule</i> (L.) DC.	Tavgha	Spontaneous	Hemicrypto- phyte	Root	Decoction	H, Cf, I	0.380	Liver disease (Lahsissène, 2010).	Antioxidant (Mosbah et al., 2020)
<b>Asteraceae</b> <i>Scolymus hispanicus</i> L.	El-Guernina	Spontaneous	Hemicrypto- phyte	Stem	Cooked	H, Cf, I	0.105	Liver diseases (Lahsissène, 2010), Icterus, and hepatic (Bellakhdar, 1997; Sbai-Juilli et al., 2017).	Anti-inflammatory, hepatoprotective, and antioxidant (Berdja et al., 2021).
<b>Asteraceae</b> <i>Waronia saharae</i> Benth et Coss.	Afessas	Spontaneous	Nanophanero - phyte	Leaf, Root	Decoction	H, Cf, I	0.040		Antioxidant (Ajeblí & Eddouks, 2019).
<b>Berberidaceae</b> <i>Berberis hispanica</i> Boiss & Reut.	Arguïs, Atizar	Spontaneous	Nanophanero - phyte	Ecr	Decoction	H, Cf, I	0.133	0.133	Liver disorders and choleretic (Bellakhdar, 1997).
<b>Brassicaceae</b> <i>Brassica rapa</i> L.	Lefte el- mahfour	Cultivated	Therophyte	Seed	Cooked	H	0.102	0.082	Antioxidant and anticancer (El Fakir et al., 2021).
<b>Brassicaceae</b> <i>Lepidium sativum</i> L.	Habb Er- rachâd, Habb errajee	Cultivated	Therophyte	Seed	Powder	H	0.069		Inflammatory disorders (Al-Asmary et al., 2014).
<b>Brassicaceae</b>	Lefjel	Cultivated	Therophyte	Root	Decoction	H, D	0.077		Antioxidant (Beevi et al., 2012).

## Ethnobotany Research and Applications

11

Raphanus sativus L.									
Caryophyllaceae <i>Corriola telephiifolia</i> Pourret.	Tasserghinte , Ssarghina	Spontaneous	Hemicryptophyte	Root	Powder	H, I	0.331	0.331	Liver diseases (Bellakhdar, 1997). Cytotoxic and antioxidant Doudach et al., 2013).
Combretaceae <i>Combretum micranthum</i> G. Don.	Kinkiliba	Imported	Phanerophyte	Leaf	Infusion	I	0.020	0.02	
Cucurbitaceae <i>Citrullus colocynthis</i> (L.) Schrad.	El-hedja, Taferzît, El handal	Spontaneous	Geophyte	Leaf, Seed	Fumigation	I	0.031	0.031	Hepatitis (Miara et al., 2019).
Cupressaceae <i>Tetraclinis articulata</i> (Vahl) Masters.	El-ar'ar, Azouka, Al qitrane laghlide	Spontaneous	Phanerophyte	Leaf	Decoction	H, I	0.012	0.012	Antioxidant (Rabib et al., 2020).
Fabaceae <i>Cassia fistula</i> L.	Aud ssalib	Imported	Phanerophyte	Fruit	Distillation	Cf	0.006	0.174	Anti-inflammatory, hepato-protective, anticancer (Chaudhari & Professor, 2013); Liver trouble (Uniyal et al., 2011). Hepatoprotective and antioxidant (Pradeep et al., 2010).
Fabaceae <i>Ceratonia siliqua</i> L.	L-kharrôb, Tikidda	Cultivated, Sup Spontaneous	Phanerophyte	Fruit	Powder	H, Cf, I, D	0.096		Anti-inflammatory and antioxidant (Rtibi et al., 2017).
Fabaceae <i>Cicer arietinum</i> L.	El hommess	Cultivated	Therophyte	Seed	Maceration	H	0.306		Jaundice (Bouayyadi et al., 2015; El-Azzouzi & Zidane, 2015; Hseini & Kahouadji, 2007; Tahir et al., 2012). Antioxidant (Ahmad Nadzri et al., 2021).
Fabaceae <i>Glycyrrhiza glabra</i> L.	Aarq assûss	Imported	Hemicryptophyte	Root	Decoction	H	0.074		Hepatoprotective, anticancer, and anti-inflammatory (Çevik et al., 2018).
Fabaceae <i>Lens culinaris</i> Medik.	L'âdесс	Cultivated	Therophyte	Seed	Powder	H	0.168		Antioxidant (Jameel et al., 2015).
Fabaceae <i>Medicago sativa</i> L.	Fessa	Cultivated	Hemicryptophyte	Seed	Powder	H, I	0.111		Hepatitis (Ben-Akka et al., 2015). Anti-inflammatory (Seddighfar et al., 2020).
Fabaceae <i>Ononis natrix</i> L.	Afesda d	Spontaneous	Chamophyte	Leafy stem	Powder	H	0.486		Icterus (Ghourri et al., 2012; Sbai-Juillili et al., 2017). Antioxidant (Sayari et al., 2016).
Fabaceae <i>Trigonella foenum-graecum</i> L.	El-halba, Tifidass	Cultivated	Therophyte	Seed	Powder, Poultice	H, I	0.142		Hepatitis and icterus (Hachi et al., 2015); depurative (Lahsissène, 2010). Hepatoprotective (Kaviaraslan et al., 2007); anti-inflammatory (Mandegary et al., 2012); anticancer (Verma et al., 2010).

## Ethnobotany Research and Applications

12

<b>Fagaceae</b> <i>Quercus ilex L.</i>	Dbagh laghlide, Lakrouche, Tassat	Spontaneous	Phanerophyte	Bark	Decoction	H, I	0.015	0.015	Antioxidant and anticancer (Amessis-Ouchemoukh et al., 2017).
<b>Gentianaceae</b> <i>Centaureum erythraea</i> Rafn.	Gosset alhayia	Spontaneous	Therophyte	Leafy stem	Infusion	Cf	0.171	0.171	Hepatoprotective (Hamza et al., 2015).
<b>Geraniaceae</b> <i>Pelargonium roseum</i> Willd.	Laatter cha	Cultivated	Geophyte	Leaf	Infusion	H, Cf, I, D	0.022	0.022	
<b>Lamiaceae</b> <i>Ajuga iva</i> (L.) Schreb.	Chenggûra, Tûf tolba	Spontaneous	Hemicryptophyte	Leafy stem, Flower	Decoction	H	0.046	0.073	Choleretic (Tahri et al., 2012). Antioxidant (Saad et al., 2019); anticancer (Bouyaha et al., 2020).
<b>Lamiaceae</b> <i>Lavandula officinalis</i> Chaix ex Villars.	El-khzama fassiya	Cultivated	Chamephyte	Flower	Decoction	I, H	0.039		choleretic (Bellakhdar, 2006).
<b>Lamiaceae</b> <i>Lavandula multifida</i> L.	EL-kohayla, Kohaylate el himir	Spontaneous	Chamephyte	Leafy stem, Infl	Pow	H, Cf, I	0.068		Hepatic (Eddouks et al., 2017).
<b>Lamiaceae</b> <i>Marrubium vulgare</i> L.	Marro ut	Spontaneous	Chamephyte	Leafy stem	Poultice	H	0.025		Icterus (Bammi & Douira, 2002; El-Azzouzi & Zidane, 2015; Lahsissène, 2010); cholagogue and jaundice (Tahri et al., 2012). Hepatoprotective (Verma et al., 2012); cytotoxic (Zarai et al., 2011); Antioxidant (Akther et al., 2013).
<b>Lamiaceae</b> <i>Origanum compactum</i> Bentham.	Zaater tadlawi	Spontaneous	Chamephyte	Leaf, Flower	Infusion	H, I	0.076		
<b>Lamiaceae</b> <i>Salvia rosmarinus</i> Spenn.= <i>Rosmarinus officinalis</i> L.	Al-Azir, Yazir	Spontaneous	Nanophanero - phyte	Leaf	Infusion	H, I	0.059		Cholagogue and choleretic (Bellakhdar, 2006; Tahri et al., 2012); anti-inflammatory, cholagogue (Ould El Hadj et al., 2003). Hepatoprotective (Abdel-Wahhab et al., 2011); antioxidant (Bajalan et al., 2017).
<b>Lamiaceae</b> <i>Thymus satureioides</i> Cosson. & Bal.	Zîtra, Azoukoni	Spontaneous	Chamephyte	Leaf	Infusion	H, I	0.151		Choleretic (Ben-Akka et al., 2015). Anti-inflammatory (Ismaili et al., 2004).
<b>Lamiaceae</b> <i>Thymus zygis</i> L.	Zaitra.	Spontaneous	Chamephyte	Leaf	Infusion	H, I	0.110		

## Ethnobotany Research and Applications

13

<b>Lamiaceae</b> <i>Vitex agnus-castus</i> L.	El-kherwa, Angarf	Spontaneous	Nanophanero - phyte	Fruit	Poultice	H, I	0.085		
<b>Lauraceae</b> <i>Cinnamomum zeylanicum</i> Nees.	Dar ssini	Imported	Phanerophyte	Bark	Decoction	I	0.120	0.096	Liver disease (El Azzouzi & Zidane, 2015). Antioxidant and anti-microbial (Ranasinghe et al., 2013).
<b>Lauraceae</b> <i>Cinnamomum cassia</i> Blum.	L-Qarfa	Imported	Phanerophyte	Bark	Decoction	I	0.093		Anti-inflammatory, and anti-apoptotic (Golshahi et al., 2019); hepatoprotective (Mohammad et al., 2021).
<b>Lauraceae</b> <i>Laurus nobilis</i> L.	War akat-sîdna Moussa, E-rrand	Spontaneous	Phanerophyte	Leaf	Infusion	H, I	0.074		Antioxidant and anti-inflammatory (Ozcan et al., 2010; Taban et al., 2018).
<b>Lythraceae</b> <i>Punica granatum</i> L.	Rommâne	Cultivated	Phanerophyte	Bark	Powder	H, Cf, I	0.215	0.215	Hepatoprotective (Yogeeta et al., 2007).
<b>Malvaceae</b> <i>Hibiscus sabdariffa</i> L.	El Karkadi	Imported	Phanerophyte	Flower	Infusion	H	0.198	0.198	Jaundice and liver disease (El-Azzouzi & Zidane, 2015). Hepatoprotective (Da-Costa-Rocha et al., 2014).
<b>Moraceae</b> <i>Ficus carica</i> L.	l-karmouss, El karma, Chreha	Cultivated	Phanerophyte	Leaf, Fruit	Powder	H	0.099	0.099	Icterus (El-Azzouzi & Zidane, 2015). Antioxidant and anticancer (Jasmine et al., 2015).
<b>Myrtaceae</b> <i>Eugenia caryophyllata</i> Thunb.	Kronffel, Uod nuwwâr	Imported	Phanerophyte	Fruit	Decoction	H, Cf, I	0.184	0.184	Anti-inflammatory and anticancer (Han & Parker, 2017).
<b>Papaveraceae</b> <i>Fumaria parviflora</i> Lam.	Hachichat as-sebyâne, Sibana, Narelbara, Chehmata el felouss	Spontaneous	Therophyte	Leafy stem	Infusion	I	0.019	0.019	Hepatoprotective (Tripathi et al., 2010).
<b>Parmeliaceae</b> <i>Evernia prunastri</i> Ach.	Lihyate E-chikh	Spontaneous	Fixed-hydrophyte	Thalle	Decoction	D	0.011	0.011	Antioxidant (Shcherbakova et al., 2021).
<b>Pinaceae</b> <i>Pinus halepensis</i> Miler.	Tayda, Aode enarr, Chejrat ssanawbar	Spontaneous, Cultivated	Phanerophyte	Bark, Leaf, Cof	Powder Infusion	H	0.005	0.005	Antioxidant and anti-inflammatory (Abbou et al., 2019).
<b>Plumbaginaceae</b> <i>Armeria maritima</i> Wallr.	Erq Awadmî. El-Aarqlahmer	Spontaneous	Hemicryptophyte	Root	Decoction	D	0.003	0.003	
<b>Poaceae</b> <i>Hordeum vulgare</i> L.	Chaâir	Cultivated	Therophyte	Seed	Maceration	H, I	0,025	0.018	Antioxidant (Deng et al., 2020).
<b>Poaceae</b> <i>Poa bulbosa</i> L.	Annadkher	Spontaneous	Hemicryptophyte	Rhizome	Decoction	H	0.012		

<b>Poaceae</b> <i>Zea mays</i> L.	Dra	Cultivated	Therophyte	Seed, Sty	Powder	H, I	0.019	Cirrhosis (Bouayyadi et al., 2015); cholagogue & choleric (Tahri & al., 2012; Saidi, 1999). Liver disorders, and inflammatory disorders (Al-Asmari et al., 2014).	Cytotoxicity (Rajkumar et al., 2019).
<b>Portulacaceae</b> <i>Portulaca oleracea</i> L.	Rejla	Spontaneous	Therophyte	Leafy stem	Cooked	H	0.151	0.151	Hepatoprotective (Farkhondeh & Samarghandian, 2019).
<b>Polygonaceae</b> <i>Emex spinosa</i> (L.) Campd.	Homayda romiya, Aycha mo thaycha	Spontaneous	Therophyte	Leaf, Root	Cooked	H	0.182	0.156	Jaundice (Bammi and Douira, 2002). Cytotoxic and antimicrobial (Donia et al., 2014).
<b>Polygonaceae</b> <i>Rumex crispus</i> L.	Tarteka, nfifiha	Spontaneous	Hemicryptophyte	Seed, Root	Cooked	H	0.130		Antioxidant, cytotoxic, and anti-acetylcholinesterase (Saoudi et al., 2021).
<b>Ranunculaceae</b> <i>Nigella sativa</i> L.	Haba ssawda, Ssanouj	Cultivated, Sup	Therophyte	Seed	Powder	H, Cf, I	0.043	0.024	Liver disease (Bouayyadi et al., 2015); cholagogue (Hmamouchi, 1999), liver tonics, anti-inflammatory, immunostimulant, and remedy for jaundice (Al-Asmari et al., 2014). Hepatoprotective (Hegazy et al., 2018).
<b>Ranunculaceae</b> <i>Ranunculus muricatus</i> L.	Wedene el halouf	Spontaneous	Therophyte	Root	Powder	H	0.006		Jaundice (Ullah et al., 2013). Antioxidant (Deghima et al., 2020).
<b>Rhamnaceae</b> <i>Rhamnus alaternus</i> L.	Amlīss, Aferzadade, Wariwri	Spontaneous	Nanophanerophyte	Leafy stem, Leaf, Bark, Fruit	Decoction	H, I, Cf	0.463	0.259	Icterus hepatic (Miara et al., 2019); jaundice (Lahissène, 2010), and liver diseases (Ljubuncic et al., 2005). Hepatoprotective (Berroukche et al., 2015).
<b>Rhamnaceae</b> <i>Ziziphus lotus</i> (L.) Lam.	Ssedra, Nbeg Azogar	Spontaneous	Nanophanero - phyte	Fruit	Powder	H, I	0.056		Anti-inflammatory (Miara et al., 2019). Antihyperlipidemic and antioxidant (Bencheikh et al., 2021).
<b>Rubiaceae</b> <i>Rubia peregrina</i> L.	Fuwwa, Tarūbia	Spontaneous	Chamephyte	Root	Powder	H, I	0.228	0.228	Hepatitis (Ouarghidi et al., 2013); jaundice and liver diseases (Bammi et Douira, 2002; El-Azzouzi & Zidane, 2015). Antioxidant and anticancer (Longo et al., 2008).

## Ethnobotany Research and Applications

15

<b>Rutaceae</b> <i>Citrus aurantium</i> L. var. <i>amara</i> Link.	Ranj, Trenje Zenboue	Cultivated	Phanerophyte	Fruit	Jus	I	0.023	0.023	Cholagogue (Sijelmassi, 2011).	Anti-inflammatory (Shen et al., 2017).
<b>Schisandraceae</b> <i>Illicium verum</i> Hook. F.	Badian a	Imported	Phanerophyte	Fruit	Decoction	H, I	0.017	0.017		Anti-inflammatory (Tuseef et al., 2021).
<b>Thymelaeaceae</b> <i>Daphne gnidium</i> L.	Alezaz, Lezaze	Spontaneous	Chamephyte	Leaf	Infusion	H	0.002	0.002	Antiinflammatory (Boudjelal et al., 2013).	Anticancer (Sanna et al., 2015); antioxidant (Chaves et al., 2020).
<b>Vitaceae</b> <i>Vitis vinifera</i> L.	Dalya, zbibe, kerma, Al- ainab	Cultivated	Phanerophyte	Fruit	Maceration	H	0.170	0.17		Hepatoprotective (Ahmad & Khan, 2012).
<b>Xanthorrhoeaceae</b> <i>Asphodelus ramosus</i> L.	El berwague, blallûze	Spontaneous	Geophyte	Rhizome	Powder	H	0.002	0.002		
<b>Zingiberaceae</b> <i>Aframomum melegueta</i> K. Schum.	El goza rqîqa ou sahrawiya	Imported	Geophyte	Seed	Powder	H	0.255	0.331		Hepatoprotective and antioxidant (Adefegha et al., 2016).
<b>Zingiberaceae</b> <i>Curcuma longa</i> L.	Kharcûm aorouk, Al airk al assfar	Imported	Geophyte	Rhizome	Powder	H, Cf	0.494		Hepatitis (Ben-Akka, et al., 2015); jaundice (Lahsissène, 2010); jaundice and liver problems (Al-Asmari et al., 2014).	Antitumor, antioxidant, hepatoprotective, and anti-inflammatory (Darvesh et al., 2012; Dhatchayani et al., 2020).
<b>Zingiberaceae</b> <i>Zingiber officinale</i> Rosc.	Skenjbîr	Imported	Geophyte	Rhizome	Powder	H	0.244		Hepatic (Hachi et al., 2015).	Antioxidant (Otunola et al., 2017).
<b>Zygophyllaceae</b> <i>Zygophyllum gaetulum</i> Emberger & Maire.	El-aaggâya	Spontaneous	Chamephyte	Seed, Flower	Decoction	I	0.026	0.026	Antiinflammatory (Bellakhdar, 1997)	Antiinflammatory (Ait El Cadi et al., 2012).

(H: hepatitis, I: Icterus, J: jaundice, D: Depurative, Cf: Liver cancer).

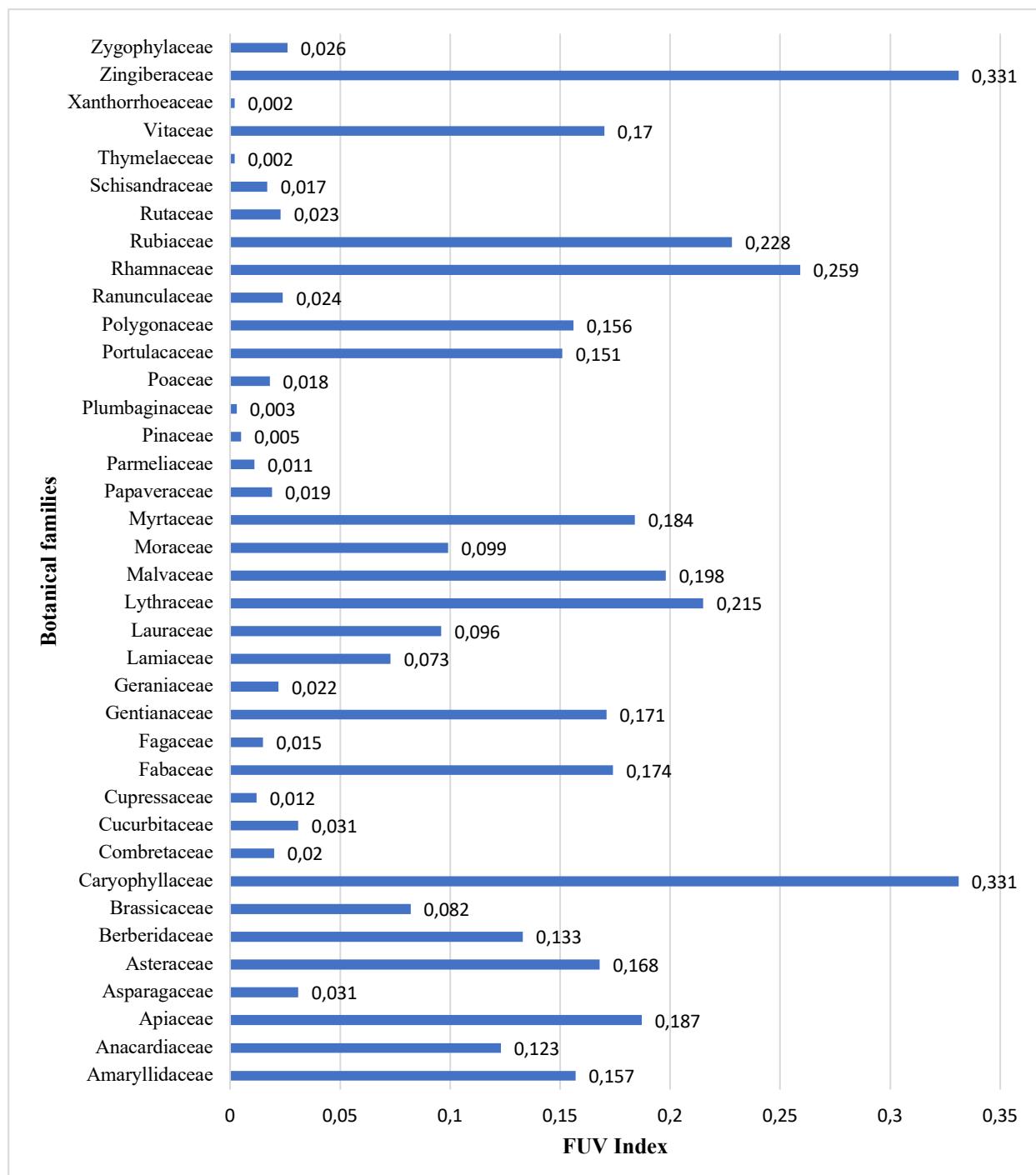


Figure 3. Family use-value (FUV) of medicinal plants.

The calculation of the UV index allowed us to extract seven species that had the highest UV because the informants cited them more: *Curcuma longa* L. (UV= 0.494), *Ononis natrix* L. (UV= 0.486), *Rhamnus alaternus* L. (UV= 0.463), *Cladanthus arabicus* (L.) Cam. (UV= 0.440), *Rhaponticum acaule* (L.) DC. (UV= 0.380), *Cynara cardunculus* L. (UV=0.324) and *Cicer arietinum* L. (UV= 0.306). However, we found two species with the smallest UV: *Asphodelus ramosus* L. and *Daphne gnidium* L. (UV= 0.002 for each). The plant species with the highest UV shoulkd be studied phytochemically and pharmacologically to identify their chemical constituents and possible active ingredients responsible for their hepatoprotective property. The analysis of the data matrix composed of all the respondents and the species identified according to the diseases treated shows a significant number of notifications spread over the 86 species, of which the most requested by the respondents are presented in Figure 4.

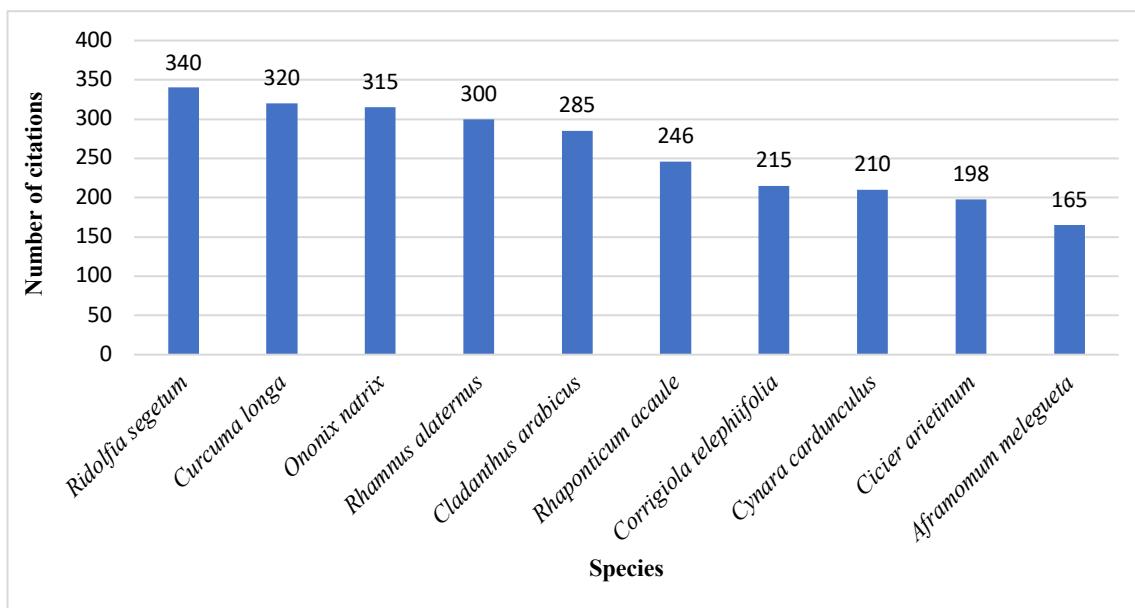


Figure 4: Distribution of the most commonly used species against liver diseases in the Al-Haouz Rehamna region with their number of citations.

Four plants were more cited in the said region (having several citations greater than or equal to 300), and they are considered effective against liver diseases:

***Ridolfia segetum*** (Apiaceae), known by the vernacular name Tebche, is an annual, therophyte plant, spontaneous in plains and low mountains, growing mainly in crops and uncultivated fields, living in arid to humid climate conditions and widely distributed in the Mediterranean countries. It was reported 340 times and is used for its leafy stems or inflorescences, in powder form, to treat hepatitis.

***Curcuma longa*** (Zingiberaceae), commonly called Kharkoum-aorouk, is a perennial plant, a geophyte, imported from Southeast Asia. It was cited 320 times, widely used for its rhizomes, in powder form, in the treatment of jaundice and hepatitis.

***Ononix natrix*** (Fabaceae), locally called Afesdade; is a spontaneous plant, chamaephyte, which lives in clear forests, matorals, ermes; in plains, low and medium mountains; in climate ranging from Saharan to subhumid zones and was reported 315 times. Its leafy stems were dried in the sun, and used in decoction in water or powder, against hepatitis.

***Rhamnus alaternus*** (Rhamnaceae) has the vernacular name Amriris; it is a spontaneous plant, phanerophyte, living in plains and mountains in the climate ranging from semi-arid to humid. This species was quoted 300 times. The leafy stems or the bark of the trunk, dried in the sun, are wildly used in decoction in the water against the hepatic affections.

All these species were included in several preparations, alone or associated with other plants.

Out of a total of 86 taxa, 48 species were cited in other similar previous ethnobotanical studies conducted nationally and internationally (Tab. 3). In addition, the hepatoprotective, antioxidant, anticancer or anti-inflammatory activity of 70 plants has been proven by pharmacological studies: *Allium sativum*, *Pistacia lentiscus*, *Ammodaucus leucotrichus*, *Carum carvi*, *Coriandrum sativum*, *Petroselinum sativum*, *Pimpinella anisum*, *Ridolfia segetum*, *Thapsia transtagana*, *Drimia marítima*, *Artemisia herba-alba*, *Carlina gummifera*, *Carthamus tinctorius*, *Centaurea maroccana*, *Chrysanthemum coronarium*, *Cladanthus arabicus*, *Cichorium intybus*, *Cynara cardunculus*, *Cynara scolymus*, *Matricaria chamomilla*, *Rhaponticum acaule*, *Scolymus hispanicus*, *Warionia saharae*, *Berberis hispanica*, *Brassica rapa*, *Lepidium sativum*, *Raphanus sativus*, *Corrigiola telephifolia*, *Tetraclinis articulata*, *Cassia fistula*, *Ceratonia siliqua*, *Cicer arietinum*, *Glycyrrhiza glabra*, *Lens culinaris*, *Medicago sativa*, *Trigonella foenum graecum*, *Quercus ilex*, *Centaurium erythraea*, *Ajuga iva*, *Marrubium vulgare*, *Rosmarinus officinalis*, *Thymus satureioides*, *Cinnamomum zeylanicum*, *Cinnamomum cassia*, *Laurus nobilis*, *Punica granatum*, *Hibiscus sabdariffa*, *Ficus carica*,

*Eugenia caryophyllata*, *Fumaria parviflora*, *Evernia prunastri*, *Pinus halepensis*, *Hordeum vulgare*, *Zea mays*, *Portulaca oleracea*, *Emex spinosa*, *Rumex crispus*, *Nigella sativa*, *Ranunculus muricatus*, *Rhamnus alaternus*, *Ziziphus lotus*, *Rubia peregrina*, *Citrus aurantium* var. *amara*, *Illicium verum*, *Vitis vinifera*, *Asphodelus ramosus*, *Aframomum melegueta*, *Curcuma longa*, *Zingiber officinale* and *Zygophyllum gaetulum* (Table 3). Therefore, it would be exciting to carry out phytochemical and pharmacological studies for the other species to identify other chemical molecules or natural drugs with hepato-protective properties.

#### Disease categories and their IAR values

In the present study, the IAR values ranged from 0.970 to 0.992 by categories of diseases treated (Tab. 4). Liver cancer with IAR = 0.992 represents the disease with the highest degree of agreement among the informants, followed by hepatitis (IAR =0.989) and jaundice (IAR =0.987). In contrast, plants used as detoxifiers were only represented by (IAR=0.970). Therefore, it can be inferred that there is a reasonable likelihood between the people involved in this study and the use of these medicinal species (Lin et al., 2002) and that the people in the study area share the knowledge of medicinal plants used to treat these liver conditions. Therefore, liver cancer was the most common disease in the said area. This may be the result of several reasons: the lack of hygiene, the high consumption of alcohol or medicinal products, and the abusive use of some toxic or hepatotoxic plants because the majority of the people concerned by this study live in rural areas, they are illiterate, and they use the spontaneous plants that are in their surroundings. Therefore, the species with high IAR should be the subject of phytochemical, pharmacological, and toxicological research to evaluate their therapeutic properties.

Table 5. IAR values by categories for treating metabolic diseases.

Categories	List of plant species used and number of uses	Nt	Nur	IAR
<b>Jaundice</b>	<i>Allium sativum</i> (102), <i>Carum carvi</i> (130), <i>Artemisia herba-alba</i> (110), <i>Calendula stellata</i> (108), <i>Cichorium intybus</i> (78), <i>Cladanthus arabicus</i> (285), <i>Cynara scolymus</i> (96), <i>Rhaponticum acaule</i> (246), <i>Scolymus hispanicus</i> (68), <i>Warionia saharae</i> (26), <i>Berberis hispanica</i> (86), <i>Corrigiola telephifolia</i> (215), <i>Combretum micranthum</i> (13), <i>Citrullus colocynthis</i> (20), <i>Tetraclinis articulata</i> (8), <i>Ceratonia siliqua</i> (62), <i>Trigonella foenum graecum</i> (92), <i>Quercus ilex</i> (10), <i>Pelargonium roseum</i> (14), <i>Lavandula officinalis</i> (25), <i>Lavandula multifida</i> (44), <i>Origanum compactum</i> (49), <i>Salvia rosmarinus</i> (38), <i>Thymus satureioides</i> (98), <i>Thymus zygis</i> (71), <i>Vitex agnus-castus</i> (55), <i>Cinnamomum zeylanicum</i> (78), <i>Cinnamomum cassia</i> (60), <i>Laurus nobilis</i> (48), <i>Punica granatum</i> (139), <i>Eugenia caryophyllata</i> (119), <i>Fumaria parviflora</i> (12), <i>Hordeum vulgare</i> (16), <i>Zea mays</i> (12), <i>Nigella sativa</i> (28), <i>Rhamnus alaternus</i> (300), <i>Ziziphus lotus</i> (36), <i>Rubia peregrina</i> (148), <i>Citrus aurantium</i> var. <i>amara</i> (15); <i>Illicium verum</i> (11), <i>Zygophyllum gaetulum</i> (17).	41	3188	0.987
<b>Hepatitis</b>	<i>Allium sativum</i> (102), <i>Pistacia lentiscus</i> (80), <i>Carum carvi</i> (130), <i>Coriandrum sativum</i> (54), <i>Petroselinum sativum</i> (80), <i>Pimpinella anisum</i> (147), <i>Thapsia transtagana</i> (30), <i>Drimia marítima</i> (20), <i>Ridolfia segetum</i> (340), <i>Artemisia atlantica</i> var. <i>Maroccana</i> (76), <i>Artemisia herba-alba</i> (110), <i>Artemisia huguetii</i> (30), <i>Calendula stellata</i> (108), <i>Carlina gummifera</i> (36), <i>Carthamus tinctorius</i> (155), <i>Centaurea maroccana</i> (162), <i>Chrysanthemum coronarium</i> (28), <i>Cichorium intybus</i> (78), <i>Cladanthus arabicus</i> (285), <i>Cynara cardunculus</i> (210), <i>Cynara humilis</i> (125), <i>Cynara scolymus</i> (96), <i>Matricaria chamomilla</i> (16), <i>Rhaponticum acaule</i> (246), <i>Scolymus hispanicus</i> (68), <i>Warionia saharae</i> (26), <i>Berberis hispanica</i> (86), <i>Brassica rapa</i> (66), <i>Lepidium sativum</i> (45), <i>Corrigiola telephifolia</i> (215), <i>Tetraclinis articulata</i> (8), <i>Ceratonia siliqua</i> (62), <i>Cicer arietinum</i> (198), <i>Glycyrrhiza glabra</i> (48), <i>Lens culinaris</i> (109), <i>Medicago sativa</i> (72), <i>Ononis natrix</i> (315), <i>Trigonella foenum graecum</i> (92), <i>Quercus ilex</i> (10), <i>Pelargonium roseum</i> (14), <i>Ajuga iva</i> (30), <i>Lavandula officinalis</i> (25), <i>Lavandula multifida</i> (44), <i>Marrubium vulgare</i> (16), <i>Origanum compactum</i> (49), <i>Salvia rosmarinus</i> (38), <i>Thymus satureioides</i> (98), <i>Thymus zygis</i> (71), <i>Vitex agnus-castus</i> (55), <i>Laurus nobilis</i> (48), <i>Punica granatum</i> (139),	73	6723	0.989

	<i>Hibiscus sabdariffa</i> (128), <i>Ficus carica</i> (64), <i>Eugenia caryophyllata</i> (119), <i>Pinus halepensis</i> (3), <i>Hordeum vulgare</i> (16), <i>Poa bulbosa</i> (8), <i>Zea mays</i> (12), <i>Portulaca oleracea</i> (98), <i>Emex spinosa</i> (118), <i>Rumex crispus</i> (84), <i>Nigella sativa</i> (28), <i>Ranunculus muricatus</i> (4), <i>Rhamnus alaternus</i> (300), <i>Ziziphus lotus</i> (36), <i>Rubia peregrina</i> (148), <i>Illicium verum</i> (11), <i>Daphne gnidium</i> (1), <i>Vitis vinifera</i> (110), <i>Asphodelus ramosus</i> (1), <i>Aframomum melegueta</i> (165), <i>Curcuma longa</i> (320), <i>Zingiber officinale</i> (158).			
<b>liver cancer</b>	<i>Ammodaucus leucotrichus</i> (120), <i>Centaurea maroccana</i> (162), <i>Cichorium intybus</i> (78), <i>Cladanthus arabicus</i> (285), <i>Cynara cardunculus</i> (210), <i>Cynara humilis</i> (125), <i>Cynara scolymus</i> (96), <i>Rhaponticum acaule</i> (246), <i>Scolymus hispanicus</i> (68), <i>Warionia saharae</i> (26), <i>Berberis hispanica</i> (86), <i>Cassia fistula</i> (4), <i>Ceratonia siliqua</i> (62), <i>Centaurium erythraea</i> (111), <i>Lavandula multifida</i> (44), <i>Punica granatum</i> (139), <i>Eugenia caryophyllata</i> (119), <i>Nigella sativa</i> (28), <i>Rhamnus alaternus</i> (300), <i>Curcuma longa</i> (320).	20	2643	0.992
<b>Detoxifying</b>	<i>Raphanus sativus</i> (50), <i>Ceratonia siliqua</i> (62), <i>Pelargonium roseum</i> (14), <i>Evernia prunastri</i> (7), <i>Armeria mauritanica</i> (2).	5	135	0.970

### Biological spectrum

In treating liver illnesses, the therophytes (30.23%) dominate the biological spectrum of all 86 species. The substantial presence of therophytes may be explained, on the one hand, by the fact that they are annual species with short life cycles that are often better suited to frequently disturbing settings (Benkhnigue *et al.* 2022; Bouhache *et al.* 2002). In contrast, these species are readily available, prolific, and simple to harvest. Phanerophytes maintain second place (19.77%), followed by chameophytes (15.12%), geophytes 13.95%; hemicryptophytes occupy the fifth place with 12.79%, nanophanerophytes come in sixth place and contribute to 6.98%. At the same time, hydrophytes are represented by only 1.16% (Fig. 5).

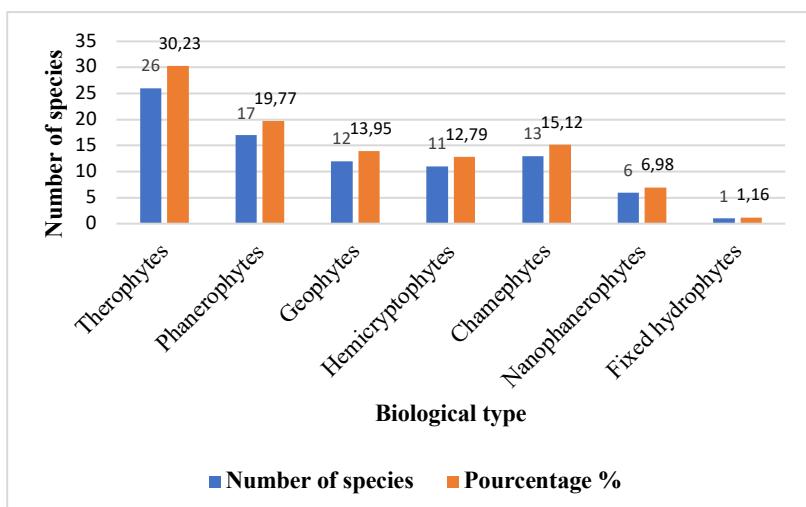


Figure 5. Distribution of the medicinal flora of the Al-Haouz Rehamna region according to their biological types

### Type of plants used

Spontaneous species dominated the other types and contributed 54.65% of the total number of species harvested, followed by cultivated species (31.40%), with species imported from other countries (e.g., Asia, India, Egypt) occupying third place with 12.79%. In comparison, naturalized species are represented by only two species (1.16%) (Fig. 6). Priority should be given to the protection of naturally occurring species since their overexploitation via overharvesting threatens their populations.

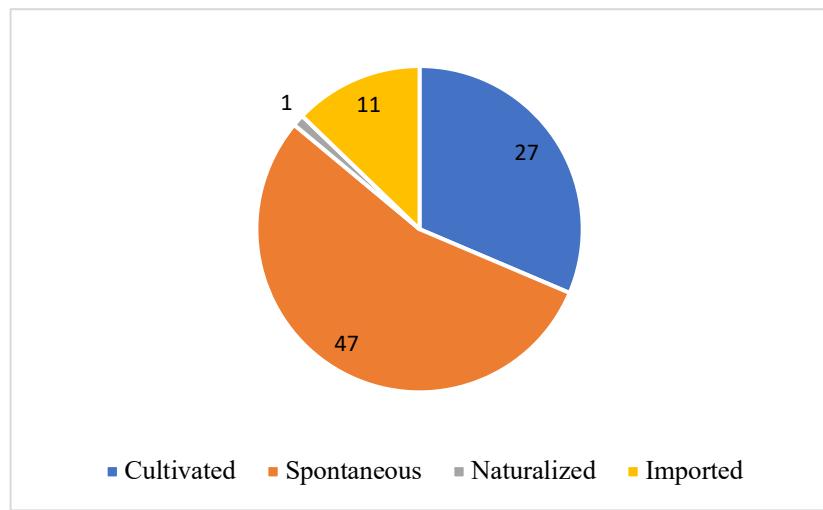


Figure 6. Distribution of plants used against liver diseases according to their degree of spontaneity.

#### Parts of medicinal plants used to treat liver diseases

Based on the PPV index (value of the part of the plant), the leaf was the most used part in local phytotherapy against liver diseases in the studied region with a PPV= 0.183; then comes the seed (PPV= 0.165), the root (PPV=0.147), the flower (PPV= 0.137) and the leafy stem (PPV= 0.128). The other parts (bulb, rhizome, stem, bark, and fruit) had a PPV index below 0.1 (Fig. 7). The great frequency of usage of leaves may be explained on the one hand by their simplicity of harvesting and, on the other hand, by their high concentration of active phytochemicals, which play an essential role in disease treatment (Ahmad *et al.* 2009). They are the primary source of alkaloids, heterosides, and essential oils (Ould El hadj *et al.* 2003).

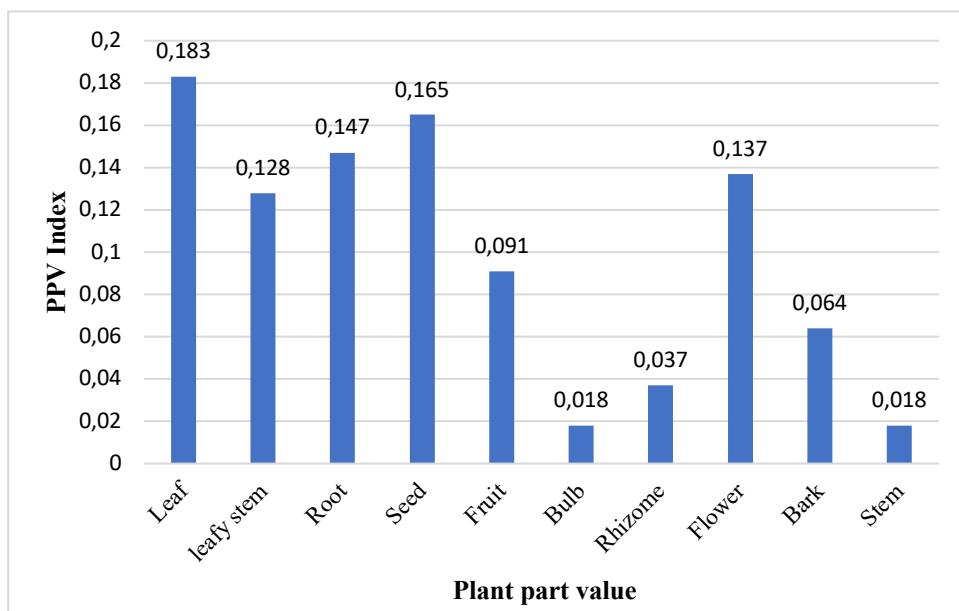


Figure 7. Plant parts used to treat Hepatic disorders in the study area.

#### Method of preparation and use, dosage and dose

Concerning the mode of preparation, we found that the majority of the persons in this study used mainly two modes to prepare anti-hepatic recipes: decoction (34.88%) and powder (23.26%) (Fig. 8). The scopes of the therapeutic formulae are administered to the patients was in most cases orally via water or by trituration with honey. These results are close to those of Sangare *et al.* (2013), who made a similar study on hepatotropic plants and the traditional use of *Gomphrena celosioides* Mart. (Amaranthaceae) in Benin. Apoultece at the abdominal level and fumigation were rarely used and were reserved for certain toxic plants whose toxicity was well known by the local people. The choice of oral administration may be justified because hepatic disorders are associated with internal organs. To aid absorption, all compounds must travel through the digestive system to reach the target

cells (Tra Bi *et al.* 2008). In addition, some users choose to supplement the medication with a diet high in vitamins derived from natural sources, such as oranges, bananas, and apples. They encourage ill individuals to avoid eating beans and eggs.

Regarding dosage, the majority of individuals (88.14%) utilized non-precise dosages (pinch, tablespoon, handful), whereas a minority (11.86%) used exact quantities. These findings corroborate prior ethnobotanical research undertaken in different parts of Morocco (Benkhnigue *et al.* 2010; 2016; Najem *et al.* 2018).

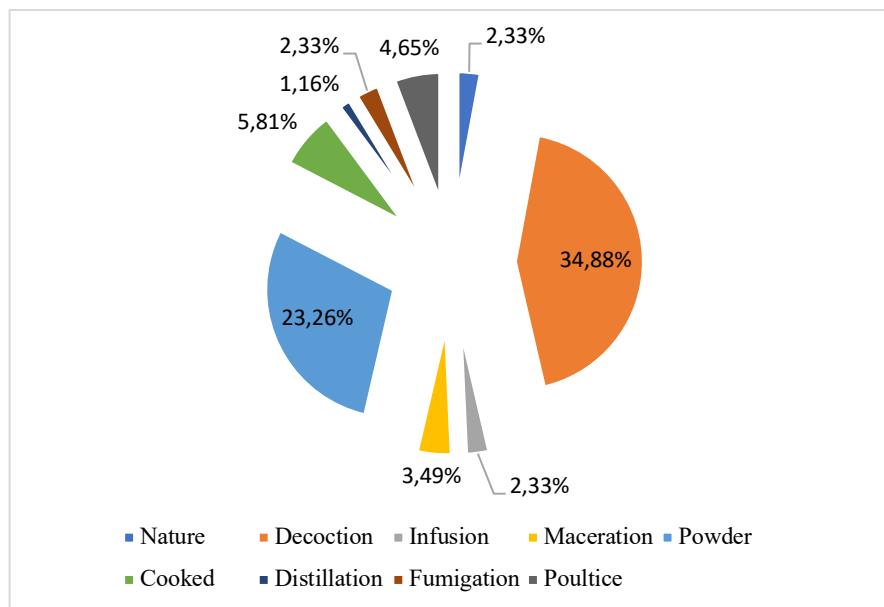


Figure 8. Frequency of the different methods of preparation.

#### Risk of toxicity related to herbal medicine in the Al-Haouz Rehamna region

Any modernization or optimization of the yields of medicinal plants does not seem feasible to us without the knowledge and assimilation of traditional practices in this field. In this perspective, we focused on the declaration of the interviewees concerning the toxicity caused by the use of certain plants.

The pharmacological analysis of the plants identified 21 species with toxicity, traditionally used against liver diseases in the region: *Allium sativum* L. (Gatsing *et al.* 2005; Filliat 2012); *Petroselinum sativum* Hoffman. (Perelman and Kuttin 1988); *Thapsia transtagana* Brot. (Bellakhdar 1997; Hammiche *et al.* 2013); *Drimia marítima* (L.) Stearn. (Hamouda *et al.* 2016); *Artemisia herba-alba* Asso. (Bellakhdar 1997); *Carlina gummifera* (L.) Less.; (Masri *et al.* 2009); *Brassica rapa* L. (Hammiche *et al.* 2013); *Lepidium sativum* L. (Bellakhdar 1997); *Corrigiola telephifolia* Pourret (Lakmichi *et al.* 2011); *Citrullus colocynthis* (L.) Schrad. (Hammiche *et al.* 2013); *Tetraclinis articulata* (Vahl.) Masters. (Zahir and Rahmani 2020); *Glycyrrhiza glabra* L. (Hmamouchi 1999); *Medicago sativa* L. (Bellakhdar 1997); *Trigonella foenum graecum* L. (Ouzir *et al.* 2016); *Quercus ilex* L. (Sijelmassi 2011); *Lavandula officinalis* Chaix. ex Villars. (Sijelmassi 2011); *Origanum compactum* Bentham. (Hmamouchi 1999); *Rosmarinus officinalis* L. (Hmamouchi 1999); *Portulaca oleracea* L. (Shafi and Tabassum 2013); *Nigella sativa* L. (Bellakhdar 1997); *Daphne gnidium* L. (Bellakhdar 1997; Sijelmassi 2011). Among these species, only six were well known locally: *Thapsia transtagana* Brot, *Drimia marítima* (L.) Stearn, *Carlina gummifera* (L.) Less, *Citrullus colocynthis* (L.) Schrad, *Marrubium vulgare* L., and *Daphne gnidium* L., while 15, were not known to be toxic. Ultimately, self-medication with these plants might be harmful. This is why pharmacological and toxicological studies should be encouraged to identify toxic molecules.

In addition, during the period of our field survey, the vast majority of users could not differentiate between the different types of liver diseases (hepatitis, jaundice, and liver cancer) and treated patients based on the characteristic symptoms associated with them (yellowish eye, yellowish body, fever, weakness of the body and anorexia. On the other hand, many patients did not respect the dosage and combined both types of medicine (traditional and modern), which may be another source of toxicity.

## Conclusion

Liver diseases are a crucial public health problem for many industrialized and developing countries. This is due to changes in nutritional habits, lack of adapted dietary measures, alcoholism, and viral or drug-induced infections. In the region of Al-Haouz Rehamna, we conducted an ethnobotanical study to identify the medicinal species used against these diseases. This study reveals that the population of this region, which has accumulated actual know-how on the virtues of medicinal plants, often resorts to traditional medicine to treat liver diseases despite the revolution in medical technology.

In addition, the ethnobotanical surveys that we conducted among the population of this region allowed us to identify 86 species belonging to 37 families and divide them into 79 genera. Among these species, 21 are toxic. The leaf is the most used plant organ, and decoction is the most frequent method of use in traditional phytotherapy in this region. Among the species cited in this region, ten species are reported to be more effective against these pathologies (jaundice, hepatitis, and liver cancer), 4 of which have several citations higher than 300, namely: *Ridolfia segetum*, *Curcuma longa*, *Ononis natrix*, and *Rhamnus alaternus*.

This study aimed to discover other unknown species, know the traditional practices used in the study area, evaluate the risks resulting from certain toxic plants' use, safeguard the local heritage, and develop the Moroccan natural resources. However, it would be interesting to extend this study to other regions of Morocco to establish complete monographs.

Finally, it would be interesting validate the effectiveness of the plants cited in this investigation through suitable testing analyses so that the use of plants is more scientifically established, as well as to pursue and isolate new bioactive molecules that may hold preventative or curative effects against these disorders.

## Declarations

**Ethical Approval:** All participants provided oral prior informed consent.

**Consent to Participate:** Not applicable.

**Consent to Publish:** Not applicable.

**Availability of data and materials:** On request, the relevant author will provide additional material for this article.

**Funding:** This study received no particular support from governmental, private, or not-for-profit funding organizations.

**Competing Interests:** We certify no conflict of interest with any financial organization regarding the manuscript.

**Authors Contributions:** OB: methodology, data collection, a compilation of literature sources, data analysis, assessment, interpretation, realization manuscript HK: Assisting with data and contributing significantly to data analysis. NC: analyzed the data and wrote the article, review and editing, LZ: Design for searching and identifying plant species. RWB: thorough revision of the manuscript and preparation of the final draft. The final paper was understood and approved by all the authors.

## Acknowledgments

The authors thank all the guides and inhabitants of the Al-Haouz-Rehamna region for their help. To all sellers of medicinal plants. The authors also acknowledge all those who participated in the achievement of this product.

## Literature Cited

Abbou A, Kadri N, Debbache N, Dairi S, Remini H, Dahmoune F, Berkani F, Adel K, Belbah A, Madani K. 2019. Effect of precipitation solvent on some biological activities of polysaccharides from *Pinus halepensis* Mill. seeds. International Journal of Biological Macromolecules 141:663-670. doi: 10.1016/j.ijbiomac.2019.08.266

Abdel-Wahhab GED, El-Shamy K, El-Beih AK, Morcy AF, Abd F, Mannaa AE. 2011. Protective effect of a natural herb (*Rosmarinus officinalis*) against hepatotoxicity in male albino rats. Comunicata Scientiae 2(1):9-17. www.ufpi.br/comunicata

Abdulmalek SA, Fessal M, El-Sayed M. 2021. Effective amelioration of hepatic inflammation and insulin response in high fat diet-fed rats via regulating AKT/mTOR signaling: Role of *Lepidium sativum* seed extracts. Journal of Ethnopharmacology 266(June 2020)113439:1-17. doi: 10.1016/j.jep.2020.113439

Abourida A. 2007 Approche hydrogéologique de la nappe du Haouz (Maroc) par Teledetection, isotopie, sig et modelisation. Thèse de Doctorat, Faculte des Sciences, Semlaliya. Universite Cadi Ayyad Marrakech, p. 146.

- Adefegha SA, Oboh G, Adefegha OM, Henle T. 2016. Alligator pepper/Grain of Paradise (*Aframomum melegueta*) modulates Angiotensin-I converting enzyme activity, lipid profile, and oxidative imbalances in a rat model of hypercholesterolemia. *Pathophysiology* 23(3):191-202. doi: 10.1016/j.pathophys.2016.05.005
- Afendy A, Kallman JB, Stepanova M, Younoszai Z, Aquino RD, Bianchi G, Marchesini G, Younossi ZM. 2009. Predictors of health-related quality of life in patients with chronic liver disease. *Alimentary Pharmacology and Therapeutics* 30(5):469-476. doi: 10.1111/j.1365-2036.2009.04061.x
- Aghraz A, Wanner J, Schmidt E, Aitdra L, Aitsidibrahim M, Tabanca N, Ali A, Nafis A, Hassani L, Markouk M, Jirovetz L, Larhsini M. 2017. Chemical composition, in vitro antioxidant, antimicrobial and insecticidal activities of essential oil from *Cladanthus arabicus*. *Journal of Essential Oil-Bearing Plants* 20(3):601-609. doi: 10.1080/0972060X.2017.1331143
- Ahmad F, Khan GM. 2012. Study of aging and hepatoprotective activity of *Vitis vinifera* L. seeds in albino rats. *Asian Pacific Journal of Tropical Biomedicine* 2(3):S1770-S1774. doi: 10.1016/S2221-1691(12)60492-4
- Ahmad M, Qureshi R, Arshad M, Ajab Khan M, Zafar M. 2009. Traditional herbal remedies used for the treatment of diabetes from district Attock (Pakistan). *Pakistan Journal of Botany* 41(6):2777-2782.
- Ahmad Nadzri FN, Tawalbeh D, Sarbon NM. 2021. Physicochemical properties and antioxidant activity of enzymatic hydrolysed chickpea (*Cicer arietinum* L.) protein as an influence by alcalase and papain enzyme. *Biocatalysis and Agricultural Biotechnology* 36.102131:1-8. doi: 10.1016/j.bcab.2021.102131
- Aissous I, Benrebai M, Cacan E, Caglar B, Erenler R, Ameddah S, Benayache S, Benayache F, Bensouici C. 2021. Antioxidant and antiproliferative activities of the n-butanol extract of *Centaurea maroccana* Ball aerial parts. *Current Issues in Pharmacy and Medical Sciences* 34(1):5-11. doi: 10.2478/cipms-2021-0002
- Ait El Cadi M, Makram S, Ansar M, Khabbal Y, Alaoui K, Faouzi MA, Cherrah Y, Taoufik J. 2012. Activité anti-inflammatoire des extraits aqueux et éthanolique de *Zygophyllum gaetulum*. *Annales Pharmaceutiques Françaises* 70(2):113-116. doi: 10.1016/j.pharma.2011.11.004
- Ait El Mekki O. 2017. Spatialisation du potentiel de recharge diffuse d'un aquifère libre sous climat semi-aride par techniques géospatiales et hydrochimiques : cas de l'aquifère du Haouz (Marrakech, Maroc). Thèse de Doctorat, Faculté des Sciences, Semlalia, Université Cadi Ayyad, Marrakech, Maroc, p. 181.
- Ajebli M, Eddouks M. 2019. Flavonoid-enriched extract from desert plant *Warionia saharae* improves glucose and cholesterol levels in diabetic rats. *Cardiovascular & Hematological Agents in Medicinal Chemistry* 17(1):28-39. doi: 10.2174/1871525717666190121143934
- Ben Akka F, El-Hilah FE, Benkhnigue O, Salhi S, Zidane L. 2015. Etude ethnobotanique des plantes médicinales Dans la province de Khouribga (Région d'Oum erbai) Ethnobotany study of medicinal plants In the province of Khouribga (Region Oum erbai). *ScienceLib Editions Mersenne* 7:150504.
- Akther N, Shawl AS, Sultana S, Chandan BK, Akhter M. 2013. Hepatoprotective activity of *Marrubium vulgare* against paracetamol-induced toxicity. *Journal of Pharmacy Research* 7(7):565-570. doi: 10.1016/j.jopr.2013.06.023
- Al-Asmari AK, Al-Elaiwi AM, Athar MT, Tariq M, Al Eid A, Al-Asmary SM. 2014. A review of hepatoprotective plants used in Saudi traditional medicine. *Hindawi Publishing Corporation, Evidence-Based Complementary and Alternative Medicine* 890842:1-22. doi: 10.1155/2014/890842
- Alilou H, Akssira M. 2021. Chemical composition, antibacterial, antioxidant, and insecticidal activities of Moroccan *Tapsia transtagana* essential oil. *Saudi Journal of Biological Sciences* 28(12):6756-6764. doi: 10.1016/j.sjbs.2021.07.052
- Amessis-Ouchemoukh N, Ouchemoukh S, Meziant N, Idiri Y, Hernanz D, Stinco CM, Rodríguez-Pulido FJ, Heredia FJ, Madani K, Luis J. 2017. Bioactive metabolites involved in the antioxidant, anticancer and anticalpain activities of *Ficus carica* L., *Ceratonia siliqua* L. and *Quercus ilex* L. extracts. *Industrial Crops and Products* 95:6-17. doi: 10.1016/j.indcrop.2016.10.007
- APG. 2016. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. *Botanical Journal of the Linnean Society* 181(1):1-20. doi: 10.1111/BOJ.12385

Bajalan I, Rouzbahani R, Pirbalouti AG, Maggi F. 2017. Antioxidant and antibacterial activities of the essential oils obtained from seven Iranian populations of *Rosmarinus officinalis*. Industrial Crops and Products 107:305-311. doi: 10.1016/j.indcrop.2017.05.063

Bammi J, Douira A. 2002. Les plantes médicinales dans la forêt de l'Achach (plateau central, Maroc). Acta Botanica Malacitana 27:131-145.

Bardaweel SK, Hudaib MM, Tawaha KA, Bashatwah RM. 2015. Studies on the in vitro antiproliferative, antimicrobial, antioxidant, and acetylcholinesterase inhibition activities associated with chrysanthemum coronarium essential oil. Hindawi Publishing Corporation, Evidence-Based Complementary and Alternative Medicine, 790838:1-6. doi: 10.1155/2015/790838

Beeby E, Magalhães M, Lemos MFL, Pires IM, Cabral C. 2021. Cytotoxic effects of *Ridolfia segetum* (L.) Moris phytoproducts in cancer cells. Journal of Ethnopharmacology 267(113515):1-2. doi: 10.1016/j.jep.2020.113515

Beevi SS, Mangamoori LN, Gowda BB. 2012. Polyphenolics profile and antioxidant properties of *Raphanus sativus* L. Natural Product Research 26(6): 557-563. doi: 10.1080/14786419.2010.521884

Belhaj S, Chaachouay N, Zidane L. 2021. Ethnobotanical and toxicology study of medicinal plants used for the treatment of diabetes in the High Atlas Central of Morocco. Journal of Pharmacy & Pharmacognosy Research 9(5):619-662.

Bellakhdar J.(1997). La pharmacopée marocaine traditionnelle. Médecine arabe ancienne et savoirs populaires. Le Fennec (ed), Paris, p. 764.

Bellakhdar J. 2006. Plantes médicinales au Maghreb et soins de base, précis de phytothérapie moderne. Le Fennec (ed), Casablanca, p. 385.

Ben Akka F, El Hilah F, Benkhnigue O, Salhi S, Zidane L. 2015. Etude ethnobotanique des plantes médicinales Dans la province de Khouribga (Région d'Oum Rbai). ScienceLib Editions Mersenne 7(150504):1-28.

Benabid A. 2000. Flore et écosystèmes du Maroc : évaluation et préservation de la biodiversité. Ibis Press (ed), Paris, p. 359.

Bencheikh N, Bouhrim M, Merrouni IA, Boutahiri S, Kharchoufa L, Addi M, Tungmunnithum D, Hano C, Eto B, Legssyer A, Elachouri M. 2021. Antihyperlipidemic and antioxidant activities of a flavonoid-rich extract of *Ziziphus lotus* (L.) Lam. fruits. Applied Sciences (Switzerland) 11 (7788):1-13. doi: 10.3390/app11177788

Benkhnigue O, Chaachouay N, Khamar H, El-Azzouzi F, Douira A, Zidane L. 2022. Ethnobotanical and ethnopharmacological study of medicinal plants used in the treatment of anemia in the region of Al-Haouz Rehamna (Morocco). Journal of Pharmacy & Pharmacognosy Research 10(2):279-302.

Benkhnigue O, Hachi M, Fadli M, Douira A, Zidan L. 2016. Catalog of the medicinal plants used in the treatment of urinary infections in the area of Al-Haouz Rhamna (Central Morocco). European Journal of Botany, Plant Sciences and Phytology 3(1):1-49.

Benkhnigue O, Zidane L, Fadli M, Elyacoubi H, Rochdi A, Douira A. 2010. Etude ethnobotanique des plantes médicinales dans la région de Mechraâ Bel Ksiri (Région du Gharb du Maroc). Acta Botanica Barcinonensis 53:191-216.

Benn M, Gul W. 2007. Pyrrolizidine alkaloids in the antipodean genus *Brachyglottis* (Asteraceae). Biochemical Systematics and Ecology 35(10):676-681. doi: 10.1016/j.bse.2007.05.003

Berdja S, Boudarene L, Smail L, Neggazi S, Boumaza S, Sahraoui A, Haffaf EM, Kacimi G, Aouichat Bouguerra S. 2021. *Scolymus hispanicus* (Golden Thistle) ameliorates hepatic steatosis and metabolic syndrome by reducing lipid accumulation, oxidative stress, and inflammation in rats under hyperfatty diet. Evidence-Based Complementary and Alternative Medicine 5588382:1-14. doi: 10.1155/2021/5588382

Berroukche A, Kahloula K, Slimani M, Denai I, Ammour K. 2015. Hepatoprotective effects of the decoction and macerated leaves of *Rhamnus alaternus* L. On rats exposed to carbon tetrachloride. Journal of Pharmacognosy and Phytotherapy 7(10):253-262. doi: 10.5897/JPP2015.0370

Billaux P, Bayssine G. 1967. Les sols du Maroc. In Les Cahiers De La Recherche Agronomique, (pp. 59-101).

- Bouabid K, Lamchouri F, Toufik H, Faouzi MEA. 2020. Phytochemical investigation, in vitro and in vivo antioxidant properties of aqueous and organic extracts of toxic plant: *Atractylis gummifera* L. Journal of Ethnopharmacology 253 (112640):1-11. doi: 10.1016/j.jep.2020.112640
- Bouayyadi L, El Hafian M, Zidane L. 2015. Étude floristique et ethnobotanique de la flore médicinale dans la région du Gharb, Maroc. Journal of Applied Biosciences 93(1):8760-8769 doi: 10.4314/jab.v93i1.10
- Boudjelal A, Henchiri C, Sari M, Sarri D, Hendel N, Benkhaled A, Ruberto G. 2013. Herbalists and wild medicinal plants in M'Sila (North Algeria): An ethnopharmacology survey. Journal of Ethnopharmacology 148(2):395-402. doi: 10.1016/j.jep.2013.03.082
- Bouhache M, Ezzahiri B, Houmy K, Rzozi SB, Taleb A. 2002. Protection phytosanitaire des céréales. IAV Hassan II. Rabat, p. 22.
- Boukhenoufa A, Benmagnhnia S, Maizi Y, Meddah A, Touil T, & Meddah B. 2021. Antifungal and antioxidant activities of *Artemisia herba-alba* Asso. European Journal of Biological Research Research Article European Journal of Biological Research 11(4):493-500. doi: 10.5281/zenodo.5552721
- Bouyahya A, El Omari N, Elmeniyi N, Guouguaou FE, Balahbib A, El-Shazly M, Chamkhi I. 2020. Ethnomedicinal use, phytochemistry, pharmacology, and toxicology of *Ajuga iva* (L.) Schreb. Journal of Ethnopharmacology 258(112875):1-20. doi: 10.1016/j.jep.2020.112875
- Bruneton J. 1999. Pharmacognosie - Phytochimie Plantes médicinales (3th ed.). Paris: Lavoisier, p. 1120.
- Bruneton J. 2009. Pharmacognosie - Phytochimie Plantes médicinales (4th ed.). Paris: Lavoisier, p. 1270.
- Cadena-González AL, Sørensen M, Theilade I. 2013. Use and valuation of native and introduced medicinal plant species in Campo Hermoso and Zetaquira, Boyacá, Colombia. Journal of Ethnobiology and Ethnomedicine 9 (1):1-14. doi: 10.1186/1746-4269-9-23
- Cavallar W. 1950. Etude des sols des différentes régions du Maroc. Société des Sciences Naturelles du Maroc, Travaux de la Section de Podologie. 1:19-45.
- Çevik D, Burçin Yılmazgöz, Kan Y, Akhan Güzelcan E, Durmaz I, Çetin-Atalay R, Kırmızıbekmez H. 2018. Bioactivity-guided isolation of cytotoxic secondary metabolites from the roots of *Glycyrrhiza glabra* and elucidation of their mechanisms of action. Industrial Crops and Products 124:389-396. doi: 10.1016/j.indcrop.2018.08.014
- Chaachouay N, Benkhnigue O, Zidane L. 2020. Ethnobotanical Study Aimed at Investigating the Use of Medicinal Plants to Treat Nervous System Diseases in the Rif of Morocco. Journal of Chiropractic Medicine 19(1):70-81. doi: 10.1016/j.jcm.2020.02.004
- Chaudhari MV. 2013. Aragvadha (*Cassia fistula* Linn.): A Phyto-Pharmacological Review. Journal of Ayurveda and Holistic Medicine 1(7):22-29.
- Chaves N, Santiago A, Alías JC. 2020. Quantification of the antioxidant activity of plant extracts: Analysis of sensitivity and hierarchization based on the method used. Antioxidants 9(76):1-14. doi: 10.3390/antiox9010076
- Da-Costa-Rocha I, Bonnlaender B, Sievers H, Pischel I, Heinrich M. 2014. *Hibiscus sabdariffa* L. A phytochemical and pharmacological review. Food Chemistry 165:424-443. doi: 10.1016/j.foodchem.2014.05.002
- Darvesh AS, Aggarwal BB, Bishayee A. 2012) Curcumin and Liver Cancer: A Review. Current Pharmaceutical Biotechnology 13(1):218-28. doi: 10.2174/138920112798868791.
- Deghima A, Righi N, Rosales-Conrado N, León-González ME, Gómez-Mejía E, Madrid Y, Baali F, Bedjou F. 2020. Bioactive polyphenols from *Ranunculus macrophyllus* Desf. Roots: Quantification, identification, and antioxidant activity. South African Journal of Botany 132:204-214. doi: 10.1016/j.sajb.2020.03.036
- Deng LN, Feng GN, Gao Y, Shen YX, Li HS, Gu Y, Luan HY. 2020. Phytochemical constituents and antioxidant enzyme activity profiles of different barley (*Hordeum vulgare* L.) cultivars at different developmental stages. Agronomy 10(37):1-11. doi: 10.3390/agronomy10010037
- Dhatchayani S, Vijayakumar S, Sarala N, Vaseeharan B, Sankaranarayanan K. 2020. Effect of curcumin sorbed selenite substituted hydroxyapatite on osteosarcoma cells: An in vitro study. Journal of Drug Delivery Science and Technology 60(101963):1-8. doi: 10.1016/j.jddst.2020.101963

- Dobignard A, Chatelain C. 2013. Index synonymique de la Flore d'Afrique du Nord volume 5: Dicotyledonae, Oleaceae à Zygophyllaceae. Vol 5. Conservatoire et Jardin botaniques de la Ville de Genève, ECWP, hors-série (11ed). Genève, 451p.
- Donia AE, Soliman GA, El-Sakhawy MA, Yusufoglu H, Zaghloul AM. 2014. Cytotoxic and antimicrobial activities of *Emex spinosa* (L.) Campd. extract. Pakistan Journal of Pharmaceutical Science 27(2):351-6.
- Doudach L, Meddah B, Faouzi M, Khatib AM, Lalou C, Hammani K, Elomri A, Cherrah Y. 2013. Cytotoxic and antioxidant activity of various extracts of *Corrigiola telephylloides* Pourr. International Journal of Pharmacy and Pharmaceutical Sciences 5(3):154-158.
- Eddouks M, Ajebli M, Hebi M. 2017. Ethnopharmacological survey of medicinal plants used in Daraa-Tafilet region (Province of Errachidia), Morocco. Journal of Ethnopharmacology 198:516-530. doi: 10.1016/j.jep.2016.12.017
- El Azzouzi F, Zidane L. 2015. La flore médicinale traditionnelle de la région de Béni- Mellal (Maroc). Journal of Applied Biosciences 91(1) 8493-8502. doi: 10.4314/jab.v91i1.8
- El Bouzidi L. 2013. Contribution à la valorisation et à la protection de quelques plantes aromatiques et médicinales marocaines : *Withania frutescens*, *Achillea ageratum*, *Thymus broussonetii*, *Thymus maroccanus* et *Thymus satureioides*. Thèse de Doctorat, Faculte des Sciences, Semlalia, Universite Cadi Ayyad, Marrakech, Maroc, p. 130.
- El Fakir L, Bouothmany K, Alotaibi A, Bourhia M, Ullah R, Zahoor S, El Mzibri M, Gmouh S, Alaoui T, Zaid A, Benbacer L. 2021. Antioxidant and understanding the anticancer properties in human prostate and breast cancer cell lines of chemically characterized methanol extract from *Berberis hispanica* Boiss. & Reut. Applied Sciences (Switzerland) 11(8)3510:1-12. doi: 10.3390/app11083510
- Elsebai MF, Mocan A, Atanasov AG. 2016. Cynaropicrin: A comprehensive research review and therapeutic potential as an anti-hepatitis C virus agent. Frontiers in Pharmacology 7(472):1-15. doi: 10.3389/fphar.2016.00472
- Emberger L. 1938. Aperçu général sur la végétation du Maroc. Berne, Hans Huber, p. 157.
- Emerenciano VP, Milit JSLT, Campos CC, Romoff P, Kaplan MAC, Zambon M, Brant AJC. 2001. Flavonoids as chemotaxonomic markers for Asteraceae. Biochemical Systematics and Ecology 29:947-957.
- Es-Safi I, Mechchate H, Amaghnouje A, Calarco A, Boukhira S, Noman OM, Mothana RA, Nasr FA, Bekkari H, Bousta D. 2020. Defatted hydroethanolic extract of *Ammodaucus leucotrichus* Cosson and Durieu seeds: Antidiabetic and anti-inflammatory activities. Applied Sciences (Switzerland) 10(24):1-11. doi: 10.3390/app10249147
- Farkondeh T, Samarghandian S. 2019. The therapeutic effects of *Portulaca oleracea* L. in hepatogastric disorders. Gastroenterología y Hepatología (English Edition) 42(2):127-132. doi: 10.1016/j.gastre.2019.02.005
- Farshori NN, Al-Sheddi ES, Al-Oqail MM, Musarrat J, Al-Khedhairy AA, Siddiqui MA. 2013. Anticancer activity of *Petroselinum sativum* seed extracts on MCF-7 human breast cancer cells. Asian Pacific Journal of Cancer Prevention 14(10):5719-5723. doi: 10.7314/APJCP.2013.14.10.5719
- Fennane M, Ibn Tattou M. 2005. Flore vasculaire du Maroc : Inventaire et chorologie. Vol. 1. Série botanique N 37. Rabat: Travailes Institute des Sciences Université Mohamed V, p. 483.
- Fennane M, Ibn Tattou M. 1998. Catalogue des plantes vasculaires rares, menacées ou endémiques du Maroc. Boccone 8(1120-4060):1-279.
- Fennane M, Ibn Tattou M. 2012. Statistiques et commentaires sur l'inventaire actuel de la flore vasculaire du Maroc. Bulletin de l'Institut Scientifique, Rabat, section Sciences de la Vie 34(1):1-9.
- Fennane M, Ibn Tattou M, El Oualidi J. 2014. Flore pratique du Maroc. Vol. 3. Série botanique N 40. Rabat: Travailes Institute des Sciences Université Mohamed 5, p. 793.
- Fennane M, Ibn Tattou M, El oualidi J, Ouyahya A. 2007. Flore pratique du Maroc. Vol. 2. Série botanique N 38. Rabat: Trav. Inst Sci. Université Mohamed V, p. 636.
- Fennane M, Ibn Tattou M, Mathez J, Ouyahya A, El Oualidi J. 1999. Flore pratique du Maroc. Vol. 1. Série botanique N 36. Rabat: Travailes Institute des Sciences Université Mohamed V, p. 558.
- Ferrari B, Tomi F, Casanova J. 2005. Terpenes and acetylene derivatives from the roots of *Santolina corsica* (Asteraceae). Biochemical Systematics and Ecology 33(4): 445-449. doi: 10.1016/j.bse.2004.11.001

Filliat P. 2012. Les plantes de la famille des Apiaceae dans les troubles digestifs. Thèse de Doctorat en pharmacie. Université Joseph Fourier Faculté De Pharmacie De Grenoble, France, p. 129.

Lakmichi H, Bakhtaoui FZ, Gadhi CA, Ezoubeiri A, El Jahiri Y, El Mansouri A, Zrara I, Loutfi K. 2011. Toxicity profile of the aqueous ethanol root extract of *Corrigiola telephifolia* Pourr. (Caryophyllaceae) in Rodents. Hindawi Publishing Corporation. Evidence-Based Complementary and Alternative Medicine 2011:1-10. doi:10.1155/2011/317090

Gakuubi MM, Wanzala W. 2012. A survey of plants and plant products traditionally used in livestock health management in Buuri district, Meru County, Kenya. Journal of Ethnobiology and Ethnomedicine, 8(39): 1-19. doi: 10.1186/1746-4269-8-39

Abdel-Wahhab GEDK, El-Shamy AK, El-Beih AN, Moryc AF, Abd F, Mannaa AEF. 2011. Protective effect of a natural herb (*Rosmarinus officinalis*) against hepatotoxicity in male albino rats. Comunicata Scientiae 2(1):9-17. www.ufpi.br/comunicata

Gatsing D, Aliyu R, Kuiate JR, Garba IH, Jaryum KH, Tedongmo N, Tchouanguep FM, Adoga GI. 2005. Toxicological evaluation of the aqueous extract of *Allium sativum* L. bulbs on laboratory mice and rats. In Cameroon Journal of Experimental Biology 1(1):39-45. doi: 10.4314/cajeb.v1i1.37926

Ghlissi Z, Kallel R, Krichen F, Hakim A, Zeghal K, Boudawara T, Bougatet A, Sahnoun Z. 2020. Polysaccharide from *Pimpinella anisum* seeds: Structural characterization, anti-inflammatory and laser burn wound healing in mice. International Journal of Biological Macromolecules 156:1530-1538. doi: 10.1016/j.ijbiomac.2019.11.201

Ghourri M, Zidane L, El Yacoub H, Rochdi A, Fadli M, Douira A. 2012. Floristic and ethnobotanical study of medicinal plants of El Ouatia (Morocco). Kastamonu Univ., Journal of Forestry Faculty 12(2):218-235.

Godron M, Daget P. 1982. Analyse fréquentielle de l'écologie des espèces. Collection Ecologia mediterranea, 18 Masson, p. 163.

Godron M. 1971. Essai sur une approche probabiliste de l'écologie des végétations. Thèse Doct. D'Etat, U.S.T.L., Montpellier, p. 247.

Golshahi H, Araghi A, Baghban F, Farzad-Mohajeri S. 2019. Protective effects of 2-methoxycinnamaldehyde, an active ingredient of *Cinnamomum cassia*, on warm hepatic ischemia/ reperfusion injury in a rat model. Iranian Journal of Basic Medical Sciences 22(12):1400-1407. doi: 10.22038/ijbms.2019.13987

Hachi M, Hachi T, Belahbib N, Dahmani J, Zidane L. 2015. Contribution à l'étude floristique et ethnobotanique de la flore medicinale utilisée au niveau de la ville de Khenifra (Maroc). International Journal of Innovation and Applied Studies 11(3):754-770.

Ben-Hamouda A, Chaieb I, Zouari L, Zarrad K, Laarif A. 2016. Toxicological effects of *Urginea maritima* (L.) against the red flour beetle (Coleoptera: Tenebrionidae). Journal of Entomology and Zoology Studies 4(1):17-20.

Han X, Parker TL. 2017. Anti-inflammatory activity of clove (*Eugenia caryophyllata*) essential oil in human dermal fibroblasts. Pharmaceutical Biology 55(1):1619-1622. doi: 10.1080/13880209.2017.1314513

HCP. 2014. Recensement général de la population et de l'habitat. Monographie de la région Marrakech-Safi. Direction régionale de Marrakech-Safi. Rabat, p. 171.

Hegazy M, Elhalfawy I, Shaban M, Abouelnour E. 2018. Potential protective role of *Nigella sativa* Linn against Sildenafil induced hepatotoxicity and nephrotoxicity in adult male rabbits. Ain Shams Journal of Forensic Medicine and Clinical Toxicology, 30(1): 101-109. doi: 10.21608/ajfm.2018.18200

Sbiai-Juimili H, Fadli A, Zidane L. 2017. Survey of ethnomedicinal plants used for the treatment of gastrointestinal disorders in Seksoua Region (Western high Moroccan Atlas). Annual Research and Review in Biology 16(5):1-9. doi: 10.9734/ARRB/2017/36112

Hmamouchi M. 1999. Les Plantes médicinales et aromatiques Marocaines : Utilisations, biologie, écologie, chimie, pharmacologie, toxicologie. Mohammed V. Rabat, p. 389.

Hseini S. 2008. Etude ethnobotanique de la flore médicinale dans la région de Rabat. Thèse de Doctorat, Faculte des Sciences Universite Mohamed V. Rabat, p. 173.

- Hseini S, Kahouadji A. 2007. Étude ethnobotanique de la flore médicinale dans la région de Rabat (Maroc occidental). *Lazaroa* 28:79-93. doi: 10.5209/LAZAROA.9741
- Hudaib M, Mohammad M, Bustanji Y, Tayyem R, Yousef M, Abuirjeie M, Aburjai T 2008. Ethnopharmacological survey of medicinal plants in Jordan, Mujib Nature Reserve and surrounding area. *Journal of Ethnopharmacology* 120(1):63-71. doi: 10.1016/j.jep.2008.07.031
- Ibn Tattou M, Fennane M. 2008. Flore vasculaire du Maroc : Inventaire et chorologie. Vol. 2. Série botanique N 39. Raba: Travailes Institut Sciences Université Mohamed 5, p. 398.
- Inta A, Trisonthi P, Trisonthi C. 2013. Analysis of traditional knowledge in medicinal plants used by Yuan in Thailand. *Journal of Ethnopharmacology* 149(1):344-351. doi: 10.1016/j.jep.2013.06.047
- Ismaili H, Milella L, Fkikh-Tetouani S, Ilidrissi A, Camporese A, Sosa S, Altinier G, Della Loggia R, Aquino R. 2004. In vivo topical anti-inflammatory and in vitro antioxidant activities of two extracts of *Thymus satureioides* leaves. *Journal of Ethnopharmacology* 91(1):31-36. doi: 10.1016/j.jep.2003.11.013
- Jameel M, Ali A, Ali M. 2015. Isolation of antioxidant phytoconstituents from the seeds of *Lens culinaris* Medik. *Food Chemistry* 175:358-365. doi: 10.1016/j.foodchem.2014.11.130
- Janakat S, Al-Merie H. 2002. Evaluation of hepatoprotective effect of *Pistacia lentiscus*, *Phillyrea latifolia*, and *Nicotiana glauca*. *Journal of Ethnopharmacology* 83(1-2):135-138. doi: 10.1016/s0378-8741(02)00241-6
- Jasmine R, Manikandan K, Karthikeyan. 2015. Evaluating the antioxidant and anticancer property of *Ficus carica* fruits. *African Journal of Biotechnology* 14(7):634-641. doi: 10.5897/ajb2014.13742
- Kaviarasan S, Naik GH, Gangabhirathi , Anuradha CV, Priyadarsini KI. 2007. In vitro studies on antiradical and antioxidant activities of fenugreek (*Trigonella foenum graecum*) seeds. *Food Chemistry* 103(1):31-37. doi: 10.1016/j.foodchem.2006.05.064
- Knippertz P, Christoph M, Speth P. 2003. Long-term precipitation variability in Morocco and the link to the large-scale circulation in recent and future climates. *Meteorology and Atmospheric Physics* 83(1-2):67-88. doi: 10.1007/s00703-002-0561-y
- Lahsissène H. 2010. Recherches ethnobotanique & floristique des plantes médicinales utilisées dans la région phytogéographique de Zaër. Thèse de doctorat, Faculté des Science Université Mohamed 5, Rabat, p. 258.
- Lin J, Puckree T, Mvelase TP. 2002. Anti-diarrhoeal evaluation of some medicinal plants used by Zulu traditional healers. *Journal of Ethnopharmacology* 79(1):53-56. doi: 10.1016/s0378-8741(01)00353-1
- Ljubuncic P, Azaizeh H, Portnaya I, Cogan U, Said O, Saleh KA, Bomzon A. 2005. Antioxidant activity and cytotoxicity of eight plants used in traditional Arab medicine in Israel. *Journal of Ethnopharmacology* 99(1):43-47. doi: 10.1016/j.jep.2005.01.060
- Longo L, Platini F, Scardino A, Alabiso O, Vasapollo G, Tessitore L. 2008. Autophagy inhibition enhances anthocyanin-induced apoptosis in hepatocellular carcinoma. *Molecular Cancer Therapeutics* 7(8):2476-2485. doi: 10.1158/1535-7163.MCT-08-0361
- Maameri Z, Djerrou Z, Halmi S, Djaalab H, Riachi F, Hamdipacha Y. 2015. Evaluation of hepatoprotective effect of *Pistacia lentiscus* L. fatty oil in rats intoxicated by carbon tetrachloride. *International Journal of Pharmacognosy and Phytochemical Research* 7(2):251-254.
- Mandegary A, Pournamdar M, Sharififar F, Pourourmohammadi S, Fardiar R, Shooli S. 2012. Alkaloid and flavonoid rich fractions of fenugreek seeds (*Trigonella foenum-graecum* L.) with antinociceptive and anti-inflammatory effects. *Food and Chemical Toxicology* 50(7):2503-2507. doi: 10.1016/j.fct.2012.04.020
- Martins N, Barros L, Santos-Buelga C, Ferreira ICFR. 2016. Antioxidant potential of two Apiaceae plant extracts: A comparative study focused on the phenolic composition. *Industrial Crops and Products* 79:188-194. doi: 10.1016/j.indcrop.2015.11.018
- Masri W, Hedhili A, Amamou M. 2009. *Attractylis gummifera* L. poisoning: Report of two clinical cases. *Revue Francophone Des Laboratoire* (413):87-91. doi: 10.1016/S1773-035X(09)74252-2

Ould El Hadj M, Hadj-Mahammed M, Zabeirou H, Chehma A. 2003. Importance des plantes spontanées Medicinales Dans La Pharmacopee Traditionnelle De La Region De Ouargla (Sahara septentrional - Est algérien). Sciences & Technologie C 20:73-78.

Mehenni C, Atmani-Kilani D, Dumarçay S, Perrin D, Gérardin P, Atmani D. 2016. Hepatoprotective and antidiabetic effects of *Pistacia lentiscus* leaf and fruit extracts. Journal of Food and Drug Analysis 24(3):653-669. doi: 10.1016/j.jfda.2016.03.002

Mejri F, Baati T, Martins A, Selmi S, Luisa Serralheiro M, Falé PL, Rauter A, Casabianca H, Hosni K. 2020. Phytochemical analysis and in vitro and in vivo evaluation of biological activities of artichoke (*Cynara scolymus* L.) floral stems: Towards the valorization of food by-products. Food Chemistry 333(127506):1-10. doi: 10.1016/j.foodchem.2020.127506.

Miara MD, Bendif H, Ouabed A, Rebbas K, Ait Hammou M, Amirat M, Greene A, Teixidor-Toneu I. 2019. Ethnoveterinary remedies used in the Algerian steppe: Exploring the relationship with traditional human herbal medicine. Journal of Ethnopharmacology 244 112164:1-13. doi: 10.1016/j.jep.2019.112164.

Miara MD, Souidi Z, Benhanifa K, Daikh A, Hammou MA, Moumenine A, Sabi IH. 2021. Diversity, natural habitats, ethnobotany, and conservation of the flora of the Macta marches (North-West Algeria). International Journal of Environmental Studies 78(5):817-837. doi: 10.1080/00207233.2020.1824867.

Michel L, Giorgi R, Villes V, Poizot-Martin I, Dellamonica P, Spire B, Protopopescu C, Carrieri MP. 2009. Withdrawal symptoms as a predictor of mortality in patients HIV-infected through drug use and receiving highly active antiretroviral therapy (HAART). Drug and Alcohol Dependence 99(1-3):96-104. doi: 10.1016/j.drugalcdep.2008.07.007

MGRMS. 2015. Monographie générale la région de Marrakech-Safi. Ministère de l'Intérieur, Direction Générale des Collectivités Locales. Rabat, p. 58.

Donia AERM. 2014. Biological Activity of *Chrysanthemum coronarium* L. Extracts. Annual Research & Review in Biology 4(16),2617-2627. doi: 10.9734/ARRB/2014/10112.

Mohammad MA, Al-Taee SK, Al-Jumaa ZM. 2021. Effect addition of *Cinnamomum cassia* on the treatment of pathological infections in *Cyprinus carpio* L. Fingerlings. Iraqi Journal of Veterinary Sciences 35(4):733-738. doi: 10.33899/ijvs.2021.128258.1564.

Mokdad AA, Lopez AD, Shahraz S, Lozano R, Mokdad AH, Stanaway J, Murray CJL, Naghavi M. 2014. Liver cirrhosis mortality in 187 countries between 1980 and 2010: A systematic analysis. BMC Medicine 12(145):1-24. doi: 10.1186/s12916-014-0145-y.

Mosbah H, Ben-Sassi A, Chahdoura H, Snoussi M, Flaminio G, Achour L, Selmi B. 2020. Antioxidant, antimicrobial, and phytotoxic activities of *Rhaponticum acaule* DC. Essential oil. Brazilian Journal of Pharmaceutical Sciences 56(e18483):1-11. doi: 10.1590/S2175-97902019000318483.

Najem M, Belaïdi R, Slimani I, Bouiamrine EH, Ibjibjen J 2018. Traditional pharmacopeia of the Zerhoun region - Morocco: ancestral knowledge and risks of toxicity. International Journal of Biological and Chemical Science 12(6):2797-2807. doi: 10.4314/ijbcs.v12i6.25.

Negre R. 1959. Recherches phytogéographiques sur l'étage de végétation méditerranéen aride (sous étage chaud) au Maroc occidental. (Issue 13). Travails Institute Scientifique Chérifien, Série Botanique N. 13.

Negre R. 1961-1962. Petite flore des régions arides du Maroc occidental. Tome 1 et 2. (CNRS). 979p

Neha M, Deepshikha PK, Vidhu A, Amitesh K, Vidushi J, Alka M, Ritu V. 2014. Determination of antioxidant and hepatoprotective ability of flavanoids of *Cichorium intybus*. International Journal of Toxicological and Pharmacological Research, 6(4): 107-112.

Nejatbakhsh F, Karegar-Borzi H, Amin G, Eslaminejad, A, Hosseini M, Bozorgi M, Gharabaghi MA. 2017. Squill Ozymel, a traditional formulation from *Drimia maritima* (L.) Stearn, as an add-on treatment in patients with moderate to severe persistent asthma: A pilot, triple-blind, randomized clinical trial. Journal of Ethnopharmacology 196:186-192. doi: 10.1016/j.jep.2016.12.032

Notas G, Kisseeleva T, Brenner D. 2009. NK and NKT cells in liver injury and fibrosis. In Clinical Immunology 130(1):16-26. doi: 10.1016/j.clim.2008.08.008

- Otunola GA, Afolayan AJ, Ajayi EO, Odeyemi SW. 2017. Characterization, antibacterial and antioxidant properties of silver nanoparticles synthesized from aqueous extracts of *Allium sativum*, *Zingiber officinale*, and *Capsicum frutescens*. *Pharmacognosy Magazine* 13(50):S201-S208. doi: 10.4103/pm.pm\_430\_16
- Ouarghidi A, Martin GJ, Powell B, Esser G, Abbad A. 2013. Botanical identification of medicinal roots collected and traded in Morocco and comparison to the existing literature. *Journal of Ethnobiology and Ethnomedicine* 9:59:1-13. <http://www.ethnobiomed.com/content/9/1/59>.
- Ouzir M, El Bairi K, Amzazi S. 2016. Toxicological properties of fenugreek (*Trigonella foenum graecum*). *Food and Chemical Toxicology* 96:145-154. doi: 10.1016/j.fct.2016.08.003
- Ozcan B, Esen M, Kemal Sangun M, Coleri A, Caliskan M. 2010. Effective antibacterial and antioxidant properties of methanolic extract of *Laurus nobilis* seed oil. In *Journal of Environmental Biology* 31(5):637-641.
- Pandey A, Bigoniya P, Raj V, Patel KK. 2011. Pharmacological screening of *Coriandrum sativum* Linn. for hepatoprotective activity. *Journal of Pharmacy and Bioallied Sciences* 3(3):435-441. doi: 10.4103/0975-7406.84462
- Perelman B, Kuttin ES. 1988. Parsley-induced photosensitivity in ostriches and ducks. *Avian Pathology* 17(1):183-192.
- Piqué A, El Hassani A, Hoepffner C. 1993. Les déformations ordoviciennes dans la zone des Sehoul (Maroc septentrional) : une orogenèse calédonienne en Afrique du Nord. *Canadian Journal of Earth Sciences* 30(7):1332-1337. doi: 10.1139/e93-114.
- Pradeep K, Victor C, Mohan R, Gobianand K, Karthikeyan S. 2010. BR Protective effect of *Cassia fistula* Linn. on diethylnitrosamine induced hepatocellular damage and oxidative stress in ethanol pretreated rats. *Biological Research* 43(1):113-125. doi: 10.4067/S0716-97602010000100013
- Rabib H, Elagdi C, Hsaine M, Fougrach H, Koussa T, Badri W. 2020. Antioxidant and antibacterial activities of the essential oil of Moroccan *Tetraclinis articulata* (Vahl) Masters. *Hindawi, Biochemistry Research International* 9638548:1-6. doi: 10.1155/2020/9638548
- Raunkiaer C. 1934. *The Life Forms of Plants and Statistical Plant Geography*. London: Clarendon Press, p. 632.
- Rajkumar T, Sapi A, Das G, Debnath T, Ansari AZ, Patra JK. 2019. Biosynthesis of silver nanoparticles using extract of *Zea mays* (corn flour) and investigation of its cytotoxicity effect and radical scavenging potential. *Journal of Photochemistry and Photobiology B: Biology* 193:1-7. doi: 10.1016/j.jphotobiol.2019.01.008
- Ranasinghe P, Pigera S, Premakumara GS, Galappaththy P, Constantine GR, Katulanda P. 2013. Medicinal properties of true Cinnamon (*Cinnamomum zeylanicum*): A systematic review. *BMC Complementary and Alternative Medicine* 13(275):1-10. doi: 10.1186/1472-6882-13-275
- Rimbau V, Cerdan C, Iglesias J. 1999. Anti-inflammatory activity of some extracts from plants used in the traditional medicine of North-African Countries (II). *Phytotherapy Research* 13:128-132. doi: 10.1002/(SICI)1099-1573(199903)13:2<128: AID-PTR399>3.0.CO;2-7
- Rtibi K, Selmi S, Grami D, Amri M, Eto B, El-benna J, Sebai H, Marzouki L. 2017. Chemical constituents and pharmacological actions of carob pods and leaves (*Ceratonia siliqua* L.) on the gastrointestinal tract: A review. *Biomedicine and Pharmacotherapy* 93:522-528. doi: 10.1016/j.biopha.2017.06.088
- Saad F, Mrabti HN, Sayah K, Bouyahya A, Salhi N, Cherrah Y, My El Abbes F. 2019. Phenolic content, acute toxicity of *Ajuga iva* extracts, and assessment of their antioxidant and carbohydrate digestive enzyme inhibitory effects. *South African Journal of Botany* 125:381-385. doi: 10.1016/j.sajb.2019.08.010
- Samia R, Abdennabi EM, Youssef D, Paul B, Ismail C, José T, Alberto G. 2018. 3D geological model of the Eastern Haouz Region (Morocco): Hydrogeological implications. *European Scientific Journal* 14(24):53-68. doi: 10.19044/esj.2018.v14n24p53
- Samojlik I, Lakić N, Mimica-Dukić N, Daković-Švajcer K, Božin B. 2010. Antioxidant and hepatoprotective potential of essential oils of coriander (*Coriandrum sativum* L.) and Caraway (*Carum carvi* L.) (Apiaceae). *Journal of Agricultural and Food Chemistry* 58(15):8848-8853. doi: 10.1021/jf101645n

- Sangare M, Sina H, Dougnon J, Bayala B, Ategbo JM, Dramane K. 2013. Etude ethnobotanique des plantes hépatotropes et de l'usage traditionnel de *Gomphrena celosioides* Mart. (Amaranthaceae) au Bénin. International Journal of Biological and Chemical Sciences 6(6):5008-5021. doi: 10.4314/ijbcs.v6i6.20
- Sanna G, Farci P, Busonera B, Murgia G, la Colla P, Giliberti G. 2015. Antiviral properties from plants of the Mediterranean flora. Natural Product Research 29(22):2065-2070. doi: 10.1080/14786419.2014.1003187
- Saoudi MM, Bouajila J, Rahmani R, Alouani K. 2021. Phytochemical composition, antioxidant, antiacetylcholinesterase, and cytotoxic activities of *Rumex crispus* L. International Journal of Analytical Chemistry 6675436:1-16. doi: 10.1155/2021/6675436
- Sayari N, Najib Saidi M, Sila A, Ellouz-Chaabouni S, Bougatef A. 2016. Chemical composition, angiotensin I-converting enzyme (ACE) inhibitory, antioxidant and antimicrobial activities of *Ononis natrix* leaves extracts. Free Radicals and Antioxidants 6(1):23-33. doi: 10.5530/fra.2016.1.3
- Seddighfar M, Mirghazanfari S, Dadpay M. 2020. Analgesic and anti-inflammatory properties of hydroalcoholic extracts of *Malva sylvestris*, *Carum carvi*, or *Medicago sativa*, and their combination in a rat model. Journal of Integrative Medicine 18(2):181-188. doi: 10.1016/j.joim.2020.02.003
- Shafi S, Tabassum N. 2013. Acute oral toxicity and hypoglycaemic study of ethanolic extract of *Portulaca oleracea* (whole plant) in swiss albino mice. International Journal of Pharmacy and Pharmaceutical Sciences 5(4):389-393.
- Shang A, Cao SY, Xu XY, Gan RY, Tang GY, Corke H, Mavumengwana V, Li HB. 2019. Bioactive compounds and biological functions of garlic (*Allium sativum* L.). Foods 8(246):1-31. doi: 10.3390/foods8070246.
- Shcherbakova A, Strömstedt AA, Göransson U, Gnezdilov O, Turanov A, Boldbaatar D, Kochkin D, Ulrich-Merzenich G, Koptina A. 2021. Antimicrobial and antioxidant activity of *Evernia prunastri* extracts and their isolates. World Journal of Microbiology and Biotechnology 37(129):1-14. doi: 10.1007/s11274-021-03099-y.
- Shebbo S, El Joumaa M, Kawach R, Borjac J. 2020. Hepatoprotective effect of *Matricaria chamomilla* aqueous extract against 1,2-Dimethylhydrazine-induced carcinogenic hepatic damage in mice. Heliyon 6(6) e04082:1-9. doi: 10.1016/j.heliyon.2020.e04082.
- Shen CY, Jiang JG, Zhu W, Ou-Yang Q. 2017. Anti-inflammatory effect of essential oil from *Citrus aurantium* L. var. *amara* Engl. Journal of Agricultural and Food Chemistry 65(39):8586-8594. doi: 10.1021/acs.jafc.7b02586
- Sjelmassi A. 2011. Les plantes médicinales du Maroc. 8ème éd. Le Fennec, Casablanca, p. 285.
- Taban A, Saharkhiz MJ, Niakousari M. 2018. Sweet bay (*Laurus nobilis* L.) essential oil and its chemical composition, antioxidant activity, and leaf micromorphology under different extraction methods. Sustainable Chemistry and Pharmacy 9:12-18. doi: 10.1016/j.scp.2018.05.001
- Tahraoui A, El-Hilaly J, Israili ZH, Lyoussi B. 2007. Ethnopharmacological survey of plants used in the traditional treatment of hypertension and diabetes in south-eastern Morocco (Errachidia province). Journal of Ethnopharmacology 110(1):105-117. doi: 10.1016/j.jep.2006.09.011
- Tahri N, Abdelkrim EL Basti A, Zidane L, Rochdi A, Douira A. 2011. Etude ethnobotanique des plantes medicinales dans la province de Settat (Maroc). Journal of Animal & Plant Sciences 12(3):1632-1652.
- Tahri Y, Koubaa I, Frikha D, Maalej S, Allouche N. 2020. Chemical investigation and biological valorization of two essential oils newly extracted from different parts of *Drimia maritima*. Journal of Essential Oil-Bearing Plants 23(5):1022-1034. doi: 10.1080/0972060X.2020.1854869
- Tra Bi F, Irie G, N'Gaman K, Mahou C. 2008. Études de quelques plantes thérapeutiques utilisées dans le traitement de l'hypertension artérielle et du diabète : deux maladies émergentes en Côte d'Ivoire. Sciences & Nature 5(1):39-48. doi: 10.4314/scinat.v5i1.42150
- Tripathi M, Singh BK, Mishra C, Raisuddin S, Kakkar P. 2010. Involvement of mitochondria-mediated pathways in hepatoprotection conferred by *Fumaria parviflora* Lam. extract against nimesulide induced apoptosis in vitro. Toxicology in Vitro 24(2):495-508. doi: 10.1016/j.tiv.2009.09.011
- Tuseef H, Raza ML, Assad T. 2021. Comparative evaluation of analgesic, antipyretic & anti-inflammatory effects of various extracts of dried fruit of *Illicium verum* Hook. f (star anise) in rodents. Walailak Journal of Science and Technology 18(9)9456:1-12. doi: 10.48048/wjst.2021.9456

- Ullah M, Khan MU, Mahmood A, Malik RN, Hussain M, Wazir SM, Daud M, Shinwari ZK. 2013. An ethnobotanical survey of indigenous medicinal plants in Wana district south Waziristan agency, Pakistan. *Journal of Ethnopharmacology*, 150(3): 918-924. doi: 10.1016/j.jep.2013.09.032
- Uniyal SK, Sharma V, Jamwal P. 2011. Folk Medicinal Practices in Kangra District of Himachal Pradesh, Western Himalaya. *Human Ecology* 39(4):479-488.
- Upadhyay Y, Asselin H, Boon EK, Yadav S, Shrestha KK. 2010. Indigenous use and bio-efficacy of medicinal plants in the Rasuwa District, Central Nepal. *Journal of Ethnobiology and Ethnomedicine* 6(3):1-10.
- Verma SK., Singh SK, Mathur A. 2010. In vitro cytotoxicity of *Calotropis procera* and *Trigonella foenum graecum* against human cancer cell lines. *Journal of Chemical and Pharmaceutical Research* 2(4):861-865.
- Verma A, Masoodi M, Ahmed B. 2012. Lead finding from the whole plant of *Marrubium vulgare* L. with Hepatoprotective Potentials through in silico methods. *Asian Pacific Journal of Tropical Biomedicine* 2(3):S1308-S1311. doi: 10.1016/S2221-1691(12)60406-7
- Hammiche V, Merad R, Azzouz M. 2013. Plantes toxiques à usage médicinal du pourtour méditerranéen. Springer-Verlag France, Paris, p. 408.
- Vitalini S, Iriti M, Puricelli C, Ciuchi D, Segale A, Fic G. 2013. Traditional knowledge on medicinal and food plants used in Val San Giacomo (Sondrio, Italy) - An alpine ethnobotanical study. *Journal of Ethnopharmacology* 145(2):517-529. doi: 10.1016/j.jep.2012.11.024
- Wang Y, Tang C, Zhang H. 2015. Hepatoprotective effects of kaempferol 3-O-rutinoside and kaempferol 3-O-glucoside from *Carthamus tinctorius* L. on CCl<sub>4</sub>-induced oxidative liver injury in mice. *Journal of Food and Drug Analysis* 23(2):310-317. doi: 10.1016/j.jfda.2014.10.002
- Wu S, Yue Y, Tian H, Li Z, Li X, He W, Ding H. 2013. Carthamus red from *Carthamus tinctorius* L. exerts antioxidant and hepatoprotective effect against CCl<sub>4</sub>-induced liver damage in rats via the Nrf2 pathway. *Journal of Ethnopharmacology* 148(2):570-578. doi: 10.1016/j.jep.2013.04.054
- Yogeeta S, Ragavender HRB, Devaki T. 2007. Antihepatotoxic effect of *Punica granatum* acetone extract against isoniazid- and rifampicin-induced hepatotoxicity. *Pharmaceutical Biology* 45(8):631-637. doi: 10.1080/13880200701538963
- Zahir I, Rahmani A. 2020. Premier cas clinique d'eczéma de contact causé par *Tetraclinis articulata*. *International Journal of Innovation and Applied Studies* 28(2):342-346 <http://www.ijias.issr-journals.org/>
- Zarai Z, Kadri A, Ben Chobba I, Ben Mansour R, Bekir A, Mejdoub H, Gharsallah N. 2011. The in-vitro evaluation of antibacterial, antifungal, and cytotoxic properties of *Marrubium vulgare* L. essential oil grown in Tunisia. *Lipids in Health and Disease* 10(161):1-8. doi: 10.1186/1476-511X-10-161