

# From fragrance to ecosystems exploring the traditional medicine and socio-economic-cultural heritage of *Pandanus odorifer* (Forssk.) Kuntze: A review

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## Review

## Abstract

*Background*: *Pandanus odorifer* (Forssk.) Kuntze, known as kewda, is a widely distributed plant species in tropical regions. This article explores the botanical features, traditional applications, and the socio-cultural and economic significance of *P. odorifer*.

*Methods:* The methodology involved an extensive search of peer-reviewed literature, scientific databases, and relevant sources to compile data on diverse attributes of *P. odorifer*.

*Results* :The Paleotropical monocot family Pandanaceae includes approximately 700 species spread across five genera: *Pandanus* Parkinson, *Freycinetia* Gaudich., *Martellidendron* (Pic. Serm.) Callm. & Chassot, *Sararanga* Helms, and *Benstonea* Callm. & Buerki. Of the five genera: three (*Pandanus, Benstonea*, and *Freycinetia*) are found in India. The unique morphology and ecological adaptations of *Pandanus odorifer* contribute to its widespread distribution and resilience. Additionally, it possesses aromatic and medicinal properties. Rich in phytochemicals like lignans, isoflavones, and alkaloids, alongside essential nutrients, it offers therapeutic and nutritional benefits. The oil extracted from its male flowers, containing phenylethyl methyl ether, finds applications in various industries such as aromatherapy, cosmetics, and food. Traditional medicinal practices utilize *P. odorifer* for treating headaches, inflammation, and colds, among other ailments. It also holds cultural significance and is utilized in handicrafts and as a flavoring agent.

*Conclusions: Pandanus odorifer* is a valuable botanical resource with significant ecological, cultural, and economic importance. Its traditional uses and diverse chemical composition demonstrate its multifunctionality. Continued research and conservation are essential to harness its potential for sustainable development and ensure its survival amid environmental changes.

Keywords: Ethnic utility, Kewda, Pandanaceae, Phytochemical constituents, Therapeutic applications

## Background

Plants play a crucial role as major sources of natural products, contributing to numerous industries such as pharmaceuticals, agrochemicals, flavor and fragrance ingredients, food additives, and pesticides (Grigoriadou et al. 2023; Sulieman et al. 2023). Secondary compounds found in plants are unique to each species or group, serving important functions by defending against herbivores, inhibiting competitors, pests, and pathogens, and attracting beneficial organisms like pollinators (Stevenson et al. 2017; Ramawat & Goyal, 2020; Divekar et al. 2022; Al-Khayri et al. 2023; Anjali et al. 2023). Additionally, many of these secondary metabolites are widely utilized in various applications, such as flavorings and pharmaceuticals. One notable plant family in this context is the screw-pine family, Pandanaceae, comprises arborescent or lianoid dioecious monocotyledonous primarily found in paleotropical regions. The family Pandanaceae represents trees, shrubs, lianas, and epiphytes with long, narrow, rigid, spirally arranged pineapple-like leaves and prop roots (Gallaher 2015; Simpson 2019). Members of the family thrive from sea level to elevations around 3000 m on diverse substrates, such as granites, limestones, ultrabasic rocks, and muddy, peaty, or clay soils. Additionally, these plants are commonly associated with littoral mangrove communities and exhibit epiphytic growth in rainforests (Stone et al. 1998). The family holds botanical significance due to its suggested pivotal role in the evolution of monocotyledons (Meeuse 1965; 1966), unusual growth forms (Guillaumet 1973; Tomlinson et al. 1970), and breeding systems (Warburg 1900; Hutchinson 1973; Stone 1968). The center of origin of Pandanaceae remains unconfirmed due to limited fossil records. Audley-Charles (1987) proposed a Gondwana origin during the later Mesozoic period, suggesting migration from eastern Gondwana through Asia and Australia via the Malay Archipelago in the Late Cretaceous period (Nadaf & Zanan 2012a). However, recent molecular studies propose that most Pandanaceae species dispersed via long-distance transport during the late Eocene, after the divergence of the Gondwana continents (Gallaher et al. 2015). Zdravchev et al. (2023) suggest that the order Pandanales likely originated in Laurasia, with its differentiation beginning in the region of Tibet. The Pandanaceae family then diverged in West Africa and subsequently spread eastward into Southeast Asia, northern Australia, and New Zealand. The largest genus, Pandanus, later repopulated Africa from Southeast Asia through long-distance transport (Zdravchev et al. 2023). The family Pandanaceae comprises around 700 species distributed among five genera: Pandanus Parkinson, Freycinetia Gaudich., Martellidendron (Pic. Serm.) Callm. & Chassot, Sararanga Helms, and Benstonea Callm. & Buerki (Stone et al 1998; Callmander et al. 2003; Buerki et al. 2012; Tan & Takayama 2019; Zdravchev et al. 2023).

*Freycinetia* is found in Southeast Asia, the Pacific Islands, and Australasia (Stone 1990). *Martellidendron* is limited to Madagascar and the granitic Seychelles (Callmander 2000, 2001; Callmander *et al.* 2003). *Sararanga* is restricted to the Philippines, New Guinea, and the Solomon Islands (Huynh 2001). *Benstonea* ranges from India to Australia, with a high diversity in Borneo and Peninsular Malaysia (Callmander 2012). Within the Pandanaceae family, the genus *Pandanus* is the most widespread, thriving in tropical and subtropical regions, including the Pacific Islands, Malaysian islands, and Australia (Stone 1974, 1982; Stone *et al.* 1998; Callmander *et al.* 2003, 2012; Buerki *et al.* 2012; Loa *et al.* 2016; Tan & Takayama 2019). *Pandanus* species adapt to diverse habitats, from coastal areas to mountainous regions, providing crucial food and habitat for various animals (Grushvitsky 1982; Frank & Lounibos 1983; Lehtinen 2002; Loa *et al.* 2016; Fusi *et al.* 2021). In many tropical regions, members of the Pandanaceae family are traditionally used for food and medicine (Langenberger *et al.* 2009; Baddu & Ouano 2018). Additionally, they serve as raw materials for woven crafts, ritual items, and construction materials for houses (Wardah 2009; Fedele *et al.* 2011; Ragragio *et al.* 2013; Ordas *et al.* 2021).

The Pandanaceae in India is represented by three genera: *Pandanus, Benstonea,* and *Freycinetia* (Nadaf & Zanan 2012b). In India, about 30 to 40 species of *Pandanus* are found in states including Odisha, Andhra Pradesh, Kerala, Tamil Nadu, West Bengal, Uttar Pradesh, Assam, Meghalaya, Andaman and Nicobar Islands, and Gujarat (Kirtikar *et al.* 1991; Chatterjee & Pakrashi 2001; Padhy *et al.* 2016; Nasim *et al.* 2018). Humans widely use various parts of the *Pandanus*, such as its fruits, leaves, inflorescences, and trunks, for a variety of purposes including food, flavoring, perfume, and fiber (Limbongan *et al.* 2009; Adkar & Bhaskar 2014).

Pandanus odorifer, commonly known as the Screw pine (also referred to as the umbrella tree), is known by various vernacular or local names such as Ketakee, Gandha pushpa, Sthira gandha, Indu Kalika, and Jambala (in Sanskrit); Kewra, Keora, Kewda, and Gagandhul (Hindi); Kedgi, Kevda, and Keora (Marathi); Kewoda (Gujarati); Kiya and Ketakee (Odia); Keya, Kedki, and Keori (Bengali); Kedige, ketake, and tale hu (Kannada); Thazhai, Thalay, and Thazhampoo (Tamil); Mogali and Gajangi (Telugu); Keura, kerada, and tarika Kiora, keura, and kevra (Nepali); Kaitha, Kaida, and Thala (Malayalam); Kiura, kevara, jambala, jambul, panshuka, and ketaki (Urdu); Adan, takonoki (Japanese) and Pandanus aromatnejshi (Russian) (Adkar & Bhaskar 2014; Padhy *et al.* 2016; Savaliram & Zanan 2022). In Odisha, *Pandanus odorifer* thrives in various natural habitats across the coastal districts of Ganjam, Jajpur, Jagatsinghpur, Khurda, Puri, Cuttack, Bhadrak, and Balasore. The plant plays a vital role in local culture, with its parts used for food, flavoring, perfume, and fiber. It serves multiple purposes, offering

aromatic luxury to the affluent and essential sustenance to rural communities, underscoring its ethno-ecological significance. Despite numerous ethnobotanical studies, comprehensive reviews on *P. odorifer*'s traditional uses, socio-cultural impact, and economic significance are lacking. This study aims to fill this gap by documenting and highlighting the integration of *P. odorifer* into local culture, emphasizing its multifaceted importance. Ongoing research and conservation efforts are crucial to unlock the plant's full potential and ensure its long-term survival amid changing environmental dynamics.

## **Materials and Methods**

### Study area

Situated on the eastern coast of India, Odisha (formerly Orissa) lies between the parallels of 17°49'N and 22°34'N latitudes and meridians of 81°27'E and 87°29'E longitudes (Figure 1). Bordered by Jharkhand to the north, Chhattisgarh to the west, Andhra Pradesh to the south, West Bengal to the northeast, and the Bay of Bengal to the southeast, it covers an area of 155,707 km<sup>2</sup>, contributing 4.87% of India's landmass. With the Bay of Bengal forming its eastern and southeastern frontiers, the Eastern Ghats range of hills almost passes through the heart of the state, with high Similipala hills to its north and around 482 km of coastline to its east. Odisha comprises 30 districts housing a total population of 4.2 crores (Census 2011. The populace is distributed across 51,349 villages and 138 urban centers. Notably, the state is home to 62 types of scheduled tribes, constituting 22.85% of the population, and 93 types of scheduled caste communities, making up 17.13% of the populace, primarily dwelling in rural and hilly regions, where they derive sustenance from traditional livelihood practices. Odisha's climate varies across its diverse topography. While the coastal regions experience a moderated climate due to the influence of the sea, with high temperatures typically recorded during April-May, the hill tracts endure more extreme climatic conditions. The vegetation prevalent in the region is primarily of the tropical moist deciduous forest type (Champion & Seth 1968).



Figure 1. Figure 1. (A) Location of Odisha state in the eastern region of India (B) Study area showing different coastal districts

#### Data Analysis

The study focused on the ethnic utility and economic prospects of *P. odorifer* in coastal districts of Odisha, India (Figure 1). To gather information, a comprehensive review of published literature was conducted, and in-depth investigations into its indigenous usage were carried out. Pertinent articles were identified to ensure the validity of the suggested theory, avoid redundancy, and gather sufficient literature for analysis. PubMed, DOAJ, Web of Science, Science Direct (Scopus), and Google Scholar were searched using specific terms such as ethnobotanical study of *Pandanus odorifer*, *P. odoratissimus*, *P. fascicularis*, screw-pines, and Kewda. Studies about the plant's ethnic utility, socio-cultural, and economic prospects were considered in this investigation. Data were gathered on the various parts of the plant used for medicine, food, flavoring, perfume, and fiber, highlighting traditional knowledge and practices documented in the literature. Information on the socio-cultural importance was extracted from studies that explored its role in local customs, rituals, and daily life. The economic potential was assessed by reviewing studies that analyzed commercial uses of products. Articles with full free text access were accessed through various search engines, while requests for full-text access were made directly to authors via research platforms like ResearchGate. Relevant papers were selected for deeper understanding, and only complete texts were considered for analysis, excluding duplicates and unrelated abstracts.

## **Results and Discussion**

Scientific ecological knowledge follows strict rules guided by academic disciplines and the scientific method (Raymond *et al.* 2010). In contrast, local ecological knowledge (LEK) is a collective body of traditional wisdom, practices, and beliefs passed down orally through generations, providing insight into local ecosystems and their management (Cebrian-Piqueras *et al.* 2020). LEK encompasses the understanding of the intricate relationship between humans and their environment, shaped by socio-historical changes within local populations (Reyes-Garcia *et al.* 2007; Fogliarini *et al.* 2021). This study analyzes the LEK of *P. odorifer* across socio-economic and ritual dimensions to understand its importance and utilization within local communities.

#### Habitat and distribution of Pandanus odorifer

*Pandanus odorifer* (Forssk.) Kuntze naturally grows in coastal regions and can withstand salt sprays and strong winds (Rashmi *et al.* 2018). This coastal species is found in various regions (Figure 2) including India, Sri Lanka, the Maldives, southern China, the Ryukyu Islands (Japan), the Philippines, Taiwan, Vietnam, Laos, Cambodia, Thailand, Myanmar, Peninsular Malaysia, Indonesia, Singapore, Borneo, Papua New Guinea, and throughout the Pacific Ocean beaches, including Solomon Islands and Fiji (Callmander *et al.* 2021). Commonly found in coastal districts of Odisha, *Pandanus odorifer* thrives in diverse habitats such as seashores, riverbanks, wastelands, agricultural fields, ponds, canals, and roadsides (Figure 3). Notably, it flourishes along the Ganjam coast, particularly in the Kewda Belt, spanning approximately 675 km<sup>2</sup> from Rushikulya River to Bahuda River. This belt, 45 km long from north to south and 15 km wide from east to west along the Bay of Bengal, hosts around 80-85% of the Kewda canopy within a 10 km radius of the coastline. Flowering persists year-round, with peaks during the rainy season (June-September). Ganjam district alone shelters an estimated 3,00,000 to 4,00,000 trees, producing approximately 1,00,00,000 flowers daily, sustaining a rich habitat for various flora and fauna.



Figure 2. Worldwide distribution of Pandanus odorifer



Figure 3 a-f. Different habitats of *Pandanus odorifer* (Forssk.) Kuntze a. riverbanks, b. near canal, c. near pond, d. wastelands, e. roadsides, f. demarcation of crop fields

#### **Botanical description**

Pandanus odorifer (Forssk.) Kuntze (Syn.: Pandanus odoratissimus L.f., P. fascicularis Lam.), profusely branched, bushy dioecious shrub or small tree, up to 8 m tall, rarely erect. The stem is supported by numerous thick terete and stilt aerial roots (Figure 4). The leaves are glaucous green, ensiform, 1-1.5 m long, with caudate-acuminate tips and coriaceous texture. Marginal spines point forward, while those on the midrib may point forward or backward. The male flowers are borne on a spadix with numerous subsessile cylindric spikes enclosed in long white fragrant caudate-acuminate spathes. Female flowers feature a solitary spadix. The fruit is oblong or globose, forming a syncarpium that is 15 to 25 cm long (Saxena & Brahmam 1996; Adkar & Bhaskar 2014). The seeds are obovoid, ellipsoid, or oblong, measuring 6-20 mm in length, red-brown on the outside and whitish inside. Propagation is by seeds and vegetative method (Saxena & Brahmam 1996).

#### Phenology

The phenology of *Pandanus odorifer* in Odisha, India, reflects its adaptation to the tropical coastal climate of the region. The plant exhibits distinct seasonal cycles influenced by local environmental factors such as temperature, rainfall patterns, and photoperiod (Saxena & Brahmam 1996). Typically, *P. odorifer* begins its growth cycle with the onset of the monsoon season, benefiting from increased rainfall and humidity conducive to germination and initial growth. As the season progresses, the plant develops its characteristic long, narrow, rigid leaves and prop roots, which are well-suited for survival in diverse substrates ranging from sandy coastal soils to inland clayey or peaty soils. Flowering usually coincides with the peak of the

rainy season or shortly thereafter, depending on local climatic variations. The aromatic inflorescences of *P. odorifer* attract essential pollinators, facilitating fruit set. Fruiting follows flowering, with the development of cone-like fruits containing edible seeds. These phenological patterns, closely tied to seasonal changes and local ecological dynamics, underscore the plant's ecological resilience and cultural importance in the region (Adkar & Bhaskar 2014).



Figure 4 a-f. Morphology of Pandanus odorifer (Forssk.) Kuntze a. whole plant, b. stem, c. leaves, d. roots, e. flower, f. fruit

#### **Reproductive biology**

Pandanus species, including P. odorifer, exhibit diverse reproductive strategies crucial for their ecological success. P. odorifer is dioecious with the female plant producing a non-scented fruit while the male produces a flower rich in volatiles. Fleming and Muchhala (2008) highlighted the genus' adaptation to wind-mediated pollination, complemented by interactions with insects, birds, and bats. Cox (1985, 1990) opined in favor of anemophily in P. odorifer because the pollen lacked pollen kitt and is easily carried off by wind, and because only the male inflorescences are visited by introduced honeybees. The plants utilize floral scents both to attract pollinators and as a defense mechanism against environmental stresses, with scent composition significantly influenced by external factors (Staudt & Bertin 1998; Gershenzon *et al.* 2000). A few nitidulid beetle

species have been recorded as visitors to the male inflorescences of *P. odorifer* (Kato 2000). Recently, Miyamoto *et al.* (2024) opined in favor of insect pollination and stated that both male and female inflorescences are thermogenic at night when insect visitation occurs. Most known *Amystrops* species are associated with the inflorescences, suggesting a widespread, specialized *Pandanus-Amystrops* association.

#### **Chemical composition**

The methanol extract of P. odorifer leaves reportedly contains phytochemical constituents such as steroids, saponins, terpenoids, glycosides, tannins, and flavonoids (Londonkar & Kamble 2009). The leaves also contain pyridine alkaloids, including pandamarilactone-1 (C18H23NO4), pandamarilactone-31 (C19H25NO4), and pandamarilactone-32 (C18H21NO3). The aroma compound 2-acetyl-1-pyrrolidine has been identified in the volatile oil of the leaves (Chilkwad et al. 2008; Kumar et al. 2010, Penu et al. 2020). Active principles isolated from the extract of the whole plant include 3-(4-(dimethylamino) cinnamoyl)-4-hydroxycoumarin, 3,3'-methylenebis(4-hydroxycoumarin), erythro-9,10-dihydroxyoctadecanoic acid. octadecanedioic acid, and dihydroagathic acid (Bharathidasan et al. 2012). Phytochemical analysis of P. odorifer root extracts led to the isolation of phenolic compounds, lignan-type compounds, and some benzofuran derivatives. Identified compounds include  $\alpha$ -terpineol,  $\beta$ -carotene,  $\beta$ -sitosterol, benzyl benzoate, pinoresinol, germacrene-B, vitamin C, viridine, tangeterine, 5,8-hydroxy-7-methoxyflavone, and vanidine (Jong & Chau 1998; Adkar & Bhaskar 2014). Additionally, P. odorifer fruit extract is rich in antioxidant compounds such as caffeoylquinic acids (Zhang et al. 2013), vitamins C, E, and β-carotene (Englberger et al. 2006; Englberger et al. 2009), as well as isopentenyl, dimethylallyl acetates, and cinnamates (Vahirua-Lechat et al. 1996; Tanna et al. 2016). An examination of multiple reports (Nigam & Ahmed 1992; Maheshwari 1995; Naqvi & Mandal 1996; Bisht et al. 1997; Misra & Rao 1997) indicates that 2-phenylethyl methyl ether constitutes the primary component (65.6-75.4%) in natural Kewda oil, followed by terpinene-4-ol (11.7-20.9%), p-cymene (1.0-3.1%), and  $\alpha$ -terpineol (1.2-2.9%) (Nigam & Ahmed 1992; Maheshwari 1995; Naqvi & Mandal 1996; Bisht et al. 1997; Misra & Rao 1997; Mishra et al. 2000).

#### Protecting agricultural landscapes

Landscape refers to the visual aspects of an area of land, often encompassing its physical features and human elements. These landscapes are frequently characterized by subsistence agriculture and a direct reliance on local ecological resources, such as fuelwood or crops for personal consumption (Tallis *et al.* 2008; Egoh *et al.* 2012; Pereponova 2023). Bio-fencing, also known as green fencing or boundary planting, is an ancient agricultural practice adopted by farmers to safeguard their cultivated fields from human and animal encroachment. This method is prevalent in India, where live fences serve to demarcate crop fields, pastures, households, and farm boundaries, creating a network of plant cover across rural landscapes. These live fences not only occur in diverse biophysical settings, including different elevations, ecological life zones, and soil types, but they also reflect distinct cultural and land use histories, particularly in areas with significant agricultural production such as vegetable plantations, pastures, and home gardens (Saucer 1979; Budowski 1987; Panda *et al.* 2018). In regions where land conversion to agriculture has been extensive, live fences often represent the primary form of remaining tree cover in the landscape. One example of a plant commonly used for bio-fencing is *P. odorifer*, whose entire plant forms a green wall (Figure 5). Farmers opt for this plant due to its cost-effectiveness and effectiveness in protecting against herbivores, due to its spiny leaves and dense growth. Furthermore, the leaves of *P. odorifer* possess unpalatable qualities, making them undesirable for herbivores. This combination of physical deterrents and unappealing taste makes it an ideal choice for bio-fencing purposes.

#### **Economic and environmental benefits**

The branches, roots, and leaves of *P. odorifer* hold significant economic value. The branches serve as pillars in small cattle huts. The trunks of aged plants are used in thatched house construction (Little & Skolmen 1989). Besides, the leaves of *Pandanus* are tough and have spines thus employed as fences for crops to protect from cattle. Additionally, Kewda leaves find application in crafting durable nets, mattresses, hats, flower baskets, money purses, handbags, and file folders (Maharana & Lenka 1993; Shiva & Jafri 1998). Dried branches and leaves are commonly used as fuel in rural areas. The trunk and branches are used as fuel wood where other fuel wood is scarce. From an environmental perspective, *P. odorifer* proves to be eco-friendly. Its soil-binding properties are crucial in preventing soil erosion in agricultural fields and stabilizing dunes along the coastline (Panda *et al.* 2007; Pattanaik *et al.* 2008; Aparna & Rajasekhar 2015). This stabilizes dunes, reducing the impact of winds and thereby protecting the coastline. *P. odorifer* emerges as an essential coastal bioresource, significantly contributing to both the socio-economy and ecology of the region.



Figure 5. *Pandanus odorifer* (Forssk.) Kuntze bio-fencing to safeguard cultivated fields from human and animal encroachment

## **Cultural significance**

During Mahashivaratri, the revered celebration dedicated to Lord Shiva, the ritualistic offering of *P. odorifer* flowers, commonly known as Ketaki flowers, holds profound symbolism deeply rooted in Hindu mythology and tradition. This practice's significance can be traced back to ancient texts like the Shiva Purana (Shastri 1970), where a celestial dispute between Lord Shiva and Lord Brahma is recounted. Central to this tale is the Ketaki flower, which, though playing a pivotal role, does so deceitfully, resulting in a curse upon both Brahma and the flower. Despite its checkered past, devotees continue the tradition of offering Ketaki flowers to Lord Shiva during Mahashivaratri, viewing it as an act of repentance and reconciliation (Figure 6a). This ritual serves as a poignant symbol of devotees' acknowledgment of past misdeeds and their earnest quest for forgiveness and divine grace. Through the acceptance of Ketaki flowers during Mahashivaratri, believers express a profound belief in redemption and spiritual growth, underscoring the transformative power of devotion and the eternal compassion of Lord Shiva.

#### Utilizing fibers: Eco-friendly solutions for sustainable industries

In recent times, there has been a significant shift in research focus towards the development of eco-friendly and sustainable materials derived from renewable natural resources. Natural fibers represent one such material category that is environmentally safe, biodegradable, and cost-effective. These fibers are sourced from various parts of plants, including the stem, bast, leaves, and roots (Demircan *et al.* 2020; Ilaiya Perumal & Sarala 2020). Natural fibers are preferred over synthetic

ones for reinforcing polymer-based composite materials. Currently, natural fiber-reinforced polymer composites find applications across a wide range of industries, including automotive, aerospace, construction, military, and industrial sectors (Brailson Mansingh *et al.* 2021). For instance, the prop roots of the *P. odorifer* plant possess high-quality fibers that penetrate deeply into the ground, resulting in improved fiber strength and quantity. These strong fibers are used to manufacture ropes, baskets, and painting brushes (Sahu *et al.* 2013). The root is also used in the thatching of houses and fencing of boundaries (Figure 6b-e). Additionally, local communities often utilize the roots as toothbrushes (Sahu & Mishra 2007). By harnessing the natural fibers from *P. odorifer*, industries can reduce their reliance on synthetic materials and contribute to the promotion of sustainable practices.



Figure 6a-f. a. The ritualistic offering of *Pandanus odorifer* (Forssk.) Kuntze flower during Mahashivaratri, the revered celebration dedicated to Lord Shiva, b. processed roots, c. selling of root in market, d use of roots for thatching of roof, e. use of root in repairing boundary wall, f. burning of *P. odorifer* plant

#### Nutritional significance of fruits

*Pandanus odorifer* fruits, popularly called "Kia Panasa" in Odisha, are consumed in various parts of Odisha (Noor & Satapathy 2020). They are a staple food in Micronesia, Tuvalu, and Kiribati (Miller *et al.* 1956; Englberger 2003). A 100 g portion of the edible pericarp is mainly comprised of water (80 g) and carbohydrates (17 g). There are also significant levels of beta-carotene (19 to 19,000 μg) and vitamin C (5 mg), along with small amounts of protein (1.3 mg), fat (0.7 mg), and fiber (3.5 g) (Englberger 2003; Englberger 2006; Englberger 2006a). The edible flesh of deeper yellow and orange-colored varieties contains higher

levels of provitamin A carotenoids. The fruit of these varieties has considerable potential for alleviating vitamin A deficiency (Englberger 2003).

#### Traditional uses and therapeutic applications

*Pandanus odorifer* is a plant of significant medicinal importance. In traditional Ayurvedic and Siddha medicine, various parts of *P. odorifer*, including flowers, leaves, fruits, and roots, are utilized either individually or in combination with other ingredients to address a diverse array of ailments (Table 1). The leaves of *P. odorifer* are particularly valued for their effectiveness in treating edema, tumors, leprosy, spasms, inflammation, scabies, leukoderma, pain, wounds, ulcers, colic, skin diseases, and rheumatoid arthritis (Warrier 1997; Chatterjee & Pakrashi 2001; Panda *et al.* 2008; Rajeswari *et al.* 2011; Adkar & Bhaskar 2014; Shim 2019). Tablets prepared from leaf extracts are orally administered to alleviate pain, inflammation, and epilepsy in traditional medical practices (Panda *et al.* 2009). The male flowers find application in the treatment of skin and heart-related infections (Adkar & Bhaskar 2014). Additionally, the roots are utilized for their antidiabetic, antidote, abortifacient, hemorrhoidal, and dermatological (for skin diseases, leprosy, scabies, and syphilis) properties (Jain 1991; Meilleur *et al.* 1997). The fruit exhibits efficacy in treating ailments such as vat, kapha, urinary discharge, and leprosy, and also serves as a male aphrodisiac, possessing notable analgesic, anti-pyretic, cytotoxic, and anti-inflammatory activities (Adkar & Bhaskar 2014; Hossain *et al.* 2020). Furthermore, the oil derived from *P. odorifer* is recommended for its efficacy in managing rheumatoid arthritis, skin diseases, earaches, headaches, arthritis, debility, depurative purposes, giddiness, as a laxative, for leprosy, and for alleviating spasms (Adkar & Bhaskar 2014).

Promoting sustainable cultivation and ethical harvesting practices for *Pandanus odorifer* can create livelihood opportunities for local communities, ensuring a steady supply of plant material for traditional medicine while preserving natural habitats. This approach supports both economic stability and environmental conservation. Establishing community-owned enterprises for the production and processing of *P. odorifer* products can economically empower indigenous populations by enhancing local knowledge on cultivation, processing, and marketing. Developing value-added products like oils, extracts, and herbal formulations derived from *P. odorifer* can boost market potential and create niche markets that respect traditional practices. Protecting and documenting the traditional knowledge associated with *P. odorifer* preserves cultural heritage and ensures communities benefit from their intellectual property. Sharing benefits from the commercial use of traditional medicine can further improve socio-economic outcomes. Integrating *P. odorifer* into mainstream healthcare systems can provide affordable healthcare, particularly in rural areas, supporting the continued practice of traditional medicine and preserving biodiversity (Warrier 1997; Bodeker & Kronenberg 2002; Panda 2004).

Parts	Disorder treated	Traditional	Country	Reference
used		medicine		
Flower	Syphilis, tumor, skin diseases, leukoderma,	Siddha	Sri Lanka,	Warrier <i>et al.</i> 2005, Raina <i>et</i>
	leprosy, generating perspiration, earache,	Medicine,	India	al. 2004, Ilanchezhian and
	bronchitis, blood diseases, asthma, urinary	Ayurveda		Joseph 2006, Khare 2011,
	tract illnesses, headache, rheumatism,			Sathasivampillai et al. 2015,
	constipation, diabetes.			2016, 2017, 2018.
Leaf	Brain diseases, heart diseases, leukoderma,	Ayurveda	Sri Lanka,	Warrier <i>et al.</i> 2005, Raina <i>et</i>
	scabies, skin diseases, smallpox, tumor,		India	<i>al.</i> 2004
	leprosy, syphilis, headache, rheumatism.			
Root	Constipation, urinary tract illnesses,	Ayurveda	India,	Nadkarni 1996, Jong and
	diabetes, syphilis, fever, thyroid disorders.		Taiwan	Chau 1998, llanchezhian
				and Joseph 2006, Khare
				2011.
Fruit	Vat, kapha, urinary discharge, leprosy, male	Ayurveda	India,	Adkar and Bhaskar 2014,
	aphrodisiac, analgesic, anti-pyretic,		Bangladesh	Hossain <i>et al.</i> 2020
	cytotoxic, anti-inflammatory activities			

Table 1. Traditional medicinal uses of P. odorifer

#### Modernizing traditional methods of application in the field of medicine

Natural products and traditional medicines, such as Traditional Chinese Medicine, Ayurveda, Kampo, Traditional Korean Medicine, and Unani, have evolved into well-regulated systems, deeply rooted in cultural heritage. These practices are increasingly integrated with modern science, particularly through the fields of ethnobotany and ethnopharmacognosy, which guide researchers in discovering new molecules from diverse sources. The flora of the tropics, with its rich biodiversity, plays

a crucial role in providing new leads for drug development. Natural products, which have evolved over millions of years, offer unique chemical diversity and drug-like properties, making them invaluable resources for creating new therapeutic agents (Fabricant & Farnsworth 2001; Suntar 2020; Pirintsos *et al.* 2022; Chaachouay & Zidane 2024). Modernizing traditional methods of application in the field of medicine involves a synergistic approach that blends centuries-old herbal wisdom with contemporary scientific advancements. The phytochemical constituents of plants like *Pandanus odorifer*, renowned for their aromatic and therapeutic properties, can be validated and enhanced through scientific research and advanced extraction techniques. Innovative technologies such as Supercritical Fluid Extraction, Microwave-Assisted Extraction, High-Performance Liquid Chromatography, Gas Chromatography, Nuclear Magnetic Resonance Spectroscopy, and Crystallography, have enabled the precise determination of active compounds, leading to the development of novel therapeutic drugs. This approach not only preserves and respects indigenous knowledge but also bridges the gap between traditional and modern medicine, offering more holistic and personalized treatment options. By integrating these traditional methods into mainstream healthcare, a wide spectrum of health concerns can be addressed, promoting overall well-being. The convergence of ancient practices with modern methodologies fosters a comprehensive approach to healthcare, ensuring that the wisdom of the past is leveraged alongside the advancements of the present to enhance therapeutic efficacy and patient outcomes. (Galm & Shen 2007; Yuan *et al.* 2016; Ponphaiboon *et al.* 2023).

#### Traditional techniques in kewda perfumery: preserving centuries-old craftsmanship

India boasts a rich perfumery tradition dating back over 5000 years to the ancient civilization of the Indus Valley. Excavations at sites such as Harappa and Mohenjo-Daro have unearthed artifacts resembling present-day Deg and Bhabka traditional distillation units, indicating the early use of water distillation techniques. During ancient times, Indian perfumes and aromatic substances such as Sandalwood, Kashmiri saffron, Assami Black agar, Himalayan Kasturi, various floral Attars, and essential herbal oils were highly sought-after commodities in trade. The perfume industry gained significant momentum during the reign of King Harshabardhana (606 to 647 AD), as evidenced by historical records. Indian fragrances found their way to Europe through Egypt, often referred to as Arabian perfumes, showcasing India's influence on global trade during that era. Cities like Kanauj, Kumbakonam, Pataliputra, Pandharpur, Poona, and Bangalore emerged as prominent centers for essential oil industries in ancient India. King Harshabardhana even imposed a business tax on perfume products, indicating the flourishing trade of aromatic substances during his reign. The Mughal emperors further enriched India's perfumery tradition with their love for exotic fragrances, as documented in Ain-e-Akbari (1547-1605) by Abul Fazl. During the Mughal period, cities like Lucknow, Aligarh, Jaunpur, Ghazipur, Delhi, and Jaipur became renowned hubs for perfume and floral extracts (Roy 1991; Saran 2005; Eraly 2007; Dwivedi 2011).

Pandanus odorifer, is an economically significant aromatic plant valued for its essential oil. This oil, extracted primarily from male flowers, is in high demand across various industries including aromatherapy, cosmetics, and food flavoring due to its distinctive fragrance. The key aromatic compound in this oil is phenylethyl methyl ether (Nasim et al. 2021). The Kewda perfume industry, a significant cottage industry in the coastal Ganjam district of Odisha, traces its origins back approximately 200 years. It commenced in the outskirts of Berhampur, Ganjam District, by Muslim immigrants from the Punjab province of undivided India (Dutta et al. 1987). Kewda Attar, a renowned fragrance in India since ancient times, primarily originates from this region, with approximately 90% of Kewda essence sourced from Ganjam District (Rout et al. 2011). Kewda attar, water, and oil (rooh) are integral perfumery products in India. The process of preparing these products involves collecting Kewra flowers in the early morning using plucking sticks, followed by distillation in distilleries to extract the perfumery components. Kewra Attar is produced by removing the outer green spiny parts and pointed tips of the spathe of the flowers. Subsequently, 1000 flowers are added to 60 liters of water and heated. The steam, along with Kewda oil vapors, is passed through sandalwood oil kept in a receiver, where the Kewda aroma is gradually absorbed, resulting in Kewda Attar. It typically contains 3-5% Kewda oil blended with sandalwood oil and is utilized in various applications such as scenting clothes, bouquets, lotions, cosmetics, soaps, hair oils, tobacco products, Pan Masala, and Agarbati (incense sticks). Kewda attar and water are used for flavouring various foods, sweets, syrups, and soft drinks. They are popular in north India, especially on festive occasions. Kewra water is obtained through a similar distillation process with water, yielding approximately 10-18 liters from 1000 flowers. It contains about 0.02% Kewda oil and is utilized for flavoring sweets, syrups, soft drinks, and other food preparations, as well as in natural packaged food items. The essential oil from Kewda flowers, known as Kewda rooh, is obtained through hydro-distillation, where flowers are repeatedly distilled without a base oil in the receiver vessel. Approximately 30 grams of pure oil can be extracted from 1000 flowers (Kusuma et al. 2012). Kewda rooh is valued for its stimulant and antispasmodic properties, often administered for headaches and rheumatism. Additionally, there is considerable potential for export to Arabian countries and the Middle East (Sahu & Misra 2007).

#### The ecological impact of burning Pandanus odorifer on biodiversity

Pandanus odorifer is recognized as a vital species extensively utilized by communities for their economic needs (Pattanaik *et al.* 2008). Groves of *P. odorifer* are considered significant vegetative formations, particularly in association with human settlements. However, the burning of *P. odorifer*, a crucial component of various ecosystems, poses substantial threats to biodiversity (Figure 4f). As a keystone species, Pandanus provides habitat and sustenance for a diverse array of organisms, including birds, insects, snakes, and small mammals (Bachan & Nasser 2015; Gurmeet & Amrita 2015; Solomon Raju 2020). The destruction of *Pandanus* disrupts these intricate ecosystems, resulting in a loss of biodiversity. Moreover, *Pandanus* plays a critical role in stabilizing coastal dunes, offering protection against erosion and storm surges (Tongway & Ludwig 2011). Its absence renders different habitats susceptible to environmental degradation, diminishing resilience to climate change impacts. Preserving *P. odorifer* is imperative for the species it directly supports and for the overall health and resilience of different ecosystems.

#### Contribution of spatiotemporal evolution and traditional knowledge in achieving sustainability goals

The spatiotemporal evolution and traditional knowledge of *Pandanus odorifer* significantly contribute to achieving sustainability goals by integrating ecological, economic, and cultural dimensions. Its wide geographical distribution and ecological adaptability inform strategies to mitigate the impacts of climate change and habitat loss, aiding biodiversity conservation and climate change adaptation (Stone 1970; Bellwood & Hughes 2001). Traditional knowledge provides deep insights into sustainable practices such as bio-fencing, soil conservation, and medicinal uses, honed over generations. These practices support local economies, cultural heritage, and sustainable resource management, fostering a harmonious relationship with nature. Integrating scientific research with traditional wisdom enhances ecosystem management and strengthens community resilience, promoting sustainable use of natural resources. This holistic approach aligns with global sustainability targets, including the UN Sustainable Development Goals (SDGs) related to life on land (SDG 15), climate action (SDG 13), sustainable communities (SDG 11), food security through sustainable agriculture (SDG 2), health benefits through ethnobotanical practices (SDG 3), and responsible consumption and production (SDG 12) (Kanie *et al.* 2019; Vijge *et al.* 2020). Thus, the spatiotemporal evolution and traditional knowledge of *P. odorifer* play a vital role in advancing sustainability efforts, enhancing ecosystem resilience, and fostering sustainable development in communities dependent on this valuable species.

#### Limitations of the study

This study on *Pandanus odorifer* encounters some limitations. One significant limitation is the phenological observations, which were restricted to a limited geographical area. This restriction potentially overlooks variations in different environmental conditions and regions. Additionally, the sustainability assessments might not fully capture the complex interactions between the species and its ecosystem, especially in the context of climate change and human activities.

## Conclusion

The goal of this paper is to explore the drivers of ecosystem services provided by *Pandanus odorifer*. The present communication delves into the diverse importance of *P. odorifer* across various domains, spanning botany, ecology, traditional medicine, perfumery, and economic sustainability. The plant's thriving presence in diverse ecosystems exhibits remarkable adaptability and ecological significance. *P. odorifer* serves multiple crucial roles, from acting as a protective barrier in agricultural bio-fencing to serving as a valuable source of essential oils for the perfumery industry. Its versatility is deeply rooted in human culture and livelihoods, reflecting its profound integration into various aspects of daily life. Moreover, the plant's nutritional value and therapeutic applications underscore its holistic importance to communities across different regions. Its multifaceted contributions underscore its pivotal role in sustaining and enriching human life. However, in some areas, it is found that people are burning the plants, which creates a negative impact on the environment and ecosystem. In this context, raising awareness among people is necessary for the proper maintenance of biodiversity. In conclusion, this study underscores the fundamental role played by *P. odorifer* in numerous aspects of human existence, ranging from its ecological contributions to its cultural and economic significance. By amalgamating traditional knowledge with scientific understanding, we emphasize the imperative of preserving indigenous practices and biodiversity for sustainable development. Moving forward, continued research and conservation efforts are essential to safeguard this invaluable plant species and unlock its full potential for the well-being of present and future generations.

## Declarations

List of abbreviations: The article does not contain abbreviations. Ethics approval and consent to participate: Not applicable.

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## Literature cited

Adkar PP, Bhaskar VH. 2014. *Pandanus odorotissimus* (Kewda): a review on ethnopharmacology, phytochemistry, and nutritional aspects. Advances in Pharmacological Sciences 2014:120895.

Al-Khayri JM, Rashmi R, Toppo V, Chole PB, Banadka A, Sudheer WN, Nagella P, Shehata WF, Al-Mssallem MQ, Alessa FM, Almaghasla MI, Rezk AA. 2023. Plant secondary metabolites: the weapons for biotic stress management. Metabolites 13(6):716.

Anjali, Kumar S, Korra T, Thakur R, Arutselvan, Kashyap AS, Nehela Y, Chaplygin V, Minkina T, Keswani C. 2023. Role of plant secondary metabolites in defence and transcriptional regulation in response to biotic stress. Plant Stress 8:100154.

Aparna S. Raja Sekhar PS. 2015. Studies on the coastal sand dune phytoresources at Visakhapatnam, Bay of Bengal, India. Asian Journal of Plant Science and Research 5:69-76.

Audley-Charles MG. 1987. Dispersal of Gondwanaland: relevance to the evolution of the angiosperms. In: Whitmore TC. (eds.). Biogeographical evolution of the Malay Archipelago. Cambridge University Press, Cambridge, U.K., Pp 5-25.

Bachan A, Nasser KMM. 2015. Conservation of screw pine *Pandanus* in Kerala, need for reestablishment of community - resource link. Meridian 4 (2):106-111.

Baddu, V, Ouano N. 2018. Ethnobotanical survey of medicinal plants used by the Y' Apayaos of Sta. Praxedes in the province of Cagayan, Philippines. Mindanao Journal of Science and Technology 16:128-153.

Bellwood DR, Hughes TP. 2001. Regional-scale assembly rules and biodiversity of coral reefs. Science 292(5521):1532-1535.

Bharathidasan R, Mahalingam R, Ambikapathy V, Panneerselvam A. 2012. Phytochemical compound analysis of *Pandanus odoratissimum*. Asian Journal of Plant Science and Research 2:228-231.

Bisht M, Sharma S, Maheshwari ML. 1997. Composition of essential oil from the flowers of Kewda (*Pandanus odoratissimus* L.). Indian Perfume 41(2):69-72.

Bodeker G, Kronenberg F. 2002. A public health agenda for traditional, complementary, and alternative medicine. American Journal of Public Health 92(10):1582-1591.

Brailson Mansingh B, Binoj JS, Prem Sai N, Hassan SA, Siengchin S, Sanjay MR, Cheng LY. 2021. Sustainable development in utilization of *Tamarindus indica* L. and its by-products in industries: A review. Current Research in Green and Sustainable Chemistry 4(1):e100207.

Budowski G. 1987. Living fences in tropical America: A widespread agroforestry practice. In: Gholz HL. (eds). Agroforestry: Realities, possibilities and potentials. Kluwer Academic Publishers, Dordrecht, Netherlands, Pp. 169-178.

Buerki S., Callmander MW, Devey DS, Chapell L, Gallaher T, Munzinger J, T. Haevermans T, Forest F. 2012. Straightening out the screw-pines: a first step in understanding phylogenetic relationships within Pandanaceae. Taxon 61:1010-1020.

Callmander MW, Chassot P, Küpfer Ph, Lowry II PP. 2003. Recognition of Martellidendron, a new genus of Pandanaceae, and its biogeographic implications. Taxon 52:747-762.

Callmander MW, Gallaher, T.J., McNeill, J., Beentje, H., Nadaf, A.B., Middleton, D.J., Buerki, S., 2021. Neotypification of *Pandanus odorifer*, the correct name for *P. odoratissimus* (Pandanaceae). Taxon 70:182-184.

Callmander MW, Lowry II PP, Forest F, Devey DS, Beentje H, Buerki S. 2012. *Benstonea* Callm. & Buerki (Pandanaceae): characterization, circumscription, and distribution of a new genus of screw-pines, with a synopsis of accepted species. Candollea 67:323-345.

Callmander MW. 2000. *Pandanus* subg. Martellidendron (Pandanaceae) part 1: New findings on *Pandanus hornei* Balf. f. (sect. Seychellea) from the Seychelles. Webbia 55:317-329.

Callmander MW. 2001. *Pandanus* subg. *Martellidendron* (Pandanaceae) part II: revision of sect. *Martellidendron* Pic. Serm. in Madagascar. Botanical Journal of Linnean Society 137:353-374.

Cebrian-Piqueras MA, Filyushkina A, Johnson DN, Lo VB, López-Rodríguez MD, March H, Oteros-Rozas E, Peppler-Lisbach C, Quintas-Soriano C, Raymond CM, Ruiz-Mallenet I, van Riper CJ, Zinngrebe I, Plieninger P. 2020. Scientific and local ecological knowledge, shaping perceptions towards protected areas and related ecosystem services. Landscape Ecology 35:2549-2567.

Chaachouay N, Zidane L. 2024. Plant-derived natural products: A source for drug discovery and development. Drugs and Drug Candidates 3(1):184-207.

Champion HG, Seth SK. 1968. A revised survey of the forest types of India. Manager of Publications, New Delhi, India, Pp.16-17.

Chatterjee A, Pakrashi SC. 2001. The Treatise on Indian medicinal plants. 6th ed. National Institute of science communication, New Delhi, India, Pp. 9-10.

Chilkwad SR, Manjunath KP, Akki KS, Savadi RV, N. Deshpande N. 2008. Pharmacognostic and Phytochemical investigation of leaves of *Pandanus odoratissimus* Linn.f. Ancient Science of Life 28 (2):3-6.

Cox PA. 1985. Islands and dioecism: insights from the reproductive ecology of *Pandanus tectorius* in Polynesia. In: White J (eds.). Studies on plant demography: A Festschrift for John L. Harper. Academic Press, London, Pp. 355-368.

Cox PA. 1990. Pollination and evolution of breeding systems in Pandanaceae. Annals Missouri Botanical Garden 77:816-840.

Demircan G, Kisa M, Ozen M, Aktas B. 2020. Surface-modified alumina nanoparticles-filled aramid fiber-reinforced epoxy nanocomposites: Preparation and mechanical properties. Iranian Polymer Journal 29 (1):253-64.

Divekar PA, Narayana S, Divekar BA, Kumar R, Gadratagi BG, Ray A, Singh AK, Rani V, Singh V, Singh AK, Kumar A, Singh RP, Meena RS, Behera TK. 2022. Plant secondary metabolites as defense tools against herbivores for sustainable crop protection. International Journal of Molecular Sciences 23(5):2690.

Dutta, P.K., Saxena, H.O. & Brahmam, M. 1987. Kewda perfume industry in India. Economic Botany 41:403-410.

Dwivedi K. 2011.Perfumery in ancient India. Indian Historical Review 38 (2):69-84.

Egoh BN, O'Farrell PJ, Charef A, Gurney LJ, Koellner T, Nibam Abi H, Egoh M, Willemen L. 2012. An African account of ecosystem service provision: use, threats and policy options for sustainable livelihoods. Ecosystem Services 2:71-81.

Englberger L, Aalbersberg W, Dolodolotawake U, Schierle J, Humphries J, luta T, Marks GC, Fitzgerald MH, Rimon B, Kaiririete M. 2006. Carotenoid content of *Pandanus* fruit cultivars and other foods of the Republic of Kiribati. Public Health Nutrition 9(5):631-643.

Englberger L, Aalbersberg W, Schierle J, Marks GC, Fitzgerald MH, Muller F, Anko Jekkein A, Alfred J, Vander Veld N. 2006a. Carotenoid content of different edible *Pandanus* fruit cultivars of the Republic of the Marshall Islands. Journal of Food Composition and Analysis 19 (6-7):484-494.

Englberger L, Schierle J, Hofmann P, Lorens A, Albert K, Levendusky A, Paul Y, Lickaneth E, Elymore A, Maddison M, deBrum I, Nemra J, Alfred J, Vander Velde N, Kraemer K. 2009. Carotenoid and vitamin content of Micronesian atoll foods: *Pandanus (Pandanus tectorius)* and garlic pear (*Crataeva speciosa*) fruit. Journal of Food Composition and Analysis 22:1-8.

Englberger L. 2003. Are Pacific islanders still enjoying the taste of pandanus? in Pacific Islands Nutrition 58:10-11.

Eraly A. 2007. The Mughal World: Life in India's Last Golden Age. Penguin Books, India.

Fabricant DS, Farnsworth NR. 2001. The value of plants used in traditional medicine for drug discovery. Environmental Health Perspectives 109 (Suppl 1):69-75.

Fedele G, Urech Z, Rehnus M, Sorg J. 2011. Impact of women's harvest practices on *Pandanus guillaumetii* in Madagascar's lowland rainforests. Economic Botany 65(2):158-168.

Fleming TH, Muchhala N. 2008. Nectar-feeding bird and bat niches in two worlds: pantropical comparisons of vertebrate pollination systems. Journal of Biogeography 35(5):764-780.

Fogliarini C, Ferreira CEL, Bornholdt J, Barbosa M, Giglio V, Bender M. 2021. Telling the same story: fishers and landing data reveal changes in fisheries on the southeastern Brazilian coast. PLoS One 16:e0252391.

Frank JH, Lounibos LP. 1983. Phytotelmata: Terrestrial plants as hosts for aquatic insect communities. Plexus Publishing, London, UK.

Fusi M, Bayi J, Ngo-Massou V *et al.* 2021. Ecology of the facultative phytotelmic crab *Platychirarma buettikoferi* (De Man, 1883) (Decapoda, Brachyura, Sesarmidae). Crustaceana 94:1043-1058.

Gallaher T, Callmander MW, Buerki S, Keeley SC. 2015. A long-distance dispersal hypothesis for the Pandanaceae and the origins of the *Pandanus tectorius* complex. Molecular Phylogenetics and Evolution 83:20-32.

Galm U, Shen B. 2007. Natural product drug discovery: The times have never been better. Chemistry and Biology 14:1098-1104.

Gershenzon J, Mcconkey ME, Croteau RB. 2000. Regulation of monoterpene accumulation in leaves of peppermint. Plant Physiology 122:205-213.

Grigoriadou K, Cheilari A, Dina E, Alexandri S, Aligiannis N. 2023. Medicinal and aromatic plants as a source of potential feed and food additives. In: Arsenos G, Giannenas I. (eds). Sustainable use of feed additives in livestock. Springer, Cham.

Grushvitsky IV. 1982. Order Pandans (Pandanales). In: Takhtajan AL (eds). Plant Life. Prosvescheniye, Moscow, Pp. 451-461 (in Russian).

Guillaumet J. 1973. Formes et developpement des Pandanus malgaches. Webbia 28:495-519.

Gurmeet S. Amrita, P. 2015. Unique *Pandanus* - Flavour, food and medicine. Journal of Pharmocognosy and Phytochemistry 5:8-14.

Hossain MS, Akhter S, Nath BD, Khanom MM, Kobir ME, Khanom A, Islam MT, Yasmin MN, Islam MR, Bayen S, Ferdousy S. 2020. Chemical group characterization and determination of pharmacological activities of *Pandanus fascicularis* L. fruit. Discovery Phytomedicine 7(3):128-137.

Hutchinson J.1973. The families of flowering plants. Oxford University Press, London, U.K.

Huynh KL. 2001. Contribution to the flower structure of *Sara ranga* (Pandanaceae). Botanical Journal of Linnean Society 136:239-245.

Ilaiya Perumal C, Sarala R. 2020. Characterization of a new natural cellulosic fiber extracted from Derris scandens stem. International Journal of Biological Macromolecules 165 (1):2303-2013.

Ilanchezhian R, Joseph R. 2006. Hepatoprotective and hepatocurative activity of the traditional medicine ketaki (*Pandanus odoratissimus* Roxb.). Asian Journal of Traditional Medicines 5:212-218.

Jain SK. 1991. Dictionary of Indian folk medicine and ethnobotany. Deep Publications, New Delhi, India.

Jong TT, Chau SW. 1998. Antioxidative activities of constituents isolated from *Pandanus odoratissimus*. Phytochemistry 49:2145-2148.

Kanie N, Griggs D, Young O, Waddell S, Shrivastava P, Haas PM, Broadgate W, Gaffney O, Korosi C. 2019. Rules to goals: Emergence of new governance strategies for sustainable development. Sustainability Science 14:1745-1749.

Kato M. 2000. Anthophilous insect community and plant-pollinator interactions on Amami Islands in the Ryukyu Archipelago, Japan. Contributions from the Biological Laboratory Kyoto University 29:157-254.

Khare CP. 2011. Indian herbal remedies: Rational western therapy, Ayurvedic and other traditional usage. Botany. Springer-Verlag Berlin, Heidelberg.

Kirtikar KR, Basu BD, Blatter E. 1991. Indian medicinal plants. Vol. 4, Indian Book Center, New Delhi, India, Pp. 115.

Kumar D, Kumar S, Kumar S, Singh J, Sharma C, Aneja KR. 2010. Antimicrobial and preliminary phytochemical screening of crude leaf extract of *Pandanus Odoratissimus* L. Pharmacologyonline 2:600-610.

Kusuma R, Reddy VP, Bhaskar BN, Venkatesh S. 2012. Phytochemical and pharmacological studies of *Pandanus odoratissimus* Linn. International Journal of Pharmacognosy and Phytochemistry 2(4):171-174.

Langenberger G, Prigge V, Martin K, Belonias B, Sauerborn J. 2009. Ethnobotanical knowledge of Philippine lowland farmers and its application in agroforestry. Agroforestry Systems 76:173-194.

Lehtinen RM. 2002. The use of screw pines (*Pandanus* spp.) by amphibians and reptiles in Madagascar. Herpetological Bulletin 82:20-5.

Limbongan J, Malik A. 2009. Peluang Pengembangan Buah Merah (*Pandanus conoideus* Lamk.) Di Provinsi Papua. Jurnal Litbang Pertanian 28:134-41.

Little EL, Skolmen RG. 1989. Common forest trees of Hawaii (Native and Introduced). Agricultural Handbook 679, USDA, Washington, DC, USA.

Loa IW, Cheng YB, Haung CC, Hwang TL, Wu CC, Liou JR, Hou MF, Yuan SS, Chang FR, Wu YC. 2016. Constituents of the leaves of *Pandanus utilis*. Natural Product Communication 11:173-176.

Londonkar R, Kamble A. 2009. Evaluation of free radical scavenging activity of *Pandanus odoratissimus*. International Journal of Pharmacology 5:377-80.

Maharana T, Lenka PC. 1993 Productivity and industrial use study in medicinal and aromatic plants of Orissa with special reference to *Pandanus* and *Cymbopogon*. Indian Council of Agricultural Research, New Delhi, India.

Maheshwari M. L. 1995. Composition of essential oil from flowers of keora (*Pandanus odoratissimus* Linn.) by capillary gas chromatography. Indian Perfume 39 (1):45 - 48.

Mary Sheeja TL, Jacob J, Balasundaram J, Gomathi Venkatachalam G. 2021. Preliminary phytochemical screening and GC-MS analysis of methanolic extract of roots of *Pandanus fascicularis*. Asian Journal of Biological and Life Sciences 10 (3):626-631.

Meeuse A.1966. Fundamentals of phytomorphology. Ronald Press, New York, U.S.A.

Meeuse ADJ.1965. Angiosperms-past and present: phylogenetic botany and interpretative floral morphology of the flowering plants. In: Vira R, Chandra L. (eds). Advancing frontiers of plant sciences. Impex, New Delhi, India, Pp. 170-182.

Meilleur BA, Maigret MB, Manshardt R. 1997. Hala and Wauke in Hawaii. Bishop Museum. Bulletin in Anthropology 7:1-55.

Miller CD, Murai M, Pen F. 1956. The use of Pandanus fruit as food in Micronesia, Pacific Science 10:3-16.

Mishra R, Dash PK, Rao YR. 2000. Chemical composition of the essential oils of Kewda and Ketaki. Journal of Essential Oil Research 12(2):175-178.

Misra R, Rao YR. 1997. A GC/MS study of Kewda (Pandanus Odoratissimus Linn.) oil. Indian Perfume 41 (4):143 - 145.

Miyamoto T, Mochizuki K, Kawakita A. 2024. Pollination of thermogenic inflorescence of *Pandanus odorifer* by a specialist *Amystrops* sap beetle that reproduces on the male inflorescence. Botanical Journal of the Linnean Society XX:1-12.

Nadaf A, Zanan R. 2012a. Biogeography of Indian Pandanaceae. In: Indian Pandanaceae - an overview. Springer, New Delhi.

Nadaf A, Zanan RL. 2012b. Indian Pandanaceae—An Overview. Springer, Germany.

Nadkarni KM. 1996. Indian materia medica: with Ayurvedic, Unani-Tibbi, Siddha, Allopathic, Homeopathic, Naturopathic & home remedies, appendices & indexes. Popular Prakashan Private Ltd, Bombay, India.

Naqvi AA, Mandal S. 1996. Investigation of the essential oil of *P.fascicularis* Lam. by GC/MS.J. Essential Oil Research 8:571 - 572.

Nasim N, Behera JK, Sandeep IR, RamaRao VV, Kar B, Mishra A, Nayak S, Mohanty S. 2018. Phytochemical analysis of flower from *Pandanus odorifer* (Forssk.) Kuntze for industrial application. Natural Product Research 32 (20):2494-2497.

Nasim N, Sandeep IS, Nayak S, Mohanty S. 2021. Cultivation and utilization of *Pandanus odorifer* for industrial application. In: Ekiert HM, Ramawat KG, Arora J. (eds). Medicinal plants. Sustainable Development and biodiversity. Springer, Cham.

Nigam MC, Ahmed A. 1992. Chemical and gas chromatographic examination of essential oil of *Pandanus odoratissimus* (keora). Indian Perfume 36 (2):93 - 95.

Noor N, Satapathy KB. 2020. Ethnobotanical exploration of wild edible fruits of Balasore district in Odisha, India - An initiative for accelerating food security and biodiversity conservation. International Journal of Botany Studies 5 (3):448-457.

Ordas JAD, Nonato MG, CECILIA B. Moran CB. 2021. Ethnobotanical uses of Pandanaceae species in selected rural communities in the Philippines. Economic Botany XX(X):1-18.

Padhy S, Dash SK, Panda BB, Misra MK, Padhi NP, Sahu D, Das M, Nayak SP, Dash B, Rama Rao VV, Mohanty PC, Padhy R. 2016. *Pandanus fasicularis* Lamk. (Kewda): The Prime vegetation in the hinterland biodiversity of coastal Odisha, with unique ethnic utility, genetic variation and economics- A review. Journal of Biodiversity 7(1):33-49.

Panda H. 2004. Handbook on medicinal herbs with uses. Asia Pacific Business Press Inc. New Delhi, India.

Panda K, Das AB, Panda BB. 2012. Genomics of *Pandanus*: A useful plant resource of coastal Odisha, India. Lambert Academic Publishing, Germany.

Panda KK, Das AB, Panda BB. 2009. Use and variation of *Pandanus tectorius* Parkinson (*P. fascicularis* Lam.) along the coastline of Orissa, India. Genetic Resources and Crop Evolution 56:629-637.

Panda KK, Panigrahy RK, Das AB, Panda BB. 2007. Analyses of chromosome number, nuclear DNA content and RAPD profile in three morphotypes of *Pandanus fascicularis* Lam. Plant Genetic Resources Newsletter 152:12-22.

Panda P, Panda DP, Panda PK, Nayak SS. 2008. Antinociceptive and anti- inflammatory activities of *Pandanus fascicularis* Lamk. leaves in animal models. Oriental Pharmacy and Experimental Medicine 7:485-493.

Panda T, Mishra N, Pradhan BK, Mohanty RB. 2018. Live fencing: an ecofriendly boundary wall in Bhadrak district of Odisha, India. International Journal of Conservation Science 9 (2):301-310.

Pattanaik C, Reddy CS, Dhal NK. 2008. Phytomedicinal study of coastal and sand dune species of Orissa. Indian Journal of Traditional Knowledge 7:263-268.

Penu FI, Ivy SM, Ahmed F, Uddin J, Hossain MS, Labu ZK. 2020. In vitro assessment of antioxidant, thrombolytic, antimicrobial activities of medicinal plant *Pandanus odoratissimus* L. leaves extract. Journal of Scientific Research 12 (3):379-390.

Pereponova A, Lischeid G, Grahmann K, Bellingrath-Kimura SD, Ewert FA. 2023. Use of the term "landscape" in sustainable agriculture research: A literature review. Heliyon 9(11):e22173.

Pirintsos S, Panagiotopoulos A, Bariotakis M, Daskalakis V, Lionis C, Sourvinos G, Karakasiliotis I, Kampa M, Castanas E. 2022. From traditional ethnopharmacology to modern natural drug discovery: a methodology discussion and specific examples. Molecules 27(13):4060.

Ponphaiboon J, Krongrawa W, Aung WW, Chinatangkul N, Limmatvapirat S, Limmatvapirat C. 2023. Advances in natural product extraction techniques, electrospun fiber fabrication, and the integration of experimental design: A comprehensive review. Molecules 28:5163.

Ragragio E, Zayas C, Obico J. 2013. Useful plants of selected Ayta communities from Porac, Pampanga, twenty years after the eruption of Mt. Pinatubo Philippine. Journal of Science 142:169-181.

Raina VK, Kumar A, Srivastava SK, Syamsundar KV, Kahol AP. 2004. Essential oil composition of 'kewda' (*Pandanus odoratissimus*) from India. Flavour and Fragrance Journal 19:434-436.

Rajeswari J, Kesavan K, Jayakar B. 2011. Phytochemical and pharmacological evaluation of prop roots of *Pandanus fascicularis* Lam. Asian Pacific Journal of Tropical Medicine 4:649-653.

Ramawat KG, Goyal S. 2020. Co-evolution of secondary metabolites during biological competition for survival and advantage: An overview. In: Mérillon JM, Ramawat K. (eds). Co-Evolution of secondary metabolites. Reference Series in Phytochemistry. Springer Cham., Pp 3-17.

Rashmi D, Barvkar VT, Nadaf A, Mundhe S, Kadoo NY. 2019. Integrative omics analysis in *Pandanus odorifer* (Forssk.) Kuntze reveals the role of Asparagine synthetase in salinity tolerance. Scientific Report 9:932.

Raymond CM, Fazey I, Reed MS, Stringer LC, Robinson GM, Evely AC. 2010. Integrating local and scientific knowledge for environmental management. Journal of Environment Management 91:1766-1777

Reyes-García V, Vadez V, Huanca T, Leonard WR, Mcdade T. 2007. Economic development and local ecological knowledge:A deadlock? quantitative research from a native Amazonian Society. Human Ecology 35:371-377.

Rout PK, Naik S and Rao YR. 2011. Liquid CO<sub>2</sub> extraction of flowers of *Pandanus fascicularis* Lam. and fractionation of floral concrete and comparative composition of the extracts. Journal of Food Biotechnology 35 (2):500-512.

Roy T. 1991. The Economic History of India: From Pre-Colonial Times to 1991. Oxford University Press, U.K.

Sahu D, Misra M. 2007. Ecology and traditional technology of screw pine perfume industry in coastal Orissa. Indian Journal of Traditional Knowledge 6(2):253-261.

Sahu SC, Pattnaik SK, Dash SS, Dhal NK. 2013. Fibre-yielding plant resources of Odisha and traditional fibre preparation knowledge – An overview. Indian Journal of Traditional Knowledge 4(4):339-347.

Saran R. 2005. Indian perfumery: Tradition and trade. Harper Collins, India.

Sathasivampillai SV, Rajamanoharan PRS, Munday M, Heinrich M. 2017. Plants currently used to treat diabetes in Sri Lankan Siddha Medicine - an ethnobotanical survey in the Eastern Province. Presented at the World Congress Integrative Medicine & Health 2017: part three. Springer Nature, Berlin, Germany, Pp. 333.

Sathasivampillai SV, Rajamanoharan PRS, Munday M, Heinrich M. 2018. Siddha medicine in eastern Sri Lanka todaycontinuity and change in the treatment of diabetes. Frontiers in Pharmacology 9:1022.

Sathasivampillai SV, Rajamanoharan PRS, Munday M, Heinrich, M. 2015. Preparations and plants used to treat diabetes in Sri Lankan Siddha Medicine. Presented at the 3rd International Conference on Ayurveda, Unani, Siddha and Traditional Medicine, Institute of Indigenous Medicine, University of Colombo, Colombo, Pp. 67.

Sathasivampillai SV, Rajamanoharan PRS, Munday M, Heinrich, M. 2016. Plants used to treat diabetes in Sri Lankan Siddha Medicine - An ethnopharmacological review of historical and modern sources. Journal of Ethnopharmacology 198:531-599.

Sauer JD. 1979. Living fences in Costa Rican agriculture, Turrialba (IICA) 29:25-261.

Savaliram GG, Zanan RL. 2022. Ethnopharmacology and phytochemistry of kewda [*Pandanus odorifer* (Forssk.) Kuntze; Family: Pandanaceae]. In: Pullaiah T. (eds). Bioactives and pharmacology of medicinal plants. Apple Academic Press, USA, Pp. 13.

Saxena HO, Brahmam M. 1996 The Flora of Orissa. Vol. I-IV. Orissa Forest Development Corporation, Bhubaneswar, Odisha, India.

Shastri, JL. 1970. The Siva Purana. Motilal Banarasidass, New Delhi, India.

Shiva V, Jafri AH. 1998. Stronger than steel: Peoples movement against globalization and the Gopalpur steel plant. Research Foundation for Science, Technology and Ecology, New Delhi, India.

Simpson MG. 2019. Diversity and classification of flowering plants: Amborellales, Nymphaeales, Austrobaileyales, Magnoliids, Monocots, and Ceratophyllales. In: Simpson MG. (eds). Plant systematics. Academic Press, Cambridge, MA, USA, Pp. 187-284.

Solomon Raju AJ, Lakshminarayana G, Ch. Prasada Rao, Dileepu Kumar B, Santhi Kumari M, Prasad KBJ, Divyasree M, Suneetha Rani T. 2020. Pollination and fruit dispersal in the Fragrant Screw Pine, *Pandanus odorifer* (Forssk.) Kuntze (Pandanaceae). Species 21(67):113-119.

Staudt M, Bertin N. 1998. Light and temperature dependence of the emission of cyclic and acyclic monoterpenes from holm oak (*Quercus ilex* L.) leaves. Plant Cell Environment 21:385-395.

Stevenson PC, Nicolson SW, Wright GA. 2017. Plant secondary metabolites in nectar: impacts on pollinators and ecological functions. Functional Ecology 31:65-75.

Stone BC, Huynh KL, Poppendieck HH. 1998. Pandanaceae. In: Kubitzki K. (eds). The families and genera of flowering plants. Springer. Berlin, Pp. 397-404.

Stone BC. 1970. The flora of Guam. Micronesica 6(1):1-659.

Stone BC. 1982. *Pandanus tectorius* Parkins. in Australia: a conservative view. Botanical Journal of the Linnean Society 85:133-46.

Stone BC. 1990. New evidence for the reconciliation of floral organisation in Pandanaceae with normal angiosperm patterns. In: Baas P, Kalkman K, Geesink R. (eds.). Plant diversity in Malaysia. Springer, Dordrecht, Pp.33-55.

Stone BC.1968. Morphological studies in the Pandanaceae. I. Staminodia and pistillodia of *Pandanus* and their hypothetical significance. Phytomorphology 18(4):498-509.

Stone BC.1974 Toward an improved infra-generic classification in *Pandanus* (Pandanaceae). Botanische Jahrbucher fur Systematik 94:459-540.

Sulieman AME, Abdallah EM, Alanazi NA, Ed-Dra A, Jamal A, Idriss H, Alshammari AS, Shommo SAM. 2023. Spices as sustainable food preservatives: A comprehensive review of their antimicrobial potential. Pharmaceuticals 16(10):1451.

Sun-Yup Shim. 2019. *Pandanus fascicularis* Lam. extract inhibits pro-inflammatory cytokines production in LPS-stimulated RAW 264.7 cells. Preventive Nutrition and Food Science 24(3):344-347.

Suntar I. 2020. Importance of ethnopharmacological studies in drug discovery: role of medicinal plants. Phytochemistry Reviews 19:1199-1209.

Tallis H, Kareiva P, Marvier M, Chang A. 2008. An ecosystem services framework to support both practical conservation and economic development. Proceedings of the National Academy of Sciences of the United States of America 105(28):9457-9464.

Tan MA, Takayama H. 2019. Recent progress in the chemistry of *pandanus* Alkaloids. Alkaloids Chemistry and Biology 82:1-28.

Tanna MTH, Amin MN, Ibrahim M, Mukul MEH, Kabir A. 2016. Evaluation of antioxidants, membrane stabilizing, cytotoxic and anthelmintic activity with phytochemical screening of *Chromolaena odorata*: A medicinal shrub. International Journal of Pharmacy 6(1):53-61.

Tomlinson P, Zimmerman M, Simpson P. 1970. Dichotomous and pseudodichotomous branching of monocotyledonous trees. Phytomorphology 20(1):28-32.

Tongway D, Ludwig J. 2011. Restoring disturbed landscapes: Putting principles into practice. Island Press, Washington, DC, USA, Pp. 189.

Vahirua-Lechat I, Menut C, Roig B, Bessiere JM, Lamaty G. 1996. Isoprene related esters, significant components of *Pandanus tectorius*. Phytochemistry 43 (6):1277-1279

Vijge MJ, Biermann F, Kim RE, Bogers M, Driel MV, Montesano FS, Yunita A, Kanie N. 2020. Governance through global goals. In: Biermann F, Kim RE. (eds.). Architectures of earth system governance: Institutional complexity and structural transformation. Cambridge University Press, Cambridge, U. K., Pp. 254.

Vinod MS, Raghavan PS, George S, Parida A. 2007. Identification of a sex-specific SCAR marker in dioecious *Pandanus fascicularis* L. (Pandanaceae). Genome 50:834-839.

Vinod MS, Sankararamasubramanian HM, Priyanka R, Ganesan G, Parida A. 2010. Gene expression analysis of volatile-rich male flowers of dioecious *Pandanus fascicularis* using expressed sequence tags. Journal of Plant Physiology 167:914-919.

Warburg O. 1900. Pandanaceae. In: Engler A. (eds.). Pflanzenreich III. Verlag von Wihem Engelmann, Leipzig, Pp. 11-97.

Wardah F. 2009. Ethnobotanical study on the genus *Pandanus* L.f. in certain areas in Java, Indonesia. Biodiversitas 10(3):146-150.

Warrier PK, Nambiar VPK, Ramankutty, C. 1995. Indian medicinal plants. Vol. 4. Orient Blackswan, Hyderabad, India.

Warrier PK. 1997. Indian medicinal plants: a compendium of 500 species. 4th ed. Orient-Longman, Kottakkal, Hyderabad, India, Pp. 206-210.

Yuan H, Ma Q, Ye L, Piao G. 2016. The traditional medicine and modern medicine from natural products. Molecules 21(5):559.

Zdravchev NS, Alexey VFCh B, Romanov MS, Lebedev IM, Sorokin AN, Timchenko AS, Mikhaylova AA, Vasekha ND, Kandidov MV, Kuptsov KV, Iovlev PS. 2023. Phylogeny and historical biogeography of the order Pandanales. Geography, Environment, Sustainability 4(16):91-104.

Zhang X, Wu C, Wu H, Sheng L, Su Y, Zhang X, Xu X. 2013. Anti-hyperlipidemic effects and potential mechanisms of action of the caffeoylquinic acid-rich *Pandanus tectorius* fruit extract in hamsters fed a high fat-diet. PLoS One 8 (4):e61922